Comparative Biology and Epidemiology of Agroup and B-group *Leptosphaeria maculans* on Winter Oilseed Rape

Yongju Huang¹, Bruce Fitt¹, Xiaojia Hu¹, Avice Hall²

¹ Rothamsted Research, Harpenden, Hertfordshire, AL5 2JQ, UK; ² University of Hertfordshire, Hatfield, Hertfordshire, AL10 9AB, UK

INTRODUCTION

Stem canker (blackleg), caused by *Leptosphaeria maculans*, is a serious disease of oilseed rape in the UK, which caused losses of over £20 M per season since 1993 (DEFRA survey results; http://www.csl.gov.uk/prodserv/cons/crop/survey/osrintro.cfm). Populations of *L. maculans* can be divided into A-group and B-group (Williams & Fitt, 1999), which recently have been named as *Leptosphaeria maculans* (A-group) and *Leptosphaeria biglobosa* (B-group), on the basis of length of neck of pseudothecia (Shoemaker & Brun, 2001). Previous epidemiology work has mainly been on the A-group, with little information about the B-group. Controlled environment experiments have mainly used as inoculum conidia of *L. maculans*, rather than the ascospores, which play the major role in the epidemiology of the disease. The aim of this study was to compare the biology and epidemiology of A-group and B-group *L. maculans*.

MATERIALS AND METHODS

Suspensions of A-group or B-group ascospores were inoculated onto water agar or oilseed rape leaf surfaces. Germination of ascospores and penetration of leaf tissues were investigated using light microscopy (Huang, 2002). Oilseed rape debris naturally infected by A-group or B-group *L. maculans* was incubated at $5 - 20^{\circ}$ C under continuous wetness. Maturation of pseudothecia was observed weekly. Oilseed rape debris naturally infected by *L. maculans* was buried or exposed on the soil surface; the presence of mature ascospores on this debris was investigated at intervals of 2 months during a period of 1 year. The ability of ascospores to germinate after exposure to different temperatures under dry conditions was investigated. Lengths of ascospores and lengths of necks of pseudothecia produced on naturally infected debris were measured.

RESULTS AND DISCUSION

The differences between A-group and B-group *L. maculans* provide further evidence that the two groups are actually different species (Table 1). The results increase our knowledge about the biology and epidemiology of A-group and B-group *L. maculans*. This will help in developing strategies to control phoma stem canker more effectively. Germination patterns of ascospores of A-group and B-group *L. maculans* were

different (Table 1), whether ascospores germinated on water agar or leaf surfaces (Huang *et al.*, 2001). This could also be used to assess proportions of the two groups

| Experiment | Parameter | A-group | B-group |
|--------------|---------------|---|----------------------------------|
| Ascospore | Germ tube | Originated from interstitial | Originated from |
| germination | position | cells | terminal cells |
| on water | Germ tube | Shorter (40 µm in 12h at | Longer (131 µm in 12h |
| agar or leaf | length | 20°C) | at 20°C) |
| surfaces | Hyphal growth | Tortuous | In almost straight lines |
| | No. of germ | More per ascospore (3.8) | Less per ascospore (3.1) |
| | tubes | | |
| | Diameter of | Thicker (1.8 µm) | Thinner (1.2 µm) |
| | germ tubes | | |
| Penetration | Stomata | Mainly through stomata | Mainly through stomata |
| leaf tissues | Wounds | Through wounds | Through wounds |
| | Appressorium | Appressorium-like | No appressorium-like |
| | | structure formed | structure formed |
| Pseudothec. | Maturation | Mature at 5-20°C, earlier | Mature at 5-20°C |
| maturation | | than B-group at 5-10°C | |
| | Position of | Whole pseudothecia | Only neck exposed; |
| | pseudothecia | exposed on stem base | under epidermis of |
| | | surface | upper stem |
| Survival on | Survival on | Survived longer (1 year) | Did not survive 2 |
| buried or | buried debris | on buried stem bases | months on buried stem |
| unburied | | | bases or upper stems |
| debris | Survival on | Survived 1 year on | Survived 1 year on |
| | unburied | unburied stem base and | unburied stem base and |
| | debris | upper stem debris | upper stem debris |
| | Survival of | Survived longer (18-36% | Survived shorter (1.2- |
| | ascospores | survived 50 days at 5- | 19% survived 50 days at |
| | | 15°C; 10% survived 35 | 5-15°C; 17% survived |
| | | days at 20° C) at 5- 20° C in | 35 days at 20° C) at 5- |
| | | dry conditions | 20°C in dry conditions |
| Size of | Ascospore | Similar length range (28- | Similar length range |
| ascospores | length | 50 μm) to B-group | (28-44 µm) to A-group |
| | | | |
| Size of | Neck length | Generally shorter neck | Generally longer neck |
| pseudothe. | _ | (76.5 µm), but 50% had | (94.5 µm), but 50% had |
| | | same neck length range as | same neck length range |
| | | B-group ($< 81 \mu m$) | as A-group (< 81 µm) |

Table 1 Comparative biology and epidemiology of A-group and B-group *L. maculans*, as shown by results of these experiments

in populations in early autumn and to predict the severity of stem canker epidemics. The main mode of penetration of oilseed leaf tissue by hyphae from both A-group and B-group ascospores was through stomata, but the percentage of germinated ascospores that penetrated stomata was greater for A-group than for B-group *L. maculans*. Furthermore, appressorium-like structures were formed by A-group not B-group *L. maculans*. These results suggest that B-group may be less efficient than A-group *L. maculans* in infecting the host tissues under the same conditions. This may explain why the proportion of ascospores which established lesions for A-group *L. maculans* (12% infection efficiency) was higher than for B-group *L. maculans* (7% infection efficiency) after inoculation with ascospores in controlled conditions (Toscano-Underwood *et al.*, 2001).

The average length of A-group ascospores (38 μ m) was longer than that of B-group ascospores (36 μ m), but they had the same length range (28-50 μ m). A-group pseudothecia were mainly produced on stem bases and exposed to the air, while B-group pseudothecia were mainly produced on upper stems under the epidermis. However, the shape of B-group pseudothecia was similar to that of the A-group. Although the average length of pseudothecial neck of B-group *L. maculans* was longer than that of A-group *L. maculans*, 50% of A-group and B-group pseudothecia had neck lengths in the same range. This suggests that the two groups cannot be reliably differentiated by the length of ascospores or length of the neck of pseudothecia (Shoemaker & Brun, 2001).

A-group pseudothecia matured earlier than B-group pseudothecia at 5-10°C, but there were no differences between the two groups at 15-20°C. These different effects of temperature on maturation of A-group and B-group ascospores could explain why isolates from upper stems were mainly B-group, while isolates from stem bases were mainly A-group (West *et al.*, 2002). Results suggest that in the UK low temperatures in winter can delay maturation of B-group ascospores more than that of A-group ascospores. In spring, as temperatures increase, maturation of B-group ascospores may increase. Subsequently, infection of upper stems is mainly associated with B-group *L. maculans*. The effects of temperature on maturation of pseudothecia of A-group and B-group *L. maculans* could also be used to forecast the first release of ascospores and timing of phoma leaf spotting, and subsequently to optimise the use of fungicides.

A-group *L. maculans* survived longer than B-group *L. maculans* on buried stem base debris, but both A-group and B-group *L. maculans* survived longer on unburied debris than on buried debris. These results suggest that deep ploughing immediately after harvest may help to decrease the severity of stem canker in crops planted the next season in the surrounding area. A-group ascospores survived longer than B-group ascospores in darkness in dry conditions at temperatures of 5-20°C. These results suggest that more A-group ascospores than B-group ascospores can survive after travelling a long distance. This suggests that there is higher risk of spread to a new area through air-borne ascospores for A-group than B-group.

ACKOWLEDGEMENTS

We thank the Perry Foundation, UK Department of the Environment, Food and Rural Affairs, Biotechnology and Biological Sciences Research Council, DFID British Council and the China Scholarship Council for supporting the work. We thank M Jedryczka for supplying oilseed rape debris from Poland.

REFERENCES

Huang YJ, 2002. Comparative biology and epidemiology of A-group and B-group *Leptosphaeria maculans* on winter oilseed rape. *PhD thesis*, August **2002**, Rothamsted Research, UK.

Huang YJ, Toscano-Underwood C, Fitt BDL, Todd AD, West JS, Koopmann B, Balesdent MH, 2001. Effects of temperature on germination and hyphal growth from ascospores of A-group and B-group *Leptosphaeria maculans* (phoma stem canker of oilseed rape). *Annals of Applied Biology* **139**, 193-207.

Shoemaker R A, Brun H. 2001. The teleomorph of the weakly aggressive segregate of *Leptosphaeria maculans. Canadian Journal of Botany* **79**, 412-419.

Toscano-Underwood C, West JS, Fitt BDL, Todd AD, Jedryczka M, 2001. Development of phoma lesions on oilseed rape leaves inoculated with ascospores of A-group or B-group *Leptosphaeria maculans* (stem canker) at different temperatures and wetness durations. *Plant Pathology* **50**, 28-41.

West JS, Balesdent MH, Rouxel T, Narcy JP, Huang YJ, Roux J, Steed JM, Fitt BDL, Schmit J, 2002. Colonisation of winter oilseed rape tissues by A/Tox+ and B/Tox0 *Leptosphaeria maculans* (phoma stem canker) in France and England. *Plant Pathology* **51**, 311-321.

Williams RH, Fitt BDL, 1999. Differentiating A and B groups of *Leptosphaeria maculans*, causal agent of stem canker (blackleg) of oilseed rape. *Plant Pathology* **48**, 161-175.