

BREEDING AND SOME BIOLOGICAL PROPERTIES OF YELLOW
 SEEDED WINTER RAPESEED (*Brassica napus* L.)

M. B e c h y n ě , Department of Genetics and Plant
 Breeding, University of Agriculture, Prague 6 - Suchbát, Czechoslovakia.

The quality of rapeseed meal as an important source of protein is limited by its high content of crude fiber in the seedcoat. In studies on different *Brassica* species (BECHYNE et al. 1979, BELL and SHIRES 1982, LIU-HOU-LI et al. 1983, STRINGAM et al. 1974, SHIRZADEGAN and RÖBBELEN 1985) have shown that yellow seeded forms have got a considerably thinner seed coat than dark ones, contain a smaller proportion of hulls, resulting in improvements in oil, protein and fiber content. At present all the cultivars of rapeseed (*B. napus*) licensed in Europe are dark in seed color.

Interspecific hybridization within the genus *Brassica* has been well recognized as an important tool for transfer the yellow seed coat color, from *B. campestris* or some other species to *B. napus* (BECHYNE 1985, 1986). The transfer of seed coat color, seed testa of interspecific hybrids offers an interesting field of study. This is the object of our experiments in this paper which presents some results of yellow seeded *B. napus*. breeding in Czechoslovakia.

MATERIALS AND METHODS

Materials utilized in interspecific crossing for obtaining yellow seeded *B. napus* originated from *Brassica campestris* - var. Yellow Sarson, *B. oleracea* var. *acephala* (selected light greyish seed) and yellow seeded *B. carinata* (selection from a land race). Plants with crossed buds were kept in isolation chamber till the stigma was seen to be developing into a pod. Progenies were raised

in pots, greenhouse beds and advanced generations were field grown.

Four classes of color for the main analyses and six for microscopic studies were identifiable and the seed was classified as either yellow, ochre, brown and black in the first case and yellow (I.) ochre (II.) yellow brown (III.) light and dark brown (IV., V.) and black (VI.). For microscopic measurements of testa, seeds were dehydrated and saturated thoroughly directly in ethanol - n-butanol mixture (15 : 85) for 12 - 24 hrs. and then saturated with dehydrogenated pure n-butanol and slowly transferred into paraffin. The sections were stained in Ehrlich hematoxylin and enclosed into Solacryl resin.^{x)} Considering the expected sterility of some hybrids, colchicin treatment was made on seedlings at 2-4 leaf stage. 0,01 colchicine solution was applied by cotton pads to the vegetative tips, five times a day. The oil content of the whole seeds was measured by NMR spectroscopy. For further analyses and measurement conventional techniques were used.

RESULTS AND DISCUSSION

Interspecific hybrids between *B. campestris* and *B. carinata* (and reciprocals) set sufficient number of yellowish-grey hybrid seed. 162 crosses and reciprocal crosses formed 17 pods with 65 seeds (0,40 seed per pollination). The seeds developed into F_1 *B. carinata* like plants with vigorous growth and disproportionally long branches with sufficient number of pods mostly without seeds. A few pods contained some misshapen siled like structures, which did not germinate in favourable conditions on moist filter

x) Microbiologic studies were provided by J. Pazourek and staff, Faculty of Natural Science Charles University, Prague.

paper. The sterility barrier could not be overcome by chromosomal duplication. The results are not in agreement with MIZUSHIMA (1950), who reported 50 % fertility in allopolyploids of his *B. campestris* x *B. carinata* hybrids. But the pods and seed like structures indicate that a successful fertilization in some cases is affected, but the embryo development is inhibited at some later stage. The embryoculture could probably overcome the sterility. The sterility barrier was the reason for not taking further full advantage of *B. campestris* x *B. carinata* crosses. A total of 120 pollinations between *B. oleracea* and *B. campestris* was made. One cross combination only proved to be successful, producing 0,18 seed per pollination. After doubling the chromosome number only one plant proved to be tetraploid and survived up to maturity. It showed many signs of intermediate character. One of the hybrid plants produced in F_2 generation yellow seeds. After repeated selfing and selecting the best shade of yellow color was the spring type of yellow seeded *B. napus* crossed with Jet Neuf. The hybrid plants formed after continuous selfing and backcrossing a winter form of yellow seeded *B. napus*. Its seed however was not bright and deep in yellow color as in *B. campestris*, *carinata* or *junceae*. The properties of the best yellow seeded plants were evaluated in four years field trials and the characters of seed coat different in color, obtained within the breeding were compared (as shown in tables and figures).

Table 1: The overwintering of *B. napus* plants in %.

| | 1982 | 1983 | 1984 | 1985 | 1986 |
|---------------|------|------|------|------|------|
| Yellow seeded | 64.4 | 91.6 | 86.4 | 89.6 | 83.1 |
| Jet Neuf | 87.5 | 96.9 | 92.7 | 91.0 | 84.3 |
| Difference | 23.1 | 5.3 | 6.3 | 2.4 | 1.2 |

The overwintering of yellow seeded plants, improving in successive generation (except for 1984) was significantly lower in 1982 - 1984, compared with Jef Neuf (Table 1).

The microscopic measurements show an evident improvement in yellow and light colored seedcoats. Yellow seeds are the longest in radius (917,9 μm), while the black seeds reached 849,3 μm only. The thinner testa, mainly due to the palisade layer, explains the differences in biological and biochemical properties of the yellow seeded material. The two measured testa layers appear to be highly positively correlated (parabolic course). The black seeded strains differ evidently from the light ones.

The means of four years field experiments (tab. 2) show that the yellow and light seeded strains do not fully reach the properties of Jet Neuf. (No. of primary branches, No. of pods, per plant and oil content.) But the seed coat thickness and crude fiber content evidently elevate the former seeds. However it should be pointed out a slight variation in seed color shade in yellow seeds. Even within single pods brown or yellow-brown spots, mostly around hilum can be seen, similarly as indicated by HEYN (1973), JÖNSSON (1975) and HAWK (1982). XIAO found out in 1982 that the lighter the color of seedcoat, the thinner the seedcoat and the higher the oil content. His results support our intentions and necessity for further breeding of light yellow color, which could improve the economic properties of yellow seeded rape plants and overtake the present cultivars.

Table 2: Biological and economic characters of plants.

| Character | Seed coat color | | | |
|-----------------------------|-----------------|--------|--------|---------------------|
| | yellow | ochre | brown | black (Jet Neuf) |
| Height of plants (mm) | 126.4 | 128.1 | 126.3 | 132.6 |
| No. of primary branches | 5.5a | 5.7a | 5.7a | 6.2b |
| No. of pods per plant | 120.6a | 121.3a | 124.0a | 131.0b |
| No. of seeds per pod | 15.6a | 15.8a | 15.6a | 16.3a |
| 1000 seed weight (g) | 4.4a | 4.3a | 4.4a | 4.5a |
| % of seed coat | 11.6a | 12.1a | 17.0b | 19.7c |
| Oil content % | 44.8 | 44.9 | 45.3 | 45.4 |
| Sum oil + protein % | 67.8 | 67.1 | 65.1 | 65.3 |
| Crude fiber % | 10.2a | 10.6a | 12.7b | 13.1b |
| Yield (t ha ⁻¹) | 2.59b | 2.52b | 2.43a | 2.77c |

Means within rows followed by different letters are significantly different (PL 0.05)

SUMMARY

Winter forms of yellow seeded rapeseed (*B. napus* L.) were obtained as a cross of yellow seeded spring *B. napus* line and winter rapeseed cv. Jet Neuf. The yellow seeded spring rape originated from the interspecific crossing (*B. oleracea* and *B. campestris*). Four classes different in seed color for main analyses and six for microscopic studies were identified. The microscopic measurements of seed coat showed evident positive characters in yellow and light colored seeds (seed size, thinner testa layers). The differences in microscopic studies support the efforts for obtaining better seed properties (less fiber, more oil and protein and pleasing light appearance of meal), even if they do not reach fully the excellent parameters of Jet Neuf in our experiments. Further breeding and overcoming a slight variation in yellow seed color is necessary.

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Table 3: Statistical parameters of seedcoat characters in six different colors (I. - VI.)

| Testa color | Seed radius (μm) | | | Testa thickness (μm) | | | Palisade l. thickness (μm) | | | Thickness of aleurone l. (μm) | | |
|-------------|----------------------------------|--------|-------|---|-------|--------|---|-------|-------|---|-------|-------|
| | \bar{x} | s | V_k | \bar{x} | s | V_k | \bar{x} | s | V_k | \bar{x} | s | V_k |
| I | 917.90 | 17.534 | 1.91 | 36.80 | 4.770 | 12.96 | 14.66 | 1.728 | 11.79 | 22.14 | 3.856 | 17.42 |
| II | 882.00 | 19.241 | 2.18 | 43.66 | 6.091 | 13.95 | 17.58 | 1.798 | 10.23 | 26.08 | 4.590 | 17.60 |
| III | 913.95 | 19.271 | 2.11 | 39.63 | 4.295 | 10.84 | 16.78 | 1.174 | 6.99 | 22.85 | 3.431 | 15.02 |
| IV | 896.55 | 5.325 | 0.59 | 40.28 | 5.360 | 13.31 | 16.62 | 2.233 | 13.38 | 23.56 | 3.734 | 15.85 |
| V | 876.55 | 12.540 | 1.43 | 38.11 | 4.526 | 11.88 | 17.17 | 1.434 | 8.35 | 20.94 | 3.417 | 16.32 |
| VI | 849.30 | 7.390 | 0.87 | 60.32 ⁺⁺ | 5.796 | 9.61 | 35.86 ⁺⁺ | 3.199 | 8.92 | 24.46 | 4.091 | 16.73 |
| | Ratio palisade- aleurone ls. | | | Thickness of palisade l. in testa (%) | | | Thickness of testa in seed radius (%) | | | Thickness of palisade l. in seed radius (%) | | |
| | \bar{x} | s | V_k | \bar{x} | s | V_k | \bar{x} | s | V_k | \bar{x} | s | V_k |
| I | 0.677 | 0.117 | 17.35 | 40.089 | 4.177 | 10.491 | 4.011 | 0.541 | 13.48 | 1.596 | 0.180 | 11.29 |
| II | 0.682 | 0.071 | 10.41 | 40.512 | 2.607 | 6.436 | 4.949 | 0.673 | 13.60 | 1.995 | 0.201 | 10.06 |
| III | 0.748 | 0.091 | 12.24 | 42.561 | 2.895 | 6.802 | 4.339 | 0.504 | 11.61 | 1.836 | 0.141 | 7.69 |
| IV | 0.714 | 0.097 | 13.66 | 41.418 | 3.578 | 8.639 | 4.491 | 0.585 | 13.03 | 1.854 | 0.242 | 13.03 |
| V | 0.832 | 0.098 | 11.82 | 45.279 | 2.927 | 6.465 | 4.348 | 0.522 | 12.01 | 1.995 | 0.155 | 7.94 |
| VI | 1.493 ⁺⁺ | 0.231 | 15.45 | 59.58 [†] | 3.810 | 6.395 | 7.108 [†] | 0.702 | 9.88 | 4.223 [†] | 0.391 | 9.25 |

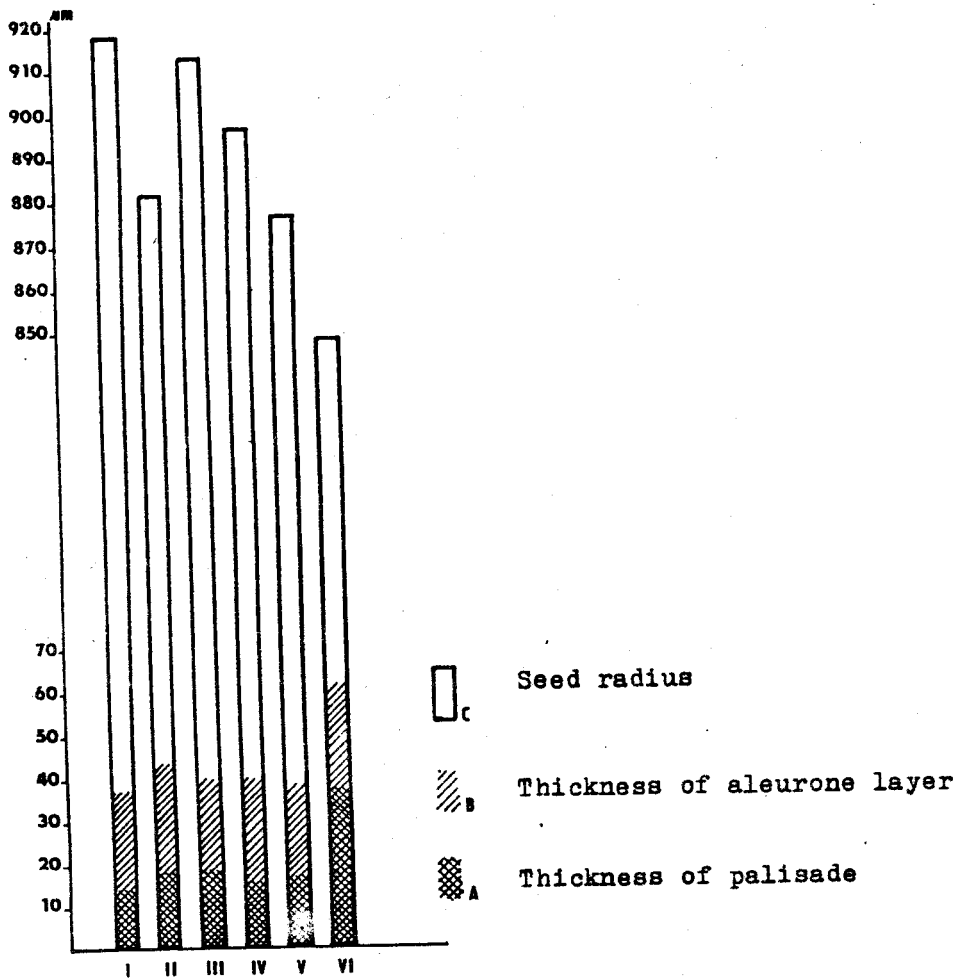
++ ... P=0.01

Table 4: Correlation index values - palisade and spongy layers

| Plant color | I | II | III | IV | V | VI |
|-------------|----------------------|----------------------|----------------------|----------------------|----------------------|----------------------|
| Plants 1 | 0.3538 ⁺⁺ | 0.2050 | 0.2842 | 0.1650 | 0.1778 | 0.5864 |
| 2 | 0.4460 ⁺⁺ | 0.1463 | 0.1654 | 0.3570 | 0.4578 ⁺⁺ | 0.5538 ⁺⁺ |
| 3 | 0.2727 | 0.5164 ⁺⁺ | 0.3250 ⁺⁺ | 0.4877 ⁺⁺ | 0.1628 | 0.4356 ⁺⁺ |
| 4 | 0.4107 ⁺⁺ | 0.6650 ⁺⁺ | 0.4380 ⁺⁺ | 0.1155 | 0.1140 | 0.2700 |
| 5 | 0.4969 ⁺⁺ | 0.5435 ⁺⁺ | 0.5011 ⁺⁺ | 0.5534 ⁺⁺ | 0.2822 | 0.2101 |
| 6 | 0.2057 | 0.5257 ⁺⁺ | 0.2703 | 0.1037 | 0.4160 ⁺⁺ | 0.5377 ⁺⁺ |
| 7 | 0.5524 ⁺⁺ | 0.2041 | 0.4949 ⁺⁺ | 0.2135 | 0.4029 ⁺⁺ | 0.1650 |
| 8 | 0.5608 ⁺⁺ | 0.1822 | 0.2979 ⁺ | -0.0098 | 0.2966 ⁺ | 0.3216 ⁺ |
| 9 | 0.1676 | 0.5803 ⁺⁺ | 0.2137 | 0.2579 | -0.1434 | 0.3323 |
| 10 | 0.1308 | 0.5100 ⁺⁺ | 0.1048 | 0.3541 | 0.2497 | 0.4092 ⁺ |
| 1-10 | 0.3172 ⁺⁺ | 0.4305 ⁺⁺ | 0.3077 ⁺⁺ | 0.3131 ⁺⁺ | 0.3120 ⁺⁺ | 0.2145 ⁺⁺ |

+ ... P=0.05, ++ ... P=0.01)

Fig. 1 Thickness of seedcoat layers and seed radius.



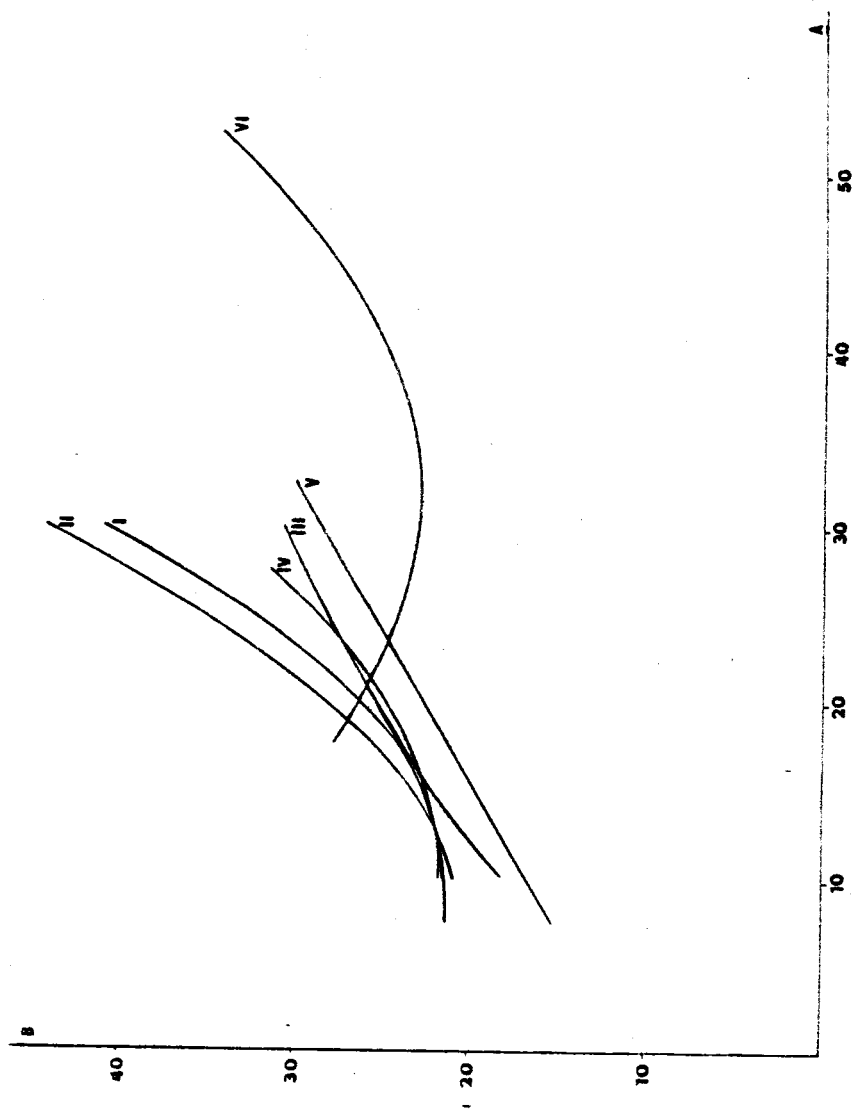


Fig. 2 Correlations between palisade (A) and aleurone layers (B) in six seed coats different in color (I. - VI.)