Quantity and quality effects in fatty acid segregation at winter oilseed rape hybrids (*Brassica napus* L.)

Jan Krzymanski, Marcin Matuszczak, Teresa Cegielska-Taras, Laura Szala, Irena Tokarczuk

Department of Oil Crops, Institute of Plant Breeding and Acclimatisation, Strzeszyńska 36, Poznań, 60-479, Poland Email:krzym@nico.ihar.poznan.pl

Abstract

Segregations of fatty acid composition in F₂ generations of rapeseed hybrids investigated till now had rather continuous character as typical for metric traits. Clearly defined zero erucic fraction occurring in relation 1:15 was found only at segregation in cross combinations "high erucic×zero erucic". (Harvey, Downey 1964, Kondra, Stefansson 1965, Krzymański at al. 1967, Krzymański, Downey 1968, Krzymański 1970, Jönsson 1977, Anand, Downey 1981, Chen, Heneen 1989, Chen, Beversdorf 1990, Scheffler at al. 1997, Barret at al. 1998, Fourmann at al. 1998, Lühs at al. 1998).

This continuous distribution observed for segregation of fatty acids may be a result:

- of control of these traits by polygenic genes working on cumulative manner or
- of masking the effects of main genes with small changes made by a number of modifying genes or
- of interference of main gene effects with effects of non heritable variability like: physiological variation, environment influences, chemical analyses errors, non proper sampling.

To explain the problem more exact study was conducted on segregation of fatty acids in hybrids of winter oilseed rape cultivated in Poland.

Key words: rapeseed, seed oil, fatty acids, biosynthesis, inheritance, segregation

Methods

The study was conducted on population of 291 doubled haploid lines (DH). These lines were obtained with the use of "in vitro" culture of microspores collected from F_1 hybrid plants (Cegielska-Taras at al. 1997, 2002). The hybrid was obtained in cross combination: "high erucic, low glucosinolate line"דzero erucic, high glucosinolate line". Parental lines differ greatly in origin and chemical composition. The population of DH lines was produced for constructing the genetic map of winter oilseed rape cultivated in Poland. It was expected that use of doubled haploid lines allow to obtain much better resolution than with inbreed lines.. DH lines are not only homozygous but also the segregation ratio should be more simple. Seeds from two self pollinated plant per line were collected and analysed. Chemical analyses of fatty acid composition in seed oil were made with gas liquid chromatography of their methyl ethers. Statistical analysis of segregation process were conducted using Microsoft Excel 97 and Statistica 5.1. programs.

Results and Discussion

Biosynthesis of fatty acids in ripening seeds of oilseed rape has following pathways (Yadav et al. 1993, Cassagne at al. 1994, Slabas at al. 1995, Lassner at. al. 1995,1996, Clemens at al. 1997, Millar, Kunst 1997) shown bellow:

where:

C 18:1 – denote acid with 18 carbon atoms in chain and with 1 double bond (oleic)

 $a = \Delta 9$ -desaturase of stearic acid

 $b = \Delta 12$ -desaturase of oleic acid

 $c = \Delta 15$ -desaturases of linolic acid

d = elongases of oleic and eicosenoic acids

Elongation of oleic acid chain is blocked in seeds of zero erucic rapeseed. Genetic source of this trait was found as natural mutant in Liho variety of spring rapeseed. (Stefansson at al. 1961,1964,).

Variability of fatty acids in studied population of 291 DH lines is shown in table 1.

Table 1. Fatty acid segregation in population of 291 DH lines of rapeseed (per cent of total fatty acid)

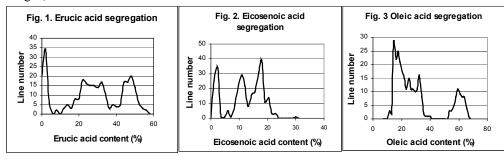
	Palmitic	Stearic	Oleic	Linolic	Linolenic	Eicosenoic	Erucic	
Mean	4,506	1,099	29,467	18,045	9,196	11,429	26,249	
Minimum	3,0	0,3	10,5	11,7	4,4	0,4	0,0	
Maximum	7,0	1,9	65,9	29,2	13,9	23,2	54,8	
C of V.	16,493	25,774	55,923	18,841	16,015	55,016	63,862	
S	0,226	0,133	1,386	0,833	0,552	1,226	0,854	
S_d	0,320	0,188	1,960	1,178	0,780	1,734	1,208	
LSD	0,640	0,375	3,920	2,356	1,560	3,468	2,415	

where: s = error of mean for line (two replication)

 s_d = error for difference between two line

LSD = least significant difference between means of two lines (P=0,05)

With the use of DH lines it was possible to isolate not only zero erucic fraction, but also to split the rest of population in two fractions: high erucic and intermediate. Intermediate fraction was irregular but it was difficult to state two subfractions connected with genes from *Brassica* rapa and *Brassica* oleracea. Frequency distribution of erucic, eicosenoic and oleic acids are shown on Fig. 1, 2 and 3.



These histograms were made with resolution approximately equal to standard deviation for line average. Histogram fractions contain many lines so differences in average values between them were very significant as it is shown in table 2.

Table 2. Least significant differences (LSD _{0,05}) for fatty acid contents by comparing fraction containing different number of lines (per cent of fatty acid)

Number of line in fraction —		Fatty acid	
Number of fine in fraction —	Oleic	Eicosenoic	Erucic
1	3,920	3,468	2,415
2	2,772	2,452	1,708
3	1,600	1,416	0,986
4	0,800	0,708	0,493
5	0,358	0,317	0,220

The ratios among the fractions set according frequency distribution of erucic, eicosenoic and oleic acid contents showed some deviations from theoretical segregation ratio 1:2:1. It was found:

zeroerucic fraction
 intermediate fraction
 high erucic fraction
 86 lines

Calculated χ^2 has value.4,649 with df=2 and probability of larger χ^2 is only P=0,098 what means that deviation from theoretical ratio is significant. This phenomenon can occur in DH lines populations. High erucic microspores (wild type) were probably preferred in haploidisation process.

Above analysis did not allow to split intermediate fraction into two subfractions connected probably with genes originated from Brassica oleracea (E_o) and from Brassica rapa (E_r). Four classes of segregates (00 00), (00 E_oE_o), (E_rE_r 00) and (E_rE_r E_oE_o) with ratio 1:1:1:1 can be expected in our population of DH lines. Method of multivariate grouping with 4 means were used to split DH lines in four groups according to contents of erucic, eicosenoic and oleic acids. Mean values of erucic, eicosenoic and oleic acids for these groups are shown in table 3 and their analyses of variance in table 4.

Table 3. Mean contents of fatty acids in calculated groups (%)

Acid	Group 1	Group 2	Group 3	Group 4
n=	60	69	73	89
Erucic	0,56	19,54	29,69	45,95
Eicosenoic	1,33	16,67	16,56	9,97
Oleic	59,51	29,74	21,42	15,61

Differences among groups according tree examined fatty acids are highly significant. Calculated χ^2 for these groups has

value.6,058.with df=3 and probability of larger χ^2 equal to P=0,109 what means that deviation from theoretical ratio are not significant on the level α =0,05. Obtained groups can be used for genes mapping (gen loci) not only for calculation of quantitave (metric) trait loci (QTL).

Table 4. An	alvses o	it variance i	tor tatty	acids in	orning

		•	•			
Acid:	SS	df	SS	df	F	p
Erucic	78110,35	3	3377,734	287	2212,299	0
Eicosenoic	10122,2	3	1342,988	287	721,0468	0
Oleic	75993,61	3	2757,898	287	2636,085	0

Conclusions

- 1) Due to higher resolution it was possible to separate not only zero erucic fraction but also to split the rest of DH line population in two clearly separated fractions: high erucic and intermediate.
- 2) Intermediate fraction did not have visible separation in two subfractions. This suggest that genes originated from *Brassica* oleracea and *B. rapa* have similar control in synthesis of erucic and eicosenoic acids..
- 3) The number of DH lines in fractions estimated according to contents of erucic and eicosenoic acids was as follow: zero erucic 60 lines, intermediate 145 lines, high erucic 86 lines. It shows some deviation from theoretical ratios 1:2:1. High erucic microspores were probably preferred in tissue culture.
- 4) It was not possible to split intermediate fraction in two subfraction using one factor analysis. This separation was done with the use the method of multivariate grouping with four means. Four groups were obtained according to contents of erucic, eicosenoic and oleic acids. These groups contain 60, 69, 73 and 89 lines. Segregation hypothesis 1:1:1:1 cannot be rejected by χ^2 test but some deviation on benefit to wild type is also visible.

References

Anand L.J., Downey R.K. - (1981) - A study of erucic acid alleles in digenomic rapeseed (Brassica napus L.) - Can. J. Plant Sci. 61: 199-203

Barret P., Delourme R., Foisset N., Renard M., Domergue F., Lessire R., Delseny M., Roscoe T. - (1998) - A rapeseed FAE1 gene is linked to the E1 locus associated with variation in the content of erucic acid - Theor. Appl. Genet. 96: 177-186

Cassagne C., Lessire R., Bessoule J.J., Moreau P., Creach A., Sscheider F., Sturbois F. - (1994) - Biosynthesis of very long fatty acids in higher plants -Prog. Lipid Res. 33: 55-69

Cegielska-Taras T. Szała L. - (1997) - Regeneracja roślin z mikrosporowych zarodków rzepaku ozimego (*Brassica* napus L.)(Plant regeneration from microspore derived embryos of winter oilseed rape (*Brassica* napus L.)) - Rośliny Oleiste - Oil Crops 18: 21-30.

Cegielska-Taras T., Tykarska T., Szała L., Kuraś L., Krzymański J. - (2002) - Direct plant development from microspore-derived embryos of winter oilseed rape Brassica napus L. ssp. oleifera (DC) Metzger - Euphytica 124/3: 341-347.

Chen J. L., Beversdorf W. D. - (1990) - A comparison of traditional and haploid-derived breeding populations of oilseed rape (*Brassica* napus L.) for fatty acid composition of the oil.- Euphytica 51: 59-65.

Chen B.Y., Heneen W.K. - (1989) - Fatty acids composition of resythesized *Brassica* napus L., *B. campestris* L. and B. alboglabra Bailey with special reference to the inheritance of erucic acid content - Heredity 63: 309-314.

Clemens S., Kunst L. - (1997) - Isolation of a *Brassica* napus cDNA (Accession No. AF009563) encoding 3-ketoacyl-CoA synthase, a condensing enzyme involved in the biosynthesis of very long chain fatty acids in seeds - Plant Physiol. 115: 313-314

Fourmann M., Berret P., Renard M., Pelletier G., Delourme R., Brunel D. - (1998) - The two genes homologous to Arabidopsis FAE1 co-segregate with the two loci governing erucic acid content in *Brassica* napus - Theor. Appl. Genet. 96: 852-858.

Harvey B. L., Downey R. K. - (1964) - The inheritance of erucic acid content in rapeseed (Brassica napus). - Can. J. Plant Sci. 44: 104-111.

Jönsson R. - 1977) - Erucic-acid heredity in rapeseed (Brassica napus L. and Brassica campestris L.) - Hereditas 86: 159-170

Kondra Z. P., Stefansson B. R. - (1965) - Inheritance of erucic and eicosenoic acid content of rapeseed oil Brassica napus). - Can. J. Genet. Cytol. 7:505-510.

Krzymański J. (1970) Genetyczne możliwości ulepszania składu chemicznego nasion rzepaku ozimego (Possibilities to improve chemical composition of winter rapeseed on genetic way) - Hodowla Roślin Aklim. Nas. 14/2: 95-133.

Krzymański J., Downey R.K. - (1969) - Inheritance of fatty acid composition in winter forms of rapeseed, Brassica napus. - Can. J. Plant Sci. 49: 313-319.

Krzymański J., Piotrowska A., Smólska B. - (1967) - Badania nad dziedziczeniem się zawartości kwasu erukowego u mieszańców rzepaku jarego (Bronowski×Liho). (Study on inheritance of erucic acid in spring rapeseed hybrids) - Biuletyn IHAR 6: 17-19.

Lassner M. W., Levering C. K., Davies H. M., Knutzon D. S. - (1995) - Lysophosphatidic acid acyltransferase from meadowfoam mediates insertion of erucic acid at the sn-2 position of triacylglycerol in transgenic rapeseed oil. - Plant Physiol, 109: 1389-1394

Lassner M. W., Lardizabal K., Metz J. G. - (1996) - A jojoba β-ketoacyl-CoA synthase cDNA complements the canola fatty acid elongation mutation in transgenic plants. - Plant Cell, 8: 281-292

Löhden I., Frentzen M. - (1992) - Triacylglycerol biosynthesis in developing seeds of Tropaeolum majus L. and Limnanthes douglasii R. Br. -Planta 188: 215-224

Lühs W., Friedt W. - (1995) - Breeding high-erucic acid rapeseed by means of Brassica napus resynthesis. Proc. 9th Intern. Rapeseed Congr., Cambridge, United Kingdom, 2: 449-451.

Millar A., Kunst L. - (1997) - Very long chain fatty acid biosynthesis is controlled through the expression and specifity of the condensing enzyme. - Plant J. 12: 121-131

Lühs W. W., Voss J., Han A., Gräfin zu Münster D., Weier F. P., Wolter F. P., Frentzen M., Friedt W. - (1998) - Genetic modification of erucic acid biosynthesis in *Brassica* napus. - Proc. XVth Eucarpia Congress, Viterbo, Italy, 20-25.9.1998 - pp.323-330.

Scheffler J. A., Sharpe A. G., Schmidt H., Sperling P., Parkin I. A. P., Lühs W., Lydiate D. J., Heinz E. - (1997) - Desaturase multigene families of *Brassica* napus arose though genome duplication. - Theor. Appl. Genet. 94: 583-591.

Slabas A. R., Simon J. W., Elborough K. M. - (1995) - Information needed to create new oil crops. - Inform 6/2: 159-166

Stefansson B. R., Hougen F. W. - (1964) - Selection of rape plants (*Brassica* napus) with seeds oil practically free from erucic acid. - Can. J. Plant Sci. 44:359-364.

Stefansson B. R., Hougen F. W., Downey R. K. - (1961) - Note on the isolation of rape plants with seeds oil free from erucic acid. - Can. J. Plant Sci. 41: 218-219
Yadav N. S., Wierzbicki A., Aegerter M., Caster C.S., Perez-Grau L., Kinney A. J., Hitz W. D., Booth J. R. Jr., Schweiger B. Stecca K. L., et al. - (1993) - Cloning of higher plant omega-3 fatty acid desaturases. - Plant Physiol. 103/2: 467-476