



Global Council for Innovation in Rapeseed and Canola

“Building a World community for Innovation on Rapeseed and Canola”

N° 7, May 2020

Contents

Table des matières

Editorial	2
Activity/ News of the association:	3
GCIRC Board meeting... on line	3
New GCIRC sponsor: NIAB	3
GCIRC Website: New design and new functions!	4
Collecting the 15th IRC presentations...	5
Value chains and regional news	5
Crops.....	5
Covid 19 Crisis.....	7
Quality of harvest 2019	7
France: 26e Carrefour de la sélection du colza / 26 th Seminar on Rapeseed Breeding. January 22-23, 2020.....	13
Scientific news	16
Publications:	16
Upcoming International and national events	32

Editorial

Rapeseed – a major global oil and protein crop: today, in the near future and beyond

*Since the invention of low-erucic rapeseed (0 type) and the subsequent, additional development of very low glucosinolate type (canola, 00) the cultivation of oilseed rape/canola has continuously increased on a global scale. Major growing areas of today are Canada, China, Australia and Western Europe (e.g. France, Germany, Poland, and the UK): Today, thirteen countries have a rapeseed acreage of more than 300,000 ha, led by Canada with an annual canola area of 6 to 9 million ha depending on the annual weather conditions. Further major producers are China and India, the latter mainly growing rapeseed mustard rather than *B. napus*.. Conspicuous advances in rapeseed cultivation and production have recently been achieved in countries of the near and middle east such as Kazakhstan, Romania, and Russia. In particular, the Russian Federation has a very large potential for rapeseed cultivation, especially in extensive areas of Western Siberia, where large amounts of rapeseed may be harvested in the future despite the current low yield potential. Other regions with high potential are Ukraine, USA, and Australia, despite the strong dependence on annual precipitation in major growing areas of these countries.*

Besides climate, other constraints can severely limit rapeseed growing and yield. Major challenges of today are numerous insect pests such as cabbage stem flea beetle, rape stem weevil, pollen beetle, cabbage seed weevil, and brassica pod midge which – depending on the site and year – can cause severe or even complete loss of the crop. In addition, fungal diseases such as blackleg, clubroot, sclerotinia and verticillium continue to damage rapeseed plants and fields. Whereas resistance breeding has led to some improvement by providing resistant or tolerant new cultivars, crop resistance against insects has not been successfully established and will probably do so for the years to come. Intensified future research activities in entomology and crop protection may help to overcome this unsatisfactory situation and improve the yield stability across environments i.e. locations and years.

In addition, the yield performance of rapeseed per se deserves substantial improvement. Intensifying research on yield physiology and broadening genetic diversity is seen as a helpful approach to higher yield of rapeseed varieties under challenging environments. Equally important, the international community in nutrition, both animal and human, will have to continue research for optimizing seed quality as a basis for optimal marketing and economical success of rapeseed production, not the least for the purpose of bioenergy creation and protect the environment.

This short description may point out the multitude of challenges rapeseed research, cultivation and use will be confronted in the near future. GCIRC as a diverse international community for research on rapeseed will be prepared to address these challenges.

Prof Wolfgang Friedt, GCIRC President

Activity/ News of the association:

GCIRC Board meeting... online

Fortunately, the International Rapeseed Congress and GCIRC General Assembly took place in 2019... Most international events and meetings have been cancelled or postponed this year due to the Covid 19 crisis. Nevertheless, the GCIRC board meeting scheduled on March 18 in Paris has been maintained through a video conference, just when Europe was closing everything. With good will of everybody (it was very late in Australia and China, and very soon in Canada), this experience was a success and proved, if needed, that international collaboration may profit a lot from the new communication technologies...

A part of the meeting has been devoted to the debriefing of the IRC15 experience, and several important issues have been treated, after the orientations given by the General Assembly in Berlin. Among these issues, 3 are especially important for the coming year:

- The next GCIRC Technical meeting is scheduled in Poznan, Poland, on May 17-20, 2021, hosted by the IHAR. Please keep the dates.
- Two major topics will be in the foreground: insects pests management strategies, and rape-seed/canola proteins.
- The practical refoundation of GCIRC scientific committees (Genetics and Breeding, Crop Protection, Agronomy and Crop Management, Quality and Products, Economy and Markets), in the perspective of sharing ideas and visions during the Technical Meeting.

Later on-line meetings have permitted to put the basis of an "Innovation and networking" group, and to pass detailed information on the Congress organization experience to the Australian team that will lead the IRC16, in 2023 in Sydney.

Further details will be given soon.

New GCIRC sponsor: NIAB

UK participated actively in the foundation of GCIRC in the 1970'ies, but its participation has been weakening in the 2000' ds. Fortunately, Simon Kightley convinced NIAB to take over, coming back to the period when UK organized a very successful IRC in 1995 in Cambridge, under the presidency of John McLeod.

NIAB was founded in 1919 as the National Institute of Agricultural Botany, to promote the improvement of British crops and to assess the merits of varieties and seed quality. It is now a major international research organisation in plant science, crop evaluation and agronomy. See: <https://www.niab.com/>

After the support in 2019 of SPZO, Union of Oilseed growers and Processors of Czech Republic, and AOF Australian Oilseed Federation, our sponsors are now 7, showing the interest of national professional sectors and research and innovation institutions in the activities of GCIRC. Thanks to the efforts of the GCIRC board members. To better know them, you may pay a visit to their websites.



GCIRC Website: New design and new functions!

The GCIRC launched its new website mid-March. Through this investment, the GCIRC wishes to go ensuring its traditional services of its website, to improve easiness and add new functionalities. The target is to favor easier interactions between the users of the website - GCIRC members... or not yet members- and GCIRC, and also between the members themselves. This new website will offer work-spaces for the GCIRC permanent thematic committees and for potential new thematic working groups. It also allows to directly modify one's personal page and to pay membership online with a credit card secured system.

We also considered this moment as a good opportunity to open a LinkedIn page for the GCIRC.

Our webmaster Laetitia Devedeux prepared of notice to make the best use of the main novelties and give some tips and tricks. You will find it as a top new at <https://www.gcirc.org/news-events/news/article/presentation-of-our-new-website>

We only report here the novelties:

NOVELTIES:

Member's profile

Now possible online modifications of your profile information: upload new cv, new photo, identification of your centers of interest, activity...

We advise GCIRC members to check and correct their profile and pay a special attention in expressing their fields of interest (several choices possible): it will favor easier interactions within the GCIRC community, notably to look for projects partners or organize working groups.

The information that appear on one's profile will be open to all, members, and non-members, except for the detailed CV that will be available only for members.

Online payment by credit card.

New members

Now possible online registration: cv, photo, interests, activity...

Online payment by credit card.

Workgroups/Collaborative activities

New collaborative spaces available for our members to work on specific subjects.

Search box

Improved research engine that searches in all site contents.

Our Newsletter

You can register your e-mail address to receive our Newsletter.

You can also read our Newsletter on-line or download the pdf file.

Our sponsors

Sponsors' logos carrousel on our Home page.

REMINDERS:

Publications

In the Archives, open to all, you will find former GCIRC Bulletins and IRC Proceedings until 2015.

GCIRC members have access to the latest IRC proceedings, Seminars/Symposia proceedings and Students thesis/reports and General Assemblies reports.

Board members can read all our Board Meetings reports.

Online directory

Members' directory & Institutions and Companies' directory

You can search Members, Institutions and Companies by:

Name, Countries, Main fields of interest, Main activity, or Type of Institution/Company.

The lists of all GCIRC members and many Institutions and Companies are also available here.

Photo library

More pictures now available. Do not hesitate to share yours so we can add them.

... Last but not least: we also need some pictures to illustrate rapeseed/canola in the different countries of the world, notably for the top parts of the website pages: this is a call for volunteers.

Collecting the 15th IRC presentations...

To the attention of the participants to the 15th IRC in Berlin, it is not too late to send us your presentations or posters as pdf files. They will be published on the new GCIRC website, coming soon.

Value chains and regional news

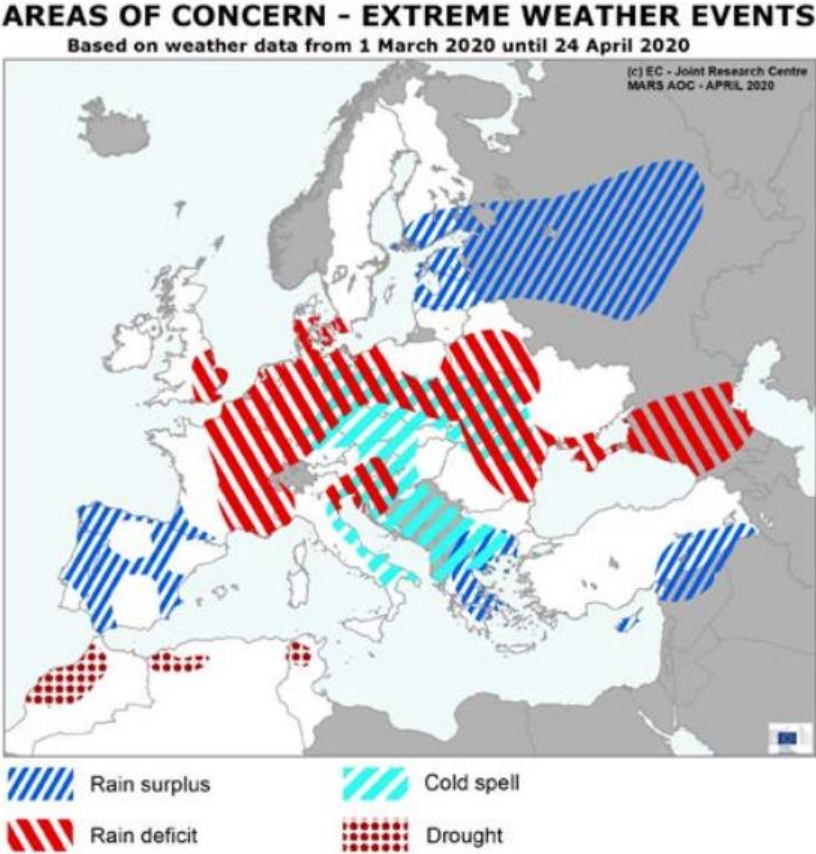
Crops

The last report of Statistics **Canada** on principal field crop areas (Issued May 7; see <https://www150.statcan.gc.ca/n1/daily-quotidien/200507/dq200507a-eng.htm>) mentions that "At the national level, farmers expect to seed fewer acres of canola in 2020 compared with one year

earlier, decreasing 1.6% to 20.6 million acres in 2020 (8,33 Million ha). If realized, this would be the second consecutive year of decrease in canola seeded area at the national level and the lowest seeded area since 2013 as farmers shift away from oilseeds to other crops.”. Acreage would be down 2.3% from 2019 in Saskatchewan, 2.8% in Alberta and 3% in Manitoba.

In Europe, The European Commission estimates the rapeseed acreage to 5,3 million ha, up 2,7% compared to 2018/19, but still 10% below the five-year acreage: as in 2018, the unfavorable weather conditions impeded rapeseed sowings in the autumn of 2019, especially in France, where some land had to be ploughed up.

According to the European Commission winter crops are still in good conditions despite lack of rain: Western Europe experienced one of the driest starts to spring since 1979 - after a very wet winter - with almost no rain since mid-March. (see details in <https://ec.europa.eu/jrc/sites/jrcsh/files/jrc-mars-bulletin-vol28-no4.pdf>),



These elements will impact international trade: rapeseed availability is expected to be very low next year in Europe. A significant import flow will certainly be necessary, first from Ukrainian rapeseed and then Canadian canola. In Ukraine, a drop in production is also expected and therefore a decrease in export capacities. Concerning Canada, exports to China restart, after being almost stopped since March 2019, more than 370,000 tons of canola were sent in March 2020. The sustained recovery in Chinese demand would be a positive factor for prices in the next season.

Covid 19 Crisis

Most sources of information report that concerning the agricultural sector, the main sources of uncertainties due to the coronavirus pandemic were logistics, and for some sectors (fruits and vegetable, labor availability). According to the EU Commission MARS Bulletin “While labour availability remains a key concern, to date we have no evidence of major COVID-19 driven impacts on the spring and summer crop sowings. The pandemic has also brought uncertainty and concerns about inputs (e.g. seeds, fertilizer, agro-chemicals) related to disruptions in logistics. So far, the supply of seed, fertilizer and pesticides seems to be adequate and no immediate disruptions are expected.”

According to the EU Short term outlook (https://ec.europa.eu/info/sites/info/files/food-farming-fisheries/farming/documents/short-term-outlook-spring-2020_en.pdf) “in the grains and oilseeds market, ample global stocks, sizeable 2019/2020 harvest in the EU and neighboring countries, and good prospects in the Americas for the next crop should allow a satisfactory supply of the market in the coming months” except perhaps in case that key exporters put barriers to exports, like recently Russia did for cereals.

The main disturbances on the European oilseeds market is linked to the drop of petroleum prices which directly impacted vegetable oils prices, and the drop of biodiesel demand which is a key outlet for rapeseed and impacts the supply of local proteins.

Quality of harvest 2019

NB: note that the results presented in the two studies in Canada and France are not expressed on the same basis of calculation.

Quality of western Canadian Canola - 2019

Weather and production review

In the prairies, seeding started early May and progressed steadily. By the end of May and the first week of June, it was completed in the three provinces. Cooler temperatures in May and June, associated with a lack of moisture, led to a much-delayed canola seed germination. Finally, germination was patchy in many fields, leading to uneven fields.

Cooler than normal temperatures (average 2 to 3°C lower) persisted during summer for most of the three provinces. In general, crops were one two weeks behind for most of the growing season due to the delayed germination in the three provinces. In Manitoba, during June, days were quite warm with temperatures reaching over 30°C; however, nights were quite cool with overnight lows in the single digit range. After the spring drought, above normal precipitation was the norm for the summer months in the prairies. However, this precipitation was uneven. During most of the growing season, the reported main causes of crop damages were either localized flooding due too much moisture or lack of moisture and high winds and hail.

The 2019 canola harvest was one to two weeks behind when compared to previous years. In September, frequent rainfalls and cooler than normal temperatures delayed harvest greatly. By mid-September, barely 10% of the canola crop was harvested compared to over 40% at the same time last year. The first snowfall occurred in Alberta and part of Saskatchewan the last week of September, it was

followed by a killer frost couples of days later. Mid-October, there was a significant snowfall in Manitoba, stopping all harvest activities. By the end of October, in Manitoba, canola harvest was considered over, with about 90% of the canola crop harvested. By mid-November, Saskatchewan canola harvest was considered finished with 91% of the canola crop harvested. About 19% of the crop in the east-central region of the province was still in the fields. By the first week of December, the Alberta canola harvest was considered complete with only 84.5% harvested. A large percentage of the crops was still in the fields, including 34% and 20% of the seeded canola respectively in the Peace River and the North-East areas of the province.

Production

As of January 2020, Statistics Canada reported that the estimated 2019 Canadian production was 18.649 million metric tonnes (MT), about 1.7 MT less than last year production and about 2.5 MT less than the record production observed in 2017 (21.328 MT), but similar to the 5-year average production (18.682 MT).

Table 1 - Seeded area and production for western Canadian canola

	Seeded area			Harvested area			Production ¹		
	thousand hectares			thousand hectares			thousand tonnes		
	2019	2018	2014-18	2019	2018	2014-18	2019	2018	2014-18
Manitoba	1,338.6	1,382.4	1,308.4	1,298.5	1,367.5	1,292.9	3,056.3	3,318.4	2,888.5
Saskatchewan	4,674.1	4,997.9	4,713.0	4,604.2	4,955.0	4,682.1	10,130.5	10,927.1	10,059.8
Alberta	2,401.2	2,755.9	2668.6	2,355.6	2,703.0	2,634.4	5,320.1	5,870.6	6,100.6
British Columbia	34.7	55.4	53.2	30.9	54.3	41.9	72.0	123.9	87.0
Canada	8,480.6	9,232.2	8,628.1	8,319.2	9,119.7	8,534.9	18,648.8	20,342.5	18,682.1

¹ For all production data please consult Statistics Canada. [Table 32-10-0359-01 Estimated areas, yield, production, average farm price and total farm value of principal field crops, in metric and imperial units at: https://www150.statcan.gc.ca/t1/tbl1/en/cv.action?pid=3210035901](https://www150.statcan.gc.ca/t1/tbl1/en/cv.action?pid=3210035901)
DOI: <https://doi.org/10.25318/3210035901-eng>

Quality of the 2019 canola harvest

This report of quality data for the 2019 harvest is based on analyses of 2,320 individual canola samples. Composites of Canola, No.1 Canada from each crop district from each province were made using these samples. Specialty oil samples, such as high oleic acid, low linolenic acid, and high erucic acid, were excluded from this report. Crop district composites of Canola, No.1 Canada samples were prepared using 1,936 samples. All quality parameters are reported on 8.5% moisture basis, except chlorophyll which is as this.

In 2019, only 85.1% of the samples were graded Canola, No. 1 Canada, compared to 74.4% in 2018 and 87.3% for the 5-year average. Since 2009, this is the 4th lowest percent of samples graded Canola, No.1 Canada, the worst average was obtained last year (74.4%) followed by the 2011 average (75.3%) and

the 2012 average (82.0%). The grade distribution of the 2019 canola crop varied greatly between provinces and between crop districts within a province. The main damages were distinctly green seed count (DGR) and admixture. Overall, distinctly green seeds (DGR) were 0.39% (0.36% in 2018) in Canola, No. 1 Canada, 3.13% (4.04% in 2018) in Canola, No. 2 Canada, 10.23% (12.01% in 2018) in Canola, No. 3 Canada and 8.52% (32.95% in 2018) in Sample.

The 2019 western Canadian canola (Canola, No.1 Canada) crop was characterized by an oil content average higher than what was observed for the 2018 crop (44.6% in 2019 versus 44.1% in 2018), and a lower protein content average (20.4% in 2019 versus 21.1% in 2018) (Table 2). Samples showing the highest oil content average were from Saskatchewan, while for the protein content, Manitoba showed a higher average than Alberta or Saskatchewan (Table 2).

The chlorophyll content average for Canola, No.1 Canada samples was higher in 2019 than in 2018 (12 versus 10 mg/kg, respectively). Chlorophyll content average (Table 2) was significantly higher in Alberta-BC Peace River (17 mg/kg) than Saskatchewan (11 mg/kg) and Manitoba (5 mg/kg). Some areas in Alberta-Peace River showed chlorophyll content averages higher than 20 mg/kg.

Table 2: 2019 Canadian canola harvest: Canola quality data by grade and province – oil, protein, chlorophyll and glucosinolate contents.

		Oil content ¹			Protein content ²			Chlorophyll content ⁵			Glucosinolates ¹		
		(%)			(%)			(mg/kg)			($\mu\text{mol/g}$)		
	Number of samples	mean	min.	max.	mean	min.	max.	mean	min.	max.	mean	min.	max.
Canola, No. 1 Canada													
Manitoba	448	43.9	38.3	50.8	21.5	15.7	26.8	5	3	53	9	3	15
Saskatchewan	839	45.1	36.0	51.0	20.0	14.4	27.1	11	3	52	9	4	19
Alberta-Peace River ³	649	44.2	37.0	51.9	20.5	14.7	27.8	17	3	56	10	4	18
Western Canada⁴	1936	44.6	36.0	51.9	20.4	14.4	27.8	12	3	56	9	3	19
Canola, No. 2 Canada													
Manitoba	13	42.7	38.8	48.0	22.0	18.0	24.7	9	4	33	9	4	14
Saskatchewan	63	44.6	39.4	49.2	20.5	15.8	25.6	36	5	91	10	5	15
Alberta-Peace River ³	114	44.4	38.5	50.6	20.1	16.0	26.8	44	5	87	11	6	17
Western Canada⁴	190	44.4	38.5	50.6	20.4	15.8	26.8	43	5	91	10	4	17
Canola, No. 3 Canada													
Manitoba	0												
Saskatchewan	31	44.5	38.9	49.2	20.7	16.0	27.1	69	4	165	11	8	16
Alberta-Peace River ³	75	44.9	38.1	50.0	20.4	15.7	24.5	79	7	138	11	5	19
Western Canada⁴	106	44.7	38.1	50.0	20.5	15.7	24.5	76	4	165	11	5	19
Canola, Sample Canada													
Manitoba	1	45.0			21.0			6					
Saskatchewan	23	44.0	38.5	48.0	19.9	17.0	23.7	45	4	157	8		
Alberta-Peace River ³	18	43.2	35.4	48.0	20.3	17.2	24.6	97	5	204	11	7	17
Western Canada⁴	42	43.7	35.4	48.0	20.1	17.0	24.6	63	4	204	12	8	23

¹ 8.5% moisture basis

² N x 6.25; 8.5% moisture basis

³ Includes part of the Peace River area that is in British Columbia

⁴ Values are weighted averages based on production by province as estimated by Statistics Canada

⁵ Individual canola samples are analyzed by Near-Infrared Spectroscopy, the accurate limit of quantification for chlorophyll is 4 mg/kg

Total seed glucosinolate averages were very similar to last year (9 $\mu\text{mol/g}$ in 2019 versus 10 $\mu\text{mol/g}$ in 2018 and for the 5-year average). There was no difference between provinces regarding the average of the total glucosinolate contents (Table 2).

The oleic acid content average of the 2019 canola crop was much lower than what was observed in 2018 (62.43% versus 64.3%) or for the 5 year-average (63.1%). The highest oleic acid content average was observed in Manitoba, whereas the lowest average was observed in Alberta (Table 3). This

decrease in oleic acid content was accompanied by a sharp increase in α -linolenic acid content (10.0% in 2019 versus 8.7% in 2018), whereas linoleic acid content increased slightly (18.7% in 2019 and 18.3% in 2018) when compared to the 2018 crop. The highest α -linolenic acid content average was observed in Alberta-BC Peace River and the lowest in Manitoba. The 2019 fatty acid composition resulted in a very different iodine value in 2019 when compared to the 2018 canola crop, 113.7 units in 2019 versus 111.0 units for 2018.

Total saturated fatty acid content for the 2019 canola crop was like what was observed in 2018 (6.6% versus 6.7%), with growing conditions having little effect on total saturates content in Canada.

The mean free fatty acids (FFA) average levels in 2019 Canola (0.16%), No.1 Canada seed was like what was observed in 2018 (0.15%) (Table 3). Individual samples showed a range of FFA level as harvest was done in wet conditions for the most part of the prairies.

Table 3 – 2019 Canadian canola harvest: Canola quality data mean by grade and province – free fatty acid content and fatty acid composition of the oil

	Relative fatty acid composition of the oil (%)						Iodine value ⁴ (Units)	Free fatty acids (%)
	C18:0	C18:1	C18:2	C18:3	C22:1	Total saturates ³		
<u>Canola, No. 1 Canada</u>								
Manitoba	1.79	63.25	18.66	9.20	0.00	6.75	112.1	0.24
Saskatchewan	1.75	62.88	18.37	9.98	0.00	6.61	113.4	0.13
Alberta-Peace River ¹	1.64	61.36	19.12	10.63	0.00	6.59	115.1	0.17
Western Canada²	1.73	62.40	18.74	10.01	0.00	6.64	113.7	0.16
<u>Canola, No. 2 Canada</u>								
Manitoba	1.70	61.71	19.45	9.83	0.00	6.80	113.8	0.76
Saskatchewan	1.65	60.96	19.37	10.74	0.00	6.59	115.5	0.23
Alberta-Peace River ¹	1.57	59.99	19.70	11.24	0.00	6.65	116.6	0.22
Western Canada²	1.61	60.52	19.54	10.94	0.00	6.63	116.0	0.25
<u>Canola, No. 3 Canada</u>								
Manitoba								
Saskatchewan	1.61	59.94	19.86	10.99	0.00	6.69	116.2	0.30
Alberta-Peace River ¹	1.54	59.86	19.78	11.27	0.02	6.60	116.8	0.25
Western Canada²	1.57	59.89	19.81	11.16	0.01	6.64	116.5	0.27
<u>Canola, Sample Canada</u>								
Manitoba	1.69	63.70	18.80	8.46	0.00	6.73	110.8	0.46
Saskatchewan	1.68	60.20	19.78	10.80	0.09	6.73	115.9	0.33
Alberta-Peace River ¹	1.51	58.12	20.39	11.94	0.20	6.61	118.3	0.72
Western Canada²	1.62	59.57	19.98	11.16	0.13	6.69	116.6	0.47

¹ Includes part of the Peace River area that is in British Columbia

² Values are weighted averages based on production by province as estimated by Statistics Canada

³ Total saturated fatty acids are the sum of palmitic (C16:0), stearic (C18:0), arachidic (C20:0), behenic (C22:0), and lignoceric (C24:0)

⁴ Calculated from fatty acid composition

Quality of winter rapeseed in France harvest 2019

Each year in France, the Observatory on the quality of winter rapeseed in France allows to assess the main qualitative criteria of the harvest. Most samples are taken just after harvest, when farmers

deliver them to the collecting body, cooperative or merchant. The harvest deals with winter rapeseed as the production of spring rapeseed is very limited in France.

In France, the 2018/19 rapeseed campaign was marked by very dry conditions late summer and autumn until the end of October in most regions, leading to a drop in sown acreages - 40 to 60% in some regions - and late emergence in part of the fields. The mild autumn and winter period allowed growth to catch up and a partial recovery of the crop. Spring was temperate and dry, drought penalized in the crops especially in shallow soils sometimes beginning in flowering times. The campaign was also marked by pest attacks, starting in autumn in the East and in spring in most regions (pollen beetle), with in the background an increasing resistance to insecticides. Overall, rapeseed was found to be very heterogeneous depending on the conditions of emergence and the water reserves of the soils in spring; yields were very variable.

Average quality characteristics Rapeseed in France, 2019 harvest					
	Nber of samples	surface weighted average	Minimum	Maximum	standard deviation on average
Impurities%	387	1,4	0,0	6,8	1,0
Water content %	519	6,2	3,6	10,0	0,8
Oil % on commercial standard basis*	516	43,0	36,8	48,2	1,3
Proteins % of DM	296	20,2	14,9	26	1,4
Proteins % of deoiled DM	296	39,2	31,9	48	2,0
Glucosinolates $\mu\text{mol/g}$ seeds at 9% humidity	108	15,7	9,8	20,8	2,4
<i>DM: Dry Matter</i>					
<i>* commercial standard for oil content: 40% oil on seeds at 9% humidity + 2 impurities</i>					

The average impurity rate of the 2019 harvest was at the same level as in 2018: 1.4%, with high variability between samples, especially in the South. Again, this year, the water content was well below the marketing standard (9%) with an average value of 6.2%. This value is the lowest recorded since the start of the seed quality survey (1991).

The average oil content was 43%, on commercial standards basis (9.0% moisture), lower than the performance in 2018 and slightly below the five-year average (43.6%). However, in view of the extreme conditions of the 2018/19 campaign, the observed oil content remained satisfactory. Especially since the proportion of samples whose oil content met the marketing standard was significant for all production areas and was better than in 2018.

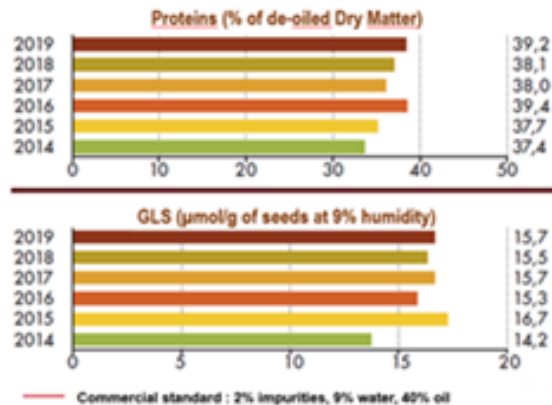
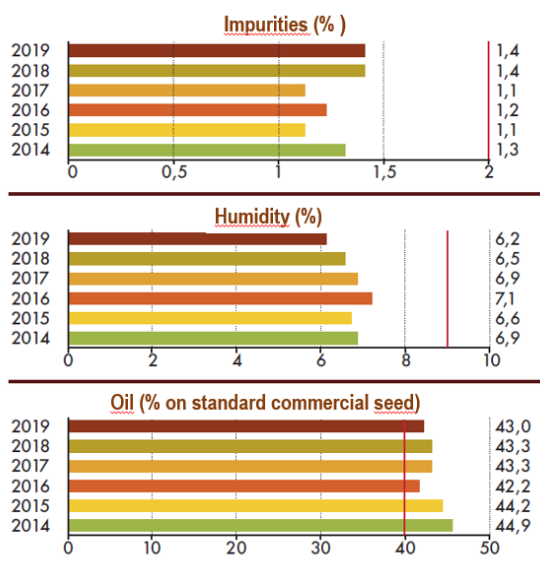
Opposite to oil content, the protein content average, 39.2% (expressed on dry and de-oiled seed), corresponded to the 2nd highest value since 2007. The low variability of the protein contents observed this year resulted from relatively homogeneous climatic conditions at the national level. The noticeable water stress at the time of seed filling led to a concentration of the protein in the seeds at the expense of the oil. It also resulted in a relatively low weight of a thousand grains, which partly explained the low national yields observed this year: 3,1 ton / ha on average according to Agreste (November 2019).

In 2019, the average value for the glucosinolate content was 15.7 $\mu\text{mol} / \text{g}$ at 9% moisture; the highest levels were found in the Atlantic coast and the Center-West, which suffered the heaviest stress (weak growth in autumn, strong pressure from pests).

Linolenic acid content (% of total fatty acids)														
Region	2006	2007	2008	2009	2010	2011	2012	2013	2014	2015	2016	2017	2018	2019
West Atlantic	8,6	10,0	10,8	9,5	9,7	8,5	9,8	9,4	10,0	9,7	9,1	7,2	9,4	10,1
Center West	7,8	9,8	10,9	9,5	9,4	9,1	9,9	9,4	8,7	8,7	9,5	8,8	8,6	9,7
East	7,7	9,2	10,1	9,1	9,1	9,6	10,2	9,9*	8,3	8,8	9,6	7,2	8,1	8,4

Panel from a multi-year follow-up of 3 variety trials representing the 3 main production basins (West Atlantic, Center-West and East).
* Average composed only of control varieties

The average linolenic acid (omega-3) content was 9.4% of the total fatty acids out of 105 controlled samples. It is well above the five-year average (8.8%). A significant decrease of more than one point was observed between the Atlantic coast and the East. The Atlantic coast benefited from early flowering, with temperatures, especially at night, relatively low, which favored the accumulation of linolenic acid. Overall, the accumulation of low temperatures, even frost temperatures, observed in the post-flowering period (between flowering and flowering + 60 days), contributed to the improvement of omega-3 contents in 2019.



France: 26e Carrefour de la sélection du colza / 26th Seminar on Rapeseed Breeding. January 22-23, 2020

Since 1977, the French association Promosol has been supporting works on oilseed crops breeding (rapeseed and sunflower) and promoting the dissemination of scientific progress. Promosol members are institutions: Terres Univia (Interprofessional Organization of the French Oil and protein crops sector), Terres Inovia (Technical Institute of the French Oil and protein crops sector), INRAE (National Institute for Agriculture, Food and Environment Research), and UFS (French seed association for seed companies & plant breeders). Promosol defines research priorities of common interest, funds key research programs and disseminates results to breeders and farmers. The association organizes the Carrefour de la Sélection du colza every year in January.

The 26th “Carrefour de la Sélection du Colza”, took place January 22nd and 23rd, 2020 in Orléans (France, Loiret). It brought together 70 participants from public research and higher education sector (INRAE, Agrocampus Ouest, Universities), GEVES (French Variety and Seed Study and Control Group), the industry and private rapeseed breeders.

The Carrefour was co-chaired by Martine Leflon, head of genetics and crop protection department at Terres Inovia, and lead of the Promosol rapeseed commission, and Maria Manzanares-Dauleux, director of the UMR IGEPP (INRAE-AgroCampus Ouest- University of Rennes I) and chair of the Promosol scientific council.

Exchanges on rapeseed genetics and breeding

The “Carrefour de la Sélection du Colza” is mainly an opportunity to take note of the progress of four research projects funded by Promosol (see below) and more widely to discuss the most important challenges of rapeseed breeding.

Experts from Terres Inovia were present: Arnaud Van Boxsom, head of varietal evaluation, presented the work in progress on the evaluation of the rapeseed vigor during autumn; Etienne Pilorgé, in charge of international relations, spoke about the evolution of the GCIRC, Christophe Jestin, in charge of genetics and crop protection, as well as Xavier Pinochet, expert in scientific strategies, were also present and participated in the exchanges.

A workshop, carried out during this event, was the opportunity to share the different visions of breeders, research organizations and actors of the sector (GEVES and technical institutes) on the varietal characteristics and their importance in different production systems and the method to evaluate them.

Update on four projects funded by Promosol

The Carrefour was an opportunity to follow the progress of the four ongoing research projects funded by PROMOSOL:

DELUGE- Unlocking the genetic fight against the winter flea beetle (coordinators: Maxime Hervé & Anne Marie Cortesero (UMR IGEPP, Rennes)

There are concerns that growing rapeseed in France might encounter a deadlock due winter flea beetle (*Psylliodes chrysocephala*) control. This project proposes a genetic strategy with the creation of canola-resistant rapeseed varieties. Three obstacles hamper the development of this approach: the absence of resistance in *B. napus*, the lack of a standardized biotest to assess resistance to larvae and the absence of methodology for rearing the insect, which would allow to work with it in the laboratory all along the year.

The DELUGE project aims to identify a rapeseed related species resistant to the winter flea beetle, which could provide genetic resistance for future introgression in rapeseed. It also looks at complementing the current adults’ resistance biotest, developing larvae resistance tests and working on flea beetle breeding.

SEEDQUAL - Characterization of the genetic diversity of the composition of rapeseed and meal for uses in animal feed (coordinator: Nathalie Nesi, UMR IGEPP, Rennes)

Population growth and rising living standards will increase food consumption of vegetable oils and proteins by 2030. Vegetable oil resources will meet the demand for food and contribute to energy and chemistry demands in the future. On the other hand, the supply of oil cakes will be tight. Reducing the shortage of vegetable protein supplies in Europe is a major challenge for the competitiveness of the agricultural sector.

The main oil and protein crops are sources of vegetable proteins, which have hitherto been valued from oil extraction cakes. However, the quality of rapeseed meal is affected by factors intrinsic to the seed and by the crushing process, which is currently optimized for oil extraction. The challenge for the oil and protein crop sector is therefore to develop, in addition to the "noble" product that is oil, oil cake as a high value-added product.

The SEEDQUAL project therefore aims to provide a detailed description of the composition of rapeseed, in particular in fibers and proteins, to identify the genetic variability available for the various constituents and to develop medium-high speed tools to assess these variables in breeding programs.

SEEDQUAL partners have complementary skills that will allow them to combine quantitative genetics, structural genomics, functional genomics, and analytical biochemistry.

DESCRIBE - Chemotypic diversity in *Brassica napus* (coordinators: Antoine Gravot and Alain Bouchereau (UMR IGEPP, Rennes))

The objective of this project is to carry out, from a large panel of *B. napus* genotypes (winter and spring type), an in-depth chemotypic analysis of a few chemical families of rapeseed secondary metabolism, considered to be determining in the interactions with the biotic environment of plants. Many of these metabolic factors also contribute to the qualitative properties of the products, particularly the seed.

The targeted analytical approach especially considers the chemical constituents of the glucosinolates family, flavonoids, and volatile terpene compounds. The first step is to improve the analytical procedures for the qualitative and quantitative description of these bioactive substances and to adapt them for the establishment of plants phytochemical profiles. The choice to take into account a genetic diversity as wide as possible leads to limit the investigations to a single phenological stage (early vegetative development) and to reduce the number of tissues and organs (seeds, roots and leaves), from plants grown under optimal conditions.

Chemotypic analyses concern a panel of 250 rapeseed accessions representative of the valuable genetic diversity and used for association genetic analyses.

Metaphor- Use of metatranscriptomic data from "LeptoLife" for the identification and monitoring of new sources of phoma resistance in rapeseed (coordinator: Thierry Rouxel, INRAE UMR BIOGER)

The Metaphor project focuses on the interaction between rapeseed and the pathogens responsible for the phoma, *Leptosphaeria maculans* and *L. biglobosa*.

Its objective is to analyse and use the data from the "Leptolife" large-scale sequencing project to:

- Characterize the mechanisms giving resistance in the adult stage and the counteraction mechanisms of the fungus,
- Identify new genes involved in resistance (leaf resistance to ascospore infection, resistance at adult stage),
- Identify new AvrLm genes of the fungus expressed only in ascospores allowing to screen new Rlm genes in rapeseed genotypes,
- Identify candidate genes for screening plant genotypes showing increased adult resistance.

We can therefore hope that this work will provide new tools for the screening of genetic resources based on our previous experience of generation of genetically improved strains for the identification of Rlm genes.

Scientific news

Publications:

GENETICS & BREEDING

- Yang J, Wang Z, Jiang Y and Fei S (2019) Editorial: **Genetics and Genomics** of Polyploid Plants. *Front. Plant Sci.* 10:934. <https://doi.org/10.3389/fpls.2019.00934>
- PREPRINT: Bird, K. A., Niederhuth, C., Ou, S., Gehan, M., Pires, J. C., Xiong, Z.... & Patrick, P. Replaying the evolutionary tape to investigate **subgenome dominance in allopolyploid**. *Zoology*, 109, 261-276. <https://www.biorxiv.org/content/10.1101/814491v2.full.pdf>
- Song, J., Guan, Z., Hu, J. et al. Eight high-quality genomes reveal **pan-genome** architecture and ecotype differentiation of *Brassica napus*. *Nat. Plants* 6, 34–45 (2020). <https://doi.org/10.1038/s41477-019-0577-7>
- Chen R, Shimono A, Aono M, Nakajima N, Ohsawa R, Yoshioka Y (2020) **Genetic diversity** and population structure of feral rapeseed (*Brassica napus* L.) in **Japan**. *PLoS ONE* 15(1): e0227990. <https://doi.org/10.1371/journal.pone.0227990>
- Gong, Q., Lian, J., Li, X. et al. The **genetic diversity** and heterotic groups of 169 Chinese semi-winter rapeseed (*Brassica napus*) cultivars and inbred lines. *Mol Breeding* 40, 35 (2020). <https://doi.org/10.1007/s11032-020-01121-z>
- Shiranifar, B., Hobson, N., Kebede, B., Yang, R. C., & Rahman, H. (2020). Potential of **rutabaga** (*Brassica napus* var. *napobrassica*) **gene pool** for use in the breeding of *B. napus* canola. *Crop Science*, 60(1), 157-171. <https://doi.org/10.1002/csc2.20074>
- Uranjargal, B., & Enkhchimeg, V. (2019). The study on **inheritance pattern**, phenotype, and nutritional analysis of atgrf2 **transgenic rapeseed** (*brassica napus* L.). *Mongolian Journal of Agricultural Sciences*, 26(01), 94-100. <https://doi.org/10.5564/mjas.v26i01.1203>
- Zhang, K., He, J., Liu, L. et al. A convenient, rapid and efficient **method for** establishing **transgenic** lines of *Brassica napus*. *Plant Methods* 16, 43 (2020). <https://doi.org/10.1186/s13007-020-00585-6>
- Zhu, A., Wang, A., Zhang, Y., Dennis, E. S., Peacock, W. J., & Greaves, I. K. (2020). Early establishment of photosynthesis and auxin biosynthesis plays a key role in **early biomass heterosis** in *Brassica napus* (Canola) hybrids. *Plant and Cell Physiology*. <https://doi.org/10.1093/pcp/pcaa038>
- Gorlova, L. A., Bochkarova, E. B., Serdyuk, V. V., Strel'nikov, Ye. A., & Pomorova, YU. (2019). The **yellow-seeded variety** of spring rapeseed Kenar. *Масличные культуры*, (3 (179)). Russian, English abstract. <https://cyberleninka.ru/article/n/pervyy-otechestvennyy-zheltosemyanny-sort-rapsa-yarovogo-kenar>
- Qu, C., Yin, N., Chen, S., Wang, S., Chen, X., Zhao, H., ... & Liu, L. (2020). Comparative Analysis of the Metabolic Profiles of **Yellow-versus Black-Seeded** Rapeseed Using UPLC–HESI–MS/MS and

- Transcriptome Analysis. *Journal of Agricultural and Food Chemistry*, 68(10), 3033-3049. <https://doi.org/10.1021/acs.jafc.9b07173>
- Liu, S., Huang, H., Yi, X., Zhang, Y., Yang, Q., Zhang, C., ... & Zhou, Y. (2019). Dissection of **Genetic architecture for glucosinolate** accumulations in leaves and seeds of *Brassica napus* by Genome-wide Association Study. *Plant Biotechnology Journal*. <https://doi.org/10.1111/pbi.13314>
- Liu, Y., Zhou, X., Yan, M. et al. Fine mapping and candidate gene analysis of a **seed glucosinolate content QTL**, qGSL-C2, in rapeseed (*Brassica napus* L.). *Theor Appl Genet* 133, 479–490 (2020). <https://doi.org/10.1007/s00122-019-03479-x>
- Li, B., Gao, J., Chen, J. et al. Identification and fine mapping of a major locus controlling **branching** in *Brassica napus*. *Theor Appl Genet* 133, 771–783 (2020). <https://doi.org/10.1007/s00122-019-03506-x>
- Zeng Xin-hua, Zhao Sheng-bo, Li Ke-qi, Yuan Rong, Wu Gang, Yan Xiao-hong. (2019). Construction of a lhrRNAi library related to rapeseed **flower development** mediated by rolling circle replication. *CHINESE JOURNAL OF OIL CROP SCIENCES*, 41(3), 340. (*Chinese, English abstract and legends*) <http://www.jouoilcrops.cn/EN/10.7505/j.issn.1007-9084.2019.03.005>
- Chen Wei, Zu Feng, Luo Yan-qing, Zhao Kai-qing, Zhang Jian-kun, Zhang Guo-jian... & Wang Jing-qiao. (2019). Factors affecting the **number of single silique** in *Brassica napus*. *CHINESE JOURNAL OF OIL CROP SCIENCES*, 41(3), 331.(*Chinese, English abstract and legends*) <http://www.jouoilcrops.cn/EN/Y2019/V41/I3/331>
- Tang, M., Tong, C., Liang, L., Du, C., Zhao, J., Xiao, L., ... & Xiang, Y. (2020). A recessive **high-density pod** mutant resource of *Brassica napus*. *Plant Science*, 110411. <https://doi.org/10.1016/j.plantsci.2020.110411>
- Zhu, Y., Ye, J., Zhan, J., Zheng, X., Zhang, J., Shi, J., ... & Wang, H. (2020). Validation and Characterization of a **Seed Number Per Silique Quantitative Trait Locus** qSN. A7 in Rapeseed (*Brassica napus* L.). *Frontiers in Plant Science*, 11, 68. <https://doi.org/10.3389/fpls.2020.00068>
- Wang, H., Yan, M., Xiong, M. et al. Genetic dissection of **thousand-seed weight** and fine mapping of cqSW.A03-2 via linkage and association analysis in rapeseed (*Brassica napus* L.). *Theor Appl Genet* 133, 1321–1335 (2020). <https://doi.org/10.1007/s00122-020-03553-9>
- Wang, X., Zheng, M., Liu, H. et al. Fine-mapping and transcriptome analysis of a candidate gene controlling **plant height** in *Brassica napus* L. *Biotechnol Biofuels* 13, 42 (2020). <https://doi.org/10.1186/s13068-020-01687-y>
- Dao-zong, C. H. E. N., Yi, L. I. U., Wen-qin, F. U., Xian-hong, G. E., & Zai-yun, L. (2019). Progress on genetics and breeding of rapeseed (*Brassica napus* L.) with **colored flowers**. *CHINESE JOURNAL OF OIL CROP SCIENCES*, 41(3), 309. (*Chinese, English abstract and legends*) <http://www.jouoilcrops.cn/EN/abstract/abstract1736.shtml>
- Wenqin Fu Daozong Chen Qi Pan Fengfeng Li Zhigang Zhao Xianhong Ge Zaiyun Li (2018). Production of **red-flowered** oilseed rape via the ectopic expression of *O rynchophragmus* violaceus O v PAP 2. *Plant biotechnology journal*, 16(2), 367-380. <https://doi.org/10.1111/pbi.12777>
- Sashidhar, N., Harloff, H. J., Potgieter, L., & Jung, C. (2020). Gene editing of three BnITPK genes in tetraploid oilseed rape leads to significant **reduction of phytic acid** in seeds. *Plant Biotechnology Journal*. <https://doi.org/10.1111/pbi.13380>
- Ding, L., Gu, S., Zhu, F. et al. Long-chain acyl-CoA synthetase 2 is involved in **seed oil production** in *Brassica napus*. *BMC Plant Biol* 20, 21 (2020). <https://doi.org/10.1186/s12870-020-2240-x>

- Channaoui, S., Hssaini, L., Velasco, L., Mazouz, H., El Fechtali, M., & Nabloussi, A. (2020). Comparative Study of **Fatty Acid Composition**, Total Phenolics, and Antioxidant Capacity in Rapeseed **Mutant Lines**. Journal of the American Oil Chemists' Society, 97(4), 397-407. <https://doi.org/10.1002/aocs.12330>
- Karunarathna, N. L., Wang, H., Harloff, H. J., Jiang, L., & Jung, C. (2020). Elevating seed oil content in a polyploid crop by induced mutations in **seed fatty acid reducer** genes. Plant Biotechnology Journal. <https://doi.org/10.1111/pbi.13381>
- Zafar, S., Tang, M. Q., Liu, S. Y., & Tan, X. L. (2020). Candidate genes association study to identify allele-specific SNP **marker of ω -3 fatty acid** in *Brassica napus*. Journal of Plant Physiology, 153159. <https://doi.org/10.1016/j.jplph.2020.153159>
- Bai, S., Wallis, J. G., Denolf, P., Engelen, S., Bengtsson, J. D., Van Thournout, M., ... & Browse, J. (2020). The biochemistry of headgroup exchange during **triacylglycerol synthesis** in canola. The Plant Journal. <https://doi.org/10.1111/tpj.14709>
- Hussain, N., Xuan, L., Zhao, X., & Jiang, L. (2019). Allelic Variation of Bnax.Vte4.B and its Association with **α -/ γ -Tocopherol Ratio** in *Brassica napus* L. Journal of Genetics and Genomes. [REFERENCE](#)
- Qamar, H., Ilyas, M., Jan, S. A., Mustafa, H. S. B., Arshad, A., Yar, M. S., ... & Shinwari, Z. K. Recent trends in molecular breeding and biotechnology for the genetic improvement of Brassica species against **drought stress**. [REFERENCE](#)
- Poveda, J. (2020). Trichoderma parareesei Favors the Tolerance of Rapeseed (*Brassica napus* L.) to **Salinity and Drought** Due to a Chorismate Mutase. Agronomy, 10(1), 118. <https://doi.org/10.3390/agronomy10010118>
- Raman, H., Raman, R., Mathews, K., Diffey, D., & Salisbury, P. P. (2020). QTL mapping reveals **genomic regions for yield** based on incremental tolerance index to **drought stress** and related agronomic traits in canola. bioRxiv. <https://doi.org/10.1101/2020.01.06.896688>
- Fan, S., Liu, H., Liu, J., Hua, W., Xu, S., & Li, J. (2020). Systematic Analysis of the DNA Methylase and Demethylase Gene Families in Rapeseed (*Brassica napus* L.) and Their Expression Variations After **Salt and Heat stresses**. International journal of molecular sciences, 21(3), 953. <https://doi.org/10.3390/ijms21030953>
- Liu, J., Yang, R., Jian, N., Wei, L., Ye, L., Wang, R., ... & Zheng, Q. (2020). **Putrescine metabolism** modulates the biphasic effects of brassinosteroids on canola and Arabidopsis salt tolerance. Plant, Cell & Environment. <https://doi.org/10.1111/pce.13757>
- Motallebinia, S., Sofalian, O., Asghari, A., Rasoulzadeh, A., & Fathi, B. (2019). Study of **Drought Tolerance** Indices and Their Relationship with ISSR Markers in some Canola (*Brassica napus* L.) Cultivars. Journal of Plant Genetic Research, 6(1), 99-114. (Persian, English abstract) <https://journals.lu.ac.ir/pgr/article-1-151-en.html>
- Hossain, S. M., Masle, J., Easton, A., Hunter, M. N., Godwin, I. D., Farquhar, G. D., & Lambrides, C. J. (2020). Genetic variation for leaf carbon isotope discrimination and its association with **transpiration efficiency** in canola (*Brassica napus*). Functional Plant Biology, 47(4), 355-367. <https://doi.org/10.1071/FP19256>
- Koscielny, C., Gardner, S., Technow, F., & Duncan, R. Linkage mapping and whole genome predictions within *Brassica napus* L. subjected to differing temperature treatments. <https://doi.org/10.1071/CP19387>

- Jian H, Xie L, Wang Y, Cao Y, Wan M, Lv D, Li J, Lu K, Xu X, Liu L. 2020. Characterization of **cold stress responses** in different rapeseed ecotypes based on metabolomics and transcriptomics analyses. PeerJ 8:e8704 <https://doi.org/10.7717/peerj.8704>
- Wan, Y., Wang, Z., Xia, J., Shen, S., Guan, M., Zhu, M., ... & Lu, K. (2020). Genome-Wide Analysis of Phosphorus Transporter Genes in Brassica and Their Roles in **Heavy Metal Stress Tolerance**. International journal of molecular sciences, 21(6), 2209. <https://doi.org/10.3390/ijms21062209>
- Zhang, F., Xiao, X., Xu, K. et al. Genome-wide association study (GWAS) reveals genetic loci of **lead (Pb) tolerance** during seedling establishment in rapeseed (*Brassica napus* L.). BMC Genomics 21, 139 (2020). <https://doi.org/10.1186/s12864-020-6558-4>
- Wang, J., Jin, Z., Zhou, M., Yu, Y., & Liang, M. (2020). Characterization of NF-Y transcription factor families in industrial rapeseed (*Brassica napus* L.) and identification of BnNF-YA3, which functions in the **abiotic stress response**. Industrial Crops and Products, 148, 112253. <https://doi.org/10.1016/j.indcrop.2020.112253>
- Lohani, N., Jain, D., Singh, M. B., & Bhalla, P. L. (2020). Engineering **Multiple Abiotic Stress Tolerance** in Canola, *Brassica napus*. Frontiers in Plant Science, 11. <https://doi.org/10.3389/fpls.2020.00003>
- Huang, Q., Lv, J., Sun, Y., Wang, H., Guo, Y., Qu, G., & Hu, S. (2020). Inheritance and Molecular Characterization of a Novel Mutated AHAS Gene Responsible for the **Resistance of AHAS-Inhibiting Herbicides** in Rapeseed (*Brassica napus* L.). International journal of molecular sciences, 21(4), 1345. <https://doi.org/10.3390/ijms21041345>
- Mei, J., Shao, C., Yang, R. et al. Introgression and pyramiding of genetic loci from wild *Brassica oleracea* into *B. napus* for improving **Sclerotinia resistance** of rapeseed. Theor Appl Genet 133, 1313–1319 (2020). <https://doi.org/10.1007/s00122-020-03552-w>
- Chittem, K., Yajima, W. R., Goswami, R. S., & del Río Mendoza, L. E. (2020). Transcriptome analysis of the plant pathogen **Sclerotinia sclerotiorum** interaction with resistant and susceptible canola (*Brassica napus*) lines. Plos one, 15(3), e0229844. <https://doi.org/10.1371/journal.pone.0229844>
- Lingyi, Z., Xiuzhen, W., Li, X., Huan, Y., Wang, C., Fan, L., ... & Xiaoping, F. (2019). Resynthesis of **clubroot** disease resistant rapeseed (AAC~ r C~ r and A~ r A~ r C~ r C~ r) through hybridization. 中国植物病理学会 2019 年学术年会论文集. (Chinese, English abstract) <http://cpfd.cnki.com.cn/Article/CPFDTOTAL-ZGVS201907001045.htm>
- Aigu, Y., Cao, T., Strelkov, I. S., Manolii, V. P., Lemoine, J., Manzaneres-Dauleux, M. J., ... & Gravot, A. (2020). Identification of winter and spring *Brassica napus* genotypes with partial resistance to Canadian isolates of **Plasmodiophora brassicae**. Canadian Journal of Plant Pathology, 1-9. <https://doi.org/10.1080/07060661.2020.1723870>
- Shah, N., Li, Q., Xu, Q., Liu, J., Huang, F., Zhan, Z., ... & Zhang, C. (2020). CRb and PbBa8. 1 Synergically Increases Resistant Genes Expression upon Infection of **Plasmodiophora brassicae** in *Brassica napus*. Genes, 11(2), 202. <https://doi.org/10.3390/genes11020202>
- Bousset, L., Ermel, M., & Delourme, R. (2020). A **Leptosphaeria maculans** set of isolates characterised on all available differentials and used as control to identify virulence frequencies in a current French population. bioRxiv. <https://doi.org/10.1101/2020.01.09.900167>
- Raman, R., Diffey, S., Barbulescu, D.M. et al. Genetic and physical mapping of loci for resistance to **blackleg disease** in canola (*Brassica napus* L.). Sci Rep 10, 4416 (2020). <https://doi.org/10.1038/s41598-020-61211-y>

Yao, L., Yang, B., Xian, B., Chen, B., Yan, J., Chen, Q., ... & Jiang, Y. Q. (2020). The R2R3-MYB transcription factor BnaMYB111L from rapeseed modulates reactive oxygen species accumulation and hypersensitive-like cell death. *Plant Physiology and Biochemistry*, 147, 280-288. <https://doi.org/10.1016/j.plaphy.2019.12.027>

BOOK:

Shabir Hussain Wani, Ajay Kumar Thakur, Yasin Jeshima Khan. *Brassica Improvement. Molecular, Genetics and Genomic Perspectives*. Springer Cham. <https://doi.org/10.1007/978-3-030-34694-2>

CROP PROTECTION

Dev, D. Pathotype variation of *Albugo candida*, the cause of white rust disease in rapeseed-mustard (Doctoral dissertation, GB Pant University of Agriculture and Technology, Pantnagar-263145 (Uttarakhand)). <https://krishikosh.egranth.ac.in/handle/1/5810136699>

Al-lami, H. F. D., You, M. P., Mohammed, A. E., & Barbetti, M. J. Virulence variability across the *Alternaria spp.* population determines incidence and severity of **Alternaria leaf spot** on rapeseed. *Plant Pathology*. <https://doi.org/10.1111/ppa.13135>

Dhaliwal, R.S., Singh, B. Effect of weather parameters and date of sowing on intensity of **Alternaria blight** of rapeseed mustard. *Indian Phytopathology* 73, 89–95 (2020). <https://doi.org/10.1007/s42360-020-00193-3>

Blagojević, J. D., Vukojević, J. B., & Ivanović, Ž. S. Occurrence and characterization of **Alternaria** species associated with leaf spot disease in rapeseed in Serbia. *Plant Pathology*. <https://doi.org/10.1111/ppa.13168>

Khun, P. R., Kulczynski, S. M., Bellé, C., Ramos, R. F., & Gabriel, M. (2020). Métodos de inoculação artificial de *Alternaria alternata* em sementes de canola. *Revista Eletrônica Científica da UERGS*, 6(1), 20-28. (Portuguese) <http://200.132.92.80/index.php/revuergs/article/view/2307>

Al-lami, H. F., You, M. P., & BARBETTI, M. J. (2020). Temperature drives contrasting **Alternaria Leaf Spot** epidemic development in canola and mustard rape from *Alternaria japonica* and *A. brassicae*. *Plant Disease*, (ja). <https://doi.org/10.1094/PDIS-10-19-2251-RE>

Monnier, N., Cordier, M., Dahi, A., Santoni, V., Guénin, S., Clément, Sarazin C., Penaud A., Dorey S., Cordelier S. & Rippa, S. (2020). Semipurified Rhamnolipid Mixes Protect *Brassica napus* Against **Leptosphaeria maculans** Early Infections. *Phytopathology*, PHYTO-07. <https://doi.org/10.1094/PHYTO-07-19-0275-R>

Peng, G., Liu, X., McLaren, D. L., McGregor, L., & Yu, F. (2020). Seed treatment with the fungicide fluopyram limits cotyledon infection by **Leptosphaeria maculans** and reduces blackleg of canola. *Canadian Journal of Plant Pathology*, 1-13. <https://doi.org/10.1080/07060661.2020.1725132>

Bousset, L., Ermel, M., & Delourme, R. (2020). A **Leptosphaeria maculans** set of isolates characterised on all available differentials and used as control to identify virulence frequencies in a current French population. *bioRxiv*. <https://doi.org/10.1101/2020.01.09.900167>

Yang, Y., Marcoft, S. J., Forsyth, L. M., Zhao, J., Li, Z., Van de Wouw, A. P., & Idnurm, A. (2020). Sterol Demethylation Inhibitor Fungicide Resistance in **Leptosphaeria maculans** is Caused by Modifications in the Regulatory Region of ERG11. *Plant Disease*, PDIS-10. <https://doi.org/10.1094/PDIS-10-19-2088-RE>

- Wang, Y., Akhavan, A., Hwang, S. F., & Strelkov, S. E. (2020). Decreased sensitivity of *Leptosphaeria maculans* to pyraclostrobin in Alberta, Canada. *Plant Disease*, (ja). <https://doi.org/10.1094/PDIS-11-19-2461-RE>
- Wang, Y., Strelkov, S. E., & Hwang, S. F. (2020). Yield losses in canola in response to **blackleg** disease. *Canadian Journal of Plant Science*, (ja). <https://doi.org/10.1139/CJPS-2019-0259>
- Sedaghatkish, A. (2020). The genomic structure and management of *Plasmodiophora brassicae* (Doctoral dissertation University of Guelph). <https://atrium2.lib.uoguelph.ca/xmlui/handle/10214/17833>
- Luo Yan-qing, Wang Yun-yue, Zu Feng, Fu Ming-lian, Zhao Kai-qin, Zhang Yun-yun... & Yuan Xiao-yan, (2019). Transcriptome analysis of *Brassica napus-Plasmodiophora brassicae* interaction during early infection. *Chinese Journal of Oil Crop Sciences*, 41(3), 421. (*Chinese, English summary & legends*). <http://www.jouoilcrops.cn/EN/Y2019/V41/I3/421>
- Fei, W., Chu, M., Jiang, Y., & Hwang, S. F. (2019). First report of *Plasmodiophora brassicae* causing **clubroot** on *Raphanus sativus* in China. *Plant Disease*, (ja). <https://doi.org/10.1094/PDIS-08-19-1629-PDN>
- Allan, J., Regmi, R., Denton-Giles, M., Kamphuis, L. G., & Derbyshire, M. C. (2019). The host generalist phytopathogenic fungus *Sclerotinia sclerotiorum* differentially expresses multiple metabolic enzymes on two different plant hosts. *Scientific Reports*, 9(1), 1-15. <https://doi.org/10.1038/s41598-019-56396-w>
- Shahoveisi, F., & del Rio Mendoza, L. E. (2020). Effect of wetness duration and incubation temperature on development of ascospore infections by *Sclerotinia sclerotiorum*. *Plant Disease*, (ja). <https://doi.org/10.1094/PDIS-06-19-1304-RE>
- Zhang, J., Mavrodi, D. V., Yang, M., Thomashow, L. S., Mavrodi, O. V., Kelton, J., & Weller, D. M. (2020). *Pseudomonas synxantha* 2-79 Transformed with Pyrrolnitrin Biosynthesis Genes Has Improved Biocontrol Activity Against **Soilborne Pathogens** of Wheat and Canola. *Phytopathology*, PHYTO-09. <https://doi.org/10.1094/PHYTO-09-19-0367-R>
- Bhatta, K., Chaulagain, L., Kafle, K., & Shrestha, J. Bio-Efficacy of Plant Extracts against **Mustard Aphid** (*Lipaphis erysimi* Kalt.) on Rapeseed (*Brassica campestris* Linn.) under Field and Laboratory Conditions. *Syrian Journal of Agricultural Research—SJAR* 6(4):557-566 December 2019 <http://agri-research-journal.net/sjar/wp-content/uploads/2019/12/v6n4p44.pdf>
- Kafle, K., & Jaishi, M. (2020). Farmer's Management Practices Adopted Against **Mustard Aphid**, *Lipaphis erysimi* (Kalt.): A Survey of Chitwan, Nepal. *International Journal of Applied Sciences and Biotechnology*, 8(1), 78-82. <https://doi.org/10.3126/ijasbt.v8i1.28255>
- Fathipour, Y., Kianpour, R., Bagheri, A., Karimzadeh, J., Hosseini Naveh, V., & Mehrabadi, M. (2020). Targeting *Plutella xylostella* digestive enzymes by applying resistant Brassicaceae host cultivars. *Journal of Crop Protection*, 9(1), 65-79. <https://jcp.modares.ac.ir/article-3-31365-en.pdf>
- Nouri-Ganbalani, G., Naseri, B., Majd-Marani, S. et al. Canola cultivars affect nutrition and cold hardiness of *Plutella xylostella* (L.) (Lepidoptera: Plutellidae). *Int J Trop Insect Sci* (2020). <https://doi.org/10.1007/s42690-020-00125-8>
- Mahdieh Jafary-Jahed, Jabraeil Razmjou, Gadir Nouri-Ganbalani, Bahram Naseri, and Mahdi Hassanpour "Bottom-Up Effects of Organic Fertilizers on *Plutella xylostella* (L) with Selected Cruciferous Crop Plants," *The Journal of the Lepidopterists' Society* 74(1), 7-17, (16 March 2020). <https://doi.org/10.18473/lepi.74i1.a2>

- Congdon, B. S., Baulch, J. R., & Coutts, B. A. (2020). Impact of **Turnip yellows virus** infection on seed yield of an open-pollinated and hybrid canola cultivar when inoculated at different growth stages. *Virus Research*, 277, 197847. <https://doi.org/10.1016/j.virusres.2019.197847>
- KİTİŞ, Yasin Emre, GRENZ, Jan Hendrik, SAUERBORN, Joachim. "Effects of some cereal root exudates on germination of **broomrapes** (*Orobancha spp.* and *Phelipanche spp.*)". *Mediterranean Agricultural Sciences* 32 / 2 (August 2019): 145-150. <https://doi.org/10.29136/mediterranean.546564>
- Mwendwa, J. M., Brown, W. B., Weston, P. A., Bagherieh-Najjar, M. B., & Preston, C. (2019). **Evaluation** of Selected Commercial Canola Cultivars for **Early Vigour, Weed Suppression** and Yield in Southern New South Wales. *Mechanisms of Weed Suppression in Wheat (*Triticum aestivum L.*) and Canola (*Brassica napus L.*) in Southeastern Australia*, 105. [REFERENCE](#)
- Shamaya, N., Raman, H., Rohan, M., Pratley, J., & Wu, H. (2020). Validation of **Competitive Ability** of Diverse Canola Accessions against Annual Ryegrass under Glasshouse and Field Conditions. *Open Journal of Genetics*, 10(02), 17. <https://doi.org/10.4236/ojgen.2020.102003>
- Munsif, F., Ali, A., Alam, J. E., Hussian, J., & Ahmad, B. (2019). Influence of Plant Population on **Weed Infestation** and Yield of Rapeseed Cultivars. *EC Agriculture*, 6(1), 01-06. <https://www.econicon.com/ecag/influence-of-plant-population-on-weed-infestation-and-yield-of-rapeseed-cultivars.php>
- Asad, M. H., & Bais, A. (2019). **Weed Detection** in Canola Fields Using Maximum Likelihood Classification and Deep Convolutional Neural Network. *Information Processing in Agriculture*. <https://doi.org/10.1016/j.inpa.2019.12.002>
- Szpyrka, E., Słowik-Borowiec, M., Książek, P. et al. The difference in dissipation of **clomazone and metazachlor** in soil under field and laboratory conditions and their uptake by plants. *Sci Rep* 10, 3747 (2020). <https://doi.org/10.1038/s41598-020-60720-0>

AGRONOMY

- Moradi Aghdam A., Sayfzadeh S., Shirani Rad A.H., Valadabadi S.A., Zakerin H.R. The assessment of **water stress and delay cropping** on quantitative and qualitative traits of rapeseed genotypes. *Industrial Crops and Products*, Volume 131, 2019, <https://doi.org/10.1016/j.indcrop.2019.01.051>
- Farhangi-Abriz, S., Tavasolee, A., Ghassemi-Golezani, K. et al. **Growth-promoting bacteria** and natural regulators mitigate **salt toxicity** and improve rapeseed plant performance. *Protoplasma* (2020). <https://doi.org/10.1007/s00709-020-01493-1>
- Elizareva, E., Yanbaev, Y., Redkina, N., Kudashkina, N., Elizaryev, A., & Khamidullin, I. (2019, December). **Accumulating capacity** of different varieties of rapeseed under conditions of anthropogenic pollution of soils by **heavy metals**. In *IOP Conference Series: Earth and Environmental Science* (Vol. 403, No. 1, p. 012182). IOP Publishing. <https://iopscience.iop.org/article/10.1088/1755-1315/403/1/012182/meta>
- Shah, T., Munsif, F., D'amato, R., & Nie, L. (2020). **Lead toxicity** induced phytotoxic impacts on rapeseed and clover can be lowered by biofilm forming lead tolerant bacteria. *Chemosphere*, 246, 125766. <https://doi.org/10.1016/j.chemosphere.2019.125766>
- Churilova, V. V., Churilov, G. I., Churilov, D. G., Polischuk, S. D., & Arapov, I. S. (2020, January). Effect of **metal nanoparticles** on the **accumulation** and structure of rapeseed carbohydrates. In *IOP*

- Conference Series: Earth and Environmental Science (Vol. 422, No. 1, p. 012089). IOP Publishing. <https://iopscience.iop.org/article/10.1088/1755-1315/422/1/012089/meta>
- Yuan, T., Gu, J., Zhou, H. et al. Translocation and **accumulation of cadmium and lead** in the tissues of 39 rape cultivars grown in a polluted farmland. Environ Sci Pollut Res (2020). <https://doi.org/10.1007/s11356-020-07697-5>
- Al-Awad, O. (2019). Resistance to **ammonium toxicity** in canola genotypes. PhD thesis University of Western Australia. <https://research-repository.uwa.edu.au/en/publications/resistance-to-ammonium-toxicity-in-canola-genotypes>
- Li Yan-hua, Shi You-ming, Zhou Yan, Huang Hua-lei, Xiao Chang-ming, Huang Shu-qin, ... & Zhang Xiao-chun (2019). **Dry matter accumulation** and NPK absorption and utilization of rapeseed for oilseed-vegetable. Chinese Journal of Oil Crop Sciences, 41(3). [REFERENCE](#)
- Feng, J., Hussain, H. A., Hussain, S., Shi, C., Cholidah, L., Men, S., ... & Wang, L. (2020). Optimum **Water and Fertilizer Management** for Better Growth and Resource Use Efficiency of Rapeseed in Rainy and Drought Seasons. Sustainability, 12(2), 703. <https://doi.org/10.3390/su12020703>
- Mamnabi, S., Nasrollahzadeh, S., Ghassemi-Golezani, K., & Raei, Y. (2020). Improving yield-related physiological characteristics of spring rapeseed by integrated **fertilizer management** under water deficit conditions. Saudi Journal of Biological Sciences, 27(3), 797-804. <https://doi.org/10.1016/j.sjbs.2020.01.008>
- Cong, R., Liu, T., Lu, P. et al. **Nitrogen fertilization** compensation the weak **photosynthesis** of Oilseed rape (*Brassica napus* L.) under haze weather. Sci Rep 10, 4047 (2020). <https://doi.org/10.1038/s41598-020-60695-y>
- Porter, M. J., Pan, W. L., Schillinger, W. F., Madsen, I. J., Sowers, K. E., & Tao, H. Winter canola response to soil and fertilizer **nitrogen** in semiarid Mediterranean conditions. Agronomy Journal. <https://doi.org/10.1002/agj2.20119>
- Porter, M. J., Pan, W. L., Schillinger, W. F., Madsen, I. J., Sowers, K. E., & Tao, H. (2020). Winter Canola Response to Soil and Fertilizer **Nitrogen**. Crops & Soils. <https://doi.org/10.1002/crso.20020>
- Swanepoel, P. A., & Labuschagne, J. (2020). Canola is injured by in-row **nitrogen placement** associated with disc openers, but not by tine openers. Crop Science, 60(1), 466-474. <https://doi.org/10.1002/csc2.20084>
- Lin, Y., Watts, D. B., Torbert, H. A., & Howe, J. A. Influence of **Nitrogen** Rate on Winter Canola Production in the **Southeastern US**. Agronomy Journal. <https://doi.org/10.1002/agj2.20197>
- Younis, M., Muhammad, A., Alam, S., & Jalal, A. (2020). **Sulphur** doses and application times on yield and oil quality of canola grown in calcareous soil. Grasas y Aceites, 71(1), 341. <https://doi.org/10.3989/gya.1176182>
- Rahman, M. N., Hanga, R., & Schoenau, J. (2020). Influence of soil temperature and moisture on micronutrient supply, plant uptake, and biomass yield of wheat, pea, and canola. Journal of Plant Nutrition, 43(6), 823-833. <https://doi.org/10.1080/01904167.2020.1711941>
- Sun, Z., Su, R. L., Xu, P., Wu, H. T., Hu, J. L., Zhao, J. S., & Hu, R. G. (2019). Effect of Phosphorus Addition on **N₂O Emissions** from Rice-Rapeseed Rotation Soils. Huan jing ke xue= Huanjing kexue, 40(7), 3355.(Chinese, English abstract) <https://www.ncbi.nlm.nih.gov/pubmed/31854738>
- Tian, X., Li, Z., Wang, L., Wang, Y., Li, B., Duan, M., & Liu, B. (2020). Effects of **Biochar** Combined with Nitrogen Fertilizer Reduction on Rapeseed Yield and Soil Aggregate Stability in Upland of Purple Soils. International Journal of Environmental Research and Public Health, 17(1), 279. <https://doi.org/10.3390/ijerph17010279>

- Ashkiani, A., Sayfzadeh, S., Shirani Rad, A. H., Valadabadi, A., & Hadidi Masouleh, E. (2020). Effects of foliar **zinc** application on yield and oil quality of rapeseed genotypes under **drought stress**. Journal of Plant Nutrition, 1-10. <https://doi.org/10.1080/01904167.2020.1739299>
- Cui, C., Xie, X., Wang, L. Y., Wang, R. L., Lei, W., Lv, J., ... & Zhou, Q. Y. (2020). Photosynthetic index and nitrogen assimilation in rapeseed seedlings transplanted in soil with ammonium **glufosinate**. Ciência Rural, 50(4). [REFERENCE](#)
- Fioreze, S. L., Vacari, J., Turek, T. L., Michelon, L. H., & Drun, R. P. (2019). Growth and yield parameters of white oat and wheat as affected by **canola residues**. Revista Ceres, 66(6), 416-421. <https://doi.org/10.1590/0034-737x201966060002>
- Monfared, B.B., Noormohamadi, G., Rad, A.H.S. et al. Effects of Sowing Date and **Chitosan** on Some Characters of Canola (*Brassica napus L.*) Genotypes. J. Crop Sci. Biotechnol. 23, 65–71 (2020). <https://doi.org/10.1007/s12892-019-0177-0>
- Liersch, A., Bocianowski, J., Poplawska, W., Wielebski, F., & Bartkowiak-Broda, I. (2020). Chemical and molecular characteristics of winter **oilseed rape** (*Brassica napus L.*) **volunteers** from the soil seed bank. Journal of Research in Weed Science, 3(3. pp. 254-411), 391-411. <https://doi.org/10.26655/JRWEEDSCI.2020.3.10>
- Umurzokov, M., Jia, W., Cho, K. M., Khaitov, B., Sohn, S. I., Cho, J. W., & Park, K. W. (2019). Persistence, Viability and Emergence Rate of Canola (*Brassica napus L.*) in Korean Soil. Weed&Turfgrass Science, 8(4), 309-318. <https://doi.org/10.5660/WTS.2019.8.4.309>
- Petrova, S.N., Andronov, E.E., Belimov, A.A. et al. **Prokaryotic Community** Structure in the Rapeseed (*Brassica napus L.*) **Rhizosphere** Depending on Addition of 1-Aminocyclopropane-1-Carboxylate-Utilizing Bacteria. Microbiology 89, 115–121 (2020). <https://doi.org/10.1134/S0026261720010117>
- Floc'h, J., Hamel, C., Harker, K.N. et al. Fungal Communities of the Canola **Rhizosphere**: Keystone Species and Substantial Between-Year Variation of the Rhizosphere Microbiome. Microb Ecol (2020). <https://doi.org/10.1007/s00248-019-01475-8>
- Wysocki, D. (2020). Can Winter Canola Be Grown on Wide **Row Spacing**?. Crops & Soils, 53(1), 10-15. <https://doi.org/10.1002/crso.20000>
- Kudnig, J. Exploring the **effects of seed size** and target plant densities on the yield of hybrid canola across Australia. [REFERENCE](#)
- Wynne, K., Neely, C. B., Adams, C., Kimura, E., DeLaune, P. B., Hathcoat, D., & Gerrish, B. Testing **row spacing** and planting rate for fall-planted spring canola in the southern United States. Agronomy Journal. <https://doi.org/10.1002/agj2.20201>
- Zaheer, S., Arif, M., Akhtar, K., Khan, A., Khan, A., Bibi, S., ... & Ain, N. U. (2020). **Grazing and Cutting** under Different Nitrogen Rates, Application Methods and Planting Density Strongly Influence Qualitative Traits and Yield of Canola Crop. Agronomy, 10(3), 404. <https://doi.org/10.3390/agronomy10030404>
- Amiri, Z., Asgharipour, M. R., Campbell, D. E., & Armin, M. (2020). Extended exergy analysis (EAA) of two canola **farming systems** in Khorramabad, **Iran**. Agricultural Systems, 180, 102789. <https://doi.org/10.1016/j.agsy.2020.102789>
- Ma, L., Wang, X., Pu, Y. et al. Ecological and economic benefits of planting winter rapeseed (*Brassica rapa L.*) in the wind erosion area of **northern China**. Sci Rep 9, 20272 (2019). <https://doi.org/10.1038/s41598-019-56678-3>

- Vasilij G. Vasin, Altynaj B. Abuova, Saniya A. Tulkubaeva, Dinara B. Zhamalova and Marat B. Tashmuhamedov. Culture of priority oil crops in the north of **Kazakhstan**. BIO Web Conf., 17 (2020) 00029. <https://doi.org/10.1051/bioconf/20201700029>
- Postovalov, A., Sukhanova, S., Plotnikov, A., Sazhina, S., & Sozinov, A. (2020). Formation of Highly Productive Agrophytocenoses of Peas and Spring Rapeseed in **Trans-Urals**. KnE Life Sciences, 475-481. <https://www.knepublishing.com/index.php/KnE-Life/article/view/6109>
- Gebeltová, Z.; Malec, K.; Maitah, M.; Smutka, L.; Appiah-Kubi, S.N.K.; Maitah, K.; Sahatqija, J.; Sirohi, J. The Impact of **Crop Mix** on Decreasing Soil Price and Soil Degradation: A Case Study of Selected Regions in Czechia (2002–2019). Sustainability 2020, 12, 444. <https://doi.org/10.3390/su12020444>
- Meng, S., Zhong, Y., Luo, C., Hu, X., Wang, X., & Huang, S. (2020). Optimal Temporal Window Selection for Winter Wheat and **Rapeseed Mapping with Sentinel-2** Images: A Case Study of Zhongxiang in China. Remote Sensing, 12(2), 226. <https://doi.org/10.3390/rs12020226>
- Xu, S., Wang, J., Tian, H., & Wang, B. (2019). **Automatic Measuring** Approach and Device for Mature Rapeseed's Plant Type Parameters. Journal of Electrical and Computer Engineering, 2019. <https://doi.org/10.1155/2019/6834290>
- Trif A, Gidea M, Cioineag C, Cimpeanu S.M. The usage of the FAE fixed wing **UAV** for the evaluation of affected rapeseed culture due to **pest attacks** [REFERENCE](#)
- Hussain, S., Gao, K., Din, M., Gao, Y., Shi, Z., & Wang, S. (2020). Assessment of **UAV-Onboard** Multispectral Sensor for Non-Destructive Site-Specific Rapeseed Crop **Phenotype** Variable at Different Phenological Stages and Resolutions. Remote Sensing, 12(3), 397. <https://doi.org/10.3390/rs12030397>
- White, J., Berg, A. A., Champagne, C., Zhang, Y., Chipanshi, A., & Daneshfar, B. (2020). Improving crop **yield forecasts** with satellite-based soil moisture estimates: An example for township level canola yield forecasts over the Canadian Prairies. International Journal of Applied Earth Observation and Geoinformation, 89, 102092. <https://doi.org/10.1016/j.jag.2020.102092>
- Rahman, G., Sohag, H., Chowdhury, R., Wahid, K. A., Dinh, A., Arcand, M., & Vail, S. (2020). SoilCam: A Fully Automated **Minirhizotron** using Multispectral Imaging for Root Activity Monitoring. Sensors, 20(3), 787. <https://doi.org/10.3390/s20030787>

Physiology

- Verdejo, J., & Calderini, D. F. (2020). **Plasticity of seed weight** in winter and spring rapeseed is higher in a narrow but different window after flowering. Field Crops Research, 250, 107777. <https://doi.org/10.1016/j.fcr.2020.107777>
- Maleki, K., Soltani, E., Alahdadi, I., & Ghorbani Javid, M. Evaluation of Primary Conditional **Dormancy** in Seeds of Oilseed Rape (*Brassica napus*) Produced in Golestan and Mazandaran Provinces. Iranian Journal of Seed Research, 31-43. http://yujs.yu.ac.ir/jisr/browse.php?a_id=405&sid=1&slc_lang=en&ftxt=0
- Tatari, S., Ghaderi-Far, F., Yamchi, A., Siahmarguee, A., Shayanfar, A., & Baskin, C. C. (2020). Application of the **hydrotime model** to assess **seed priming** effects on the germination of rapeseed (*Brassica napus L.*) in response to water stress. Botany, (999), 1-9. <https://doi.org/10.1139/cjb-2019-0192>
- Shayanfar, A., Ghaderi-Far, F., Behmaram, R., Soltani, A., & Sadeghipour, H. R. (2020). Impacts of fire cues on the germination of *Brassica napus L.* seeds with high and low levels of **secondary dormancy**. Plant Biology. <https://doi.org/10.1111/plb.13115>

- Malek, M., Ghaderi-Far, F., Torabi, B., & Sadeghipour, H. R. The Effect of **Priming** on Seed Viability of Canola (*Brassica napus*) Cultivars under Different **Storage** Conditions. Iranian Journal of Seed Research, 45-60. <http://yujs.yu.ac.ir/jisr/article-1-409-en.html&sw=Seed+Longevity>
- Maliba, B. G. (2019). Photosynthetic responses of canola and wheat to elevated levels of CO₂ and O₃ in open-top chambers (Doctoral dissertation, North-West University (South Africa)). <http://repository.nwu.ac.za/handle/10394/33877>
- Kovaleski, S., Heldwein, A. B., Dalmago, G. A., & de Gouvêa, J. A. (2020). **Frost damage** to canola (*Brassica napus* L.) during reproductive phase in a controlled environment. Agrometeoros, 27(2). <http://seer.sct.embrapa.br/index.php/agrometeoros/article/view/26463>
- Doğru, A., Çakırlar, H. Effects of leaf age on chlorophyll fluorescence and antioxidant enzymes activity in winter rapeseed leaves under **cold acclimation** conditions. Braz. J. Bot 43, 11–20 (2020). <https://doi.org/10.1007/s40415-020-00577-9>
- Davoudi, A., Zeinalzadeh-Tabrizi, H., & Shirani-Rad, A. Effect of **selenium** foliar application on some quantitative and qualitative characteristics of rapeseed cultivars under end-season **thermal stress**. Journal of Crop Breeding, 0-0. (Persian, English abstract) http://jcb.sanru.ac.ir/browse.php?a_id=1076&sid=1&slc_lang=en
- Wang, F., Zhong, F., Zhang, S., Zhang, P., Chen, F., Li, W., & ZHANG, S. (2020). **Triadimefon** increases **drought tolerance** through the regulation of photosynthesis and carbohydrate metabolism in rapeseed at bolting stage. Photosynthetica, 58(1), 100-109. <https://ps.ueb.cas.cz/pdfs/phs/2020/01/12.pdf>
- Dai, L., Li, J., Harmens, H., Zheng, X., & Zhang, C. (2020). Melatonin enhances **drought resistance** by regulating leaf stomatal behaviour, root growth and catalase activity in two contrasting rapeseed (*Brassica napus* L.) genotypes. Plant Physiology and Biochemistry, 149, 86-95. <https://doi.org/10.1016/j.plaphy.2020.01.039>
- Keshavarzian, M., Toorchi, M., & Shakiba, M. R. Sodium Chloride **Salt Tolerance** Evaluation and Classification of Spring Rapeseed (*Brassica napus* L.). Journal of Crop Breeding, 0-0. (Persian, English abstract) http://jcb.sanru.ac.ir/browse.php?a_id=410&sid=1&slc_lang=en&ftxt=0
- Akhavan Hezaveh, T., Pourakbar, L., Rahmani, F., & Alipour, H. (2020). Effects of ZnO NPs on **phenolic compounds** of rapeseed seeds under **salinity stress**. Journal of Plant Process and Function, 8(34), 11-18. https://jispp.iut.ac.ir/browse.php?a_code=A-10-69-4&slc_lang=en&sid=1
- Naveed, M., Sajid, H., Mustafa, A., Niamat, B., Ahmad, Z., Yaseen, M., ... & Chen, J. T. (2020). Alleviation of **Salinity**-Induced Oxidative Stress, Improvement in Growth, Physiology and Mineral Nutrition of Canola (*Brassica napus* L.) through Calcium-Fortified Composted Animal Manure. Sustainability, 12(3), 846. <https://doi.org/10.3390/su12030846>
- Ma Ni, Wan Lin, Zhao Wei, Liu H. F., Li Jun, & ZHANG C. L. (2020). Exogenous **strigolactones** promote lateral **root growth** by reducing the endogenous auxin level in rapeseed. Journal of Integrative Agriculture, 19(2), 465-482. [https://doi.org/10.1016/S2095-3119\(19\)62810-8](https://doi.org/10.1016/S2095-3119(19)62810-8)

QUALITY and PRODUCTS

- Barthet V. J., Petryk M.W.P., Siemens B. Rapid **Nondestructive Analysis** of Intact Canola Seeds Using a Handheld Near-Infrared Spectrometer <https://doi.org/10.1002/aocs.12335>

- Rokosik, Ewa, Krzysztof Dwiecki, and Aleksander Siger. "The quality of **cold-pressed rapeseed oil** obtained from seeds of *Brassica napus* L. with increased moisture content." *Acta Scientiarum Polonorum Technologia Alimentaria* 18.2 (2019): 205-218. <https://doi.org/10.17306/J.AFS.2019.0672>
- Chew, S. C. (2020). **Cold-pressed rapeseed (*Brassica napus*) oil**: Chemistry and functionality. *Food Research International*, 108997. <https://doi.org/10.1016/j.foodres.2020.108997>
- Mohseni, N. M., Mirzaei, H. O., & Moghimi, M. (2020). Optimization of producing oil and meal from canola seeds using **microwave– pulsed electric field pretreatment**. *OCL*, 27, 2.<https://doi.org/10.1051/ocl/2019050>
- Romero-Guzmán, M. J., Jung, L., Kyriakopoulou, K., Boom, R. M., & Nikiforidis, C. V. (2020). Efficient single-step rapeseed **oleosome extraction** using twin-screw press. *Journal of Food Engineering*, 276, 109890. <https://doi.org/10.1016/j.jfoodeng.2019.109890>
- Romero-Guzmán, M. J., Köllmann, N., Zhang, L., Boom, R. M., & Nikiforidis, C. V. (2020). Controlled **oleosome extraction** to produce a plant-based mayonnaise-like emulsion using solely rapeseed seeds. *LWT*, 123, 109120. <https://doi.org/10.1016/j.lwt.2020.109120>
- Asl, P. J., Niazmand, R., & Yahyavi, F. (2020). **Extraction of phytosterols and tocopherols** from rapeseed oil waste by supercritical CO₂ plus co-solvent: A comparison with conventional solvent extraction. *Heliyon*, 6(3), e03592. <https://doi.org/10.1016/j.heliyon.2020.e03592>
- Sikorska, E., Wójcicki, K., Kozak, W., Gliszczyńska-Świgło, A., Khmelinskii, I., Górecki, T., ... & Pasqualone, A. (2019). Front-Face Fluorescence Spectroscopy and Chemometrics for **Quality Control of Cold-Pressed Rapeseed Oil** During Storage. *Foods*, 8(12), 665. <https://doi.org/10.3390/foods8120665>
- Wang, M., Zhang, J., Chen, J., Jing, B., Zhang, L., & Yu, X. (2019). Characterization of Differences in **Flavor in Virgin Rapeseed Oils** by Using Gas Chromatography–Mass Spectrometry, Electronic Nose, and Sensory Analysis. *European Journal of Lipid Science and Technology*, 1900205. <https://doi.org/10.1002/ejlt.201900205>
- Jing, B., Guo, R., Wang, M., Zhang, L., & Yu, X. (2020). Influence of seed roasting on the quality of glucosinolate content and **flavor** in virgin rapeseed oil. <https://doi.org/10.1016/j.lwt.2020.109301>
- Zhang, Y., Wu, G., Chang, C., Lv, Y., Lai, W., Zhang, H., ... & Jin, Q. Determination of Origin of Commercial Flavor Rapeseed Oil by the Pattern of **Volatile Compounds** obtained via GC/MS and Flash GC **Electronic Nose**. *European Journal of Lipid Science and Technology*, 1900332. <https://doi.org/10.1002/ejlt.201900332>
- Hou, Z., Cao, X., Cao, L., Ling, G., Yu, Z., Pang, M., ... & Jiang, S. (2020). The **removal of phospholipid** from crude rapeseed oil by enzyme-membrane binding. *Journal of Food Engineering*, 109910. <https://doi.org/10.1016/j.jfoodeng.2020.109910>
- Abdellah, M. H., Scholes, C. A., Liu, L., & Kentish, S. E. (2020). Efficient **degumming** of crude canola oil using **ultrafiltration** membranes and bio-derived solvents. *Innovative Food Science & Emerging Technologies*, 59, 102274. <https://doi.org/10.1016/j.ifset.2019.102274>
- Kasprzak, M., Rudzińska, M., Przybylski, R., Kmiecik, D., Siger, A., & Olejnik, A. (2020). The **degradation of bioactive compounds** and formation of their oxidation derivatives in refined rapeseed oil during **heating** in model system. *LWT*, 123, 109078. <https://doi.org/10.1016/j.lwt.2020.109078>
- Kmiecik, D., Fedko, M., Siger, A., & Kulczyński, B. (2019). Degradation of Tocopherol Molecules and Its Impact on the Polymerization of Triacylglycerols during Heat Treatment of Oil. *Molecules*, 24(24), 4555. <https://doi.org/10.3390/molecules24244555>
- HE, Mogeng et KARATANI, Naohiro. **Oil and fat for suppressing bloom**. U.S. Patent Application No 16/484,114, 23 janv. 2020. <https://patents.google.com/patent/US20200022380A1/en>

- Mao, X., Chen, W., Huyan, Z. et al. Impact of linolenic acid on **oxidative stability** of rapeseed oils. *J Food Sci Technol* (2020). <https://doi.org/10.1007/s13197-020-04349-x>
- Schmid, T., Baumer, B., Rüegg, R., Näf, P., Kinner, M., & Müller, N. (2020). Evaluation of innovative technological approaches to **replace palm fat** with physically modified Swiss rapeseed oil in bakery products. *International Journal of Food Science & Technology*. <https://doi.org/10.1111/ijfs.14564>
- Zewuhn, Merle, Dorothe Fiedler, and Sepideh Reshad. "Lipid mixture of octyldodecanol and **hydrogenated rapeseed oil**." U.S. Patent Application No. 16/491,212. <https://patents.google.com/patent/US20190388333A1/en>
- Hussain, S., Rehman, A. U., Luckett, D. J., Blanchard, C. L., Obied, H. K., & Strappe, P. (2020). **Phenolic Compounds** with Antioxidant Properties from Canola Meal Extracts Inhibit Adipogenesis. *International Journal of Molecular Sciences*, 21(1), 1. <https://doi.org/10.3390/ijms21010001>
- Hao, Y., Wang, Z., Zou, Y., He, R., Ju, X., & Yuan, J. (2020). Effect of Static-State Fermentation on **Volatile Composition in Rapeseed Meal**. *Journal of the Science of Food and Agriculture*. <https://doi.org/10.1002/jsfa.10238>
- Tie, Y., Li, L., Liu, J., Liu, C., Fu, J., Xiao, X., ... & Wang, J. (2020). Two-step biological approach for **treatment of rapeseed meal**. *Journal of Food Science*, 85(2), 340-348. <https://doi.org/10.1111/1750-3841.15011>
- Zhang, Z., Wen, M., & Chang, Y. (2020). Degradation of **glucosinolates** in rapeseed meal by *Lactobacillus delbrueckii* and *Bacillus subtilis*. *Grain & Oil Science and Technology*. <https://doi.org/10.1016/j.gaost.2020.02.003>
- Zhan, Q., Yan, J., Bian, J., Tan, C., Huang, H., & Lin, H. (2020). Study on Ultrasonic Extraction and Stability of **Glucosinolate** in Rapeseed Cake. *E&ES*, 446(2), 022066. <https://iopscience.iop.org/article/10.1088/1755-1315/446/2/022066/meta>
- Herrmann, K. R., Ruff, A. J., & Schwaneberg, U. (2020). Phytase-Based **Phosphorus Recovery** Process for 20 Distinct Press Cakes. *ACS Sustainable Chemistry & Engineering*, 8(9), 3913-3921. <https://doi.org/10.1021/acssuschemeng.9b07433>
- Sánchez-Moya, A., García-Meilán, I., Riera-Heredia, N., Vélez, E. J., Lutfi, E., Fontanillas, R., ... & Navarro, I. (2020). Effects of different dietary vegetable **oils** on growth and intestinal performance, lipid metabolism and flesh quality in **gilthead sea bream**. *Aquaculture*, 519, 734881. <https://doi.org/10.1016/j.aquaculture.2019.734881>
- Gan, L., Feng, L., Jiang, W. D., Wu, P., Liu, Y., Jiang, J., ... & Zhou, X. Q. (2020). **Erucic acid** impairs intestinal immune function of on-growing **grass carp** (*Ctenopharyngodon idella*). *Aquaculture*, 519, 734916. <https://doi.org/10.1016/j.aquaculture.2019.734916>
- Sissener, N. H., Araujo, P., Sæle, Ø., Rosenlund, G., Stubhaug, I., & Sanden, M. (2020). Dietary 18: 2n-6 affects EPA (20: 5n-3) and ARA (20: 4n-6) content in cell membranes and eicosanoid production in **Atlantic salmon** (*Salmo salar* L.). *Aquaculture*, 522, 735098. <https://doi.org/10.1016/j.aquaculture.2020.735098>
- Kaczmarek, S. A., Hejdysz, M., Kubiś, M., Nowaczewski, S., Mikuła, R., & Rutkowski, A. (2019). Effects of feeding intact, ground and/or pelleted rapeseed on nutrient digestibility and growth performance of **broiler chickens**. *Archives of Animal Nutrition*, 1-15. <https://doi.org/10.1080/1745039X.2019.1688557>
- Kuźniacka, J., Biesek, J., Banaszak, M., Rutkowski, A., Kaczmarek, S., Adamski, M., & Hejdysz, M. (2020). Effect of Dietary Protein Sources Substituting Soybean Meal on Growth Performance and Meat Quality in **Ducks**. *Animals*, 10(1), 133. <https://doi.org/10.3390/ani10010133>

- Kawabata, F., Dey, B., Yoshida, Y., Nishimura, S., & Tabata, S. (2019). Bitter Taste Receptor Antagonists Inhibit the Bitter taste of Canola Meal Extract in **Chickens**. *The Journal of Poultry Science*, 0190099. <https://doi.org/10.2141/jpsa.0190099>
- Hanna, C., Munoz, J., Utterback, P., & Parsons, C. M. (2020). Research Note: **Phosphorus digestibility** in conventional canola meal determined using different balance assays. *Poultry Science*. <https://doi.org/10.1016/j.psj.2019.12.057>
- Kowalska, D., Strychalski, J., Zwoliński, C., Gugolek, A., & MATUSEVICIUS, P. (2020). The Effect of Mixture of Rapeseed Meal, White Lupin Seed, and Pea Seed in **Rabbit** Diets on Performance Indicators and Fatty Acid Profile of Meat and Fat. *Kafkas Universitesi Veteriner Fakultesi Dergisi*. Kars: Kafkas Univ, Veteriner Fakultesi Dergisi, 2020, vol. 00, no. 00. http://www.vetdergikafkas.org/uploads/pdf/pdf_KVFD_2694.pdf or <https://doi.org/10.9775/kvfd.2019.23222>
- Grabež, V., Egelanddal, B., Kjos, N. P., Håkenåsen, I. M., Mydland, L. T., Vik, J. O., ... & Øverland, M. (2020). Replacing soybean meal with rapeseed meal and faba beans in a **growing-finishing pig** diet: Effect on growth performance, meat quality and metabolite changes. *Meat Science*, 108134. <https://doi.org/10.1016/j.meatsci.2020.108134>
- Vergara, D., López, O., Bustamante, M., & Shene, C. (2020). An in vitro digestion study of encapsulated lactoferrin in rapeseed phospholipid- based liposomes. *Food Chemistry*, 126717. <https://doi.org/10.1016/j.foodchem.2020.126717>
- Mohammadi, M., Imani, A., Farhangi, M., Gharaei, A., & Hafezieh, M. (2020). Replacement of fishmeal with processed canola meal in diets for juvenile **Nile tilapia** (*Oreochromis niloticus*): Growth performance, mucosal innate immunity, hepatic oxidative status, liver, and intestine histology. *Aquaculture*, 518, 734824. <https://doi.org/10.1016/j.aquaculture.2019.734824>
- Paula, E. M., Broderick, G. A., & Faciola, A. P. (2020). Effects of replacing soybean meal with canola meal for lactating **dairy cows** fed 3 different ratios of alfalfa to corn silage. *Journal of Dairy Science*, 103(2), 1463-1471. <https://doi.org/10.3168/jds.2019-16947>
- Vatandoost, M., & Didarkhah, M. Effect of Different Levels of Soybean Oil, Soybean and Canola Meal on Production Performance, Rumination Activity and Nutrient Digestibility in Holstein **Dairy Cows**. *Research On Animal Production (Scientific and Research)*, 0-0. http://rap.sanru.ac.ir/browse.php?a_id=1008&sid=1&slc_lang=en
- Lee, J. W. (2019). Optimization of Canola Co-product Utilization in **Swine**. PhD thesis South Dakota State University : <https://openprairie.sdstate.edu/etd/3654/>
- Brock, D., Koder, A., Rabl, H. P., Touraud, D., & Kunz, W. (2020). Optimising the **biodiesel** production process: Implementation of glycerol derivatives into biofuel formulations and their potential to form hydrofuels. *Fuel*, 264, 116695. <https://doi.org/10.1016/j.fuel.2019.116695>
- Likhanov, V. A., Lopatin, O. P., & Yurlov, A. S. (2020, January). **Biofuel** based on methanol and methyl ester of rapeseed oil for diesel engine. In *IOP Conference Series: Materials Science and Engineering* (Vol. 734, No. 1, p. 012208). IOP Publishing. <https://iopscience.iop.org/article/10.1088/1757-899X/734/1/012208/meta>
- Zieniuk, B., Wołoszynowska, M., & Białecka-Florjańczyk, E. (2020). Enzymatic Synthesis of **Biodiesel** by Direct Transesterification of Rapeseed Cake. *International Journal of Food Engineering*, 16(3). <https://doi.org/10.1515/ijfe-2019-0089>
- Encinar, J. M., Nogales-Delgado, S., Sánchez, N., & González, J. F. (2020). Biolubricants from Rapeseed and Castor Oil **Transesterification** by Using Titanium Isopropoxide as a Catalyst: Production and Characterization. *Catalysts*, 10(4), 366. <https://doi.org/10.3390/catal10040366>

- Tan, L., Zhong, J., Jin, Y. L., Sun, Z. Y., Tang, Y. Q., & Kida, K. (2020). Production of **bioethanol** from unwashed-pretreated rapeseed straw at high solid loading. *Bioresource Technology*, 303, 122949. <https://doi.org/10.1016/j.biortech.2020.122949>
- Xu, C., Xia, T., Wang, J. et al. Selectively Desirable Rapeseed and Corn Stalks Distinctive for Low-Cost **Bioethanol** Production and High-Active Biosorbents. *Waste Biomass Valor* (2020). <https://doi.org/10.1007/s12649-020-01026-0>
- Hýsková, P., Hýsek, Š., Schönfelder, O., Šedivka, P., Lexa, M., & Jarský, V. (2020). Utilization of agricultural rests: Straw-based **composite panels** made from enzymatic modified wheat and rapeseed straw. *Industrial Crops and Products*, 144, 112067. <https://doi.org/10.1016/j.indcrop.2019.112067>
- Sheridan, J., Sonebi, M., Taylor, S., & Amziane, S. (2020). The effect of long-term weathering on hemp and **rapeseed concrete**. *Cement and Concrete Research*, 131, 106014. <https://doi.org/10.1016/j.cemconres.2020.106014>
- Moreno-González, M., Girish, V., Keulen, D., Wijngaard, H., Lauteslager, X., Ferreira, G., & Ottens, M. (2020). Recovery of **sinapic acid** from canola/rapeseed meal extracts by adsorption. *Food and Bioproducts Processing*, 120, 69-79. <https://doi.org/10.1016/j.fbp.2019.12.002>
- Cong, Y., Zheng, M., Huang, F., Liu, C., & Zheng, C. (2020). **Sinapic acid** derivatives in microwave-pretreated rapeseeds and minor components in oils. *Journal of Food Composition and Analysis*, 87, 103394. <https://doi.org/10.1016/j.jfca.2019.103394>
- Fetzer, A., Hintermayr, C., Schmid, M. et al. Effect of Acylation of **Rapeseed Proteins** with Lauroyl and Oleoyl Chloride on Solubility and **Film-Forming** Properties. *Waste Biomass Valor* (2020). <https://doi.org/10.1007/s12649-020-01012-6>
- Fitzpatrick, S.E., Deb-Choudhury, S., Ranford, S. et al. Novel **protein-based bio-aerogels** derived from canola seed meal. *J Mater Sci* 55, 4848–4863 (2020). <https://doi.org/10.1007/s10853-019-04330-w>
- Ivdre, A., Abolins, A., Sevastyanova, I., Kirpluks, M., Cabulis, U., & Merijs-Meri, R. (2020). Rigid Polyurethane Foams with Various Isocyanate Indices Based on **Polyols** from Rapeseed Oil and Waste PET. *Polymers*, 12(4), 738. <https://doi.org/10.3390/polym12040738>
- Chen, W., Li, X., Ma, X. et al. Simultaneous hydrolysis with lipase and fermentation of rapeseed cake for **iturin A production** by *Bacillus amyloliquefaciens* CX-20. *BMC Biotechnol* 19, 98 (2019). <https://doi.org/10.1186/s12896-019-0591-x>
- Yu, G., Guo, T., & Huang, Q. (2020). Preparation of rapeseed oil with superhigh **canolol** content and superior quality characteristics by steam explosion pretreatment technology. *Food Science & Nutrition*. <https://doi.org/10.1002/fsn3.1502>

ECONOMY and MARKET

- Deepayan Debnath, Jarrett Whistance, Patrick Westhoff, Mike Helmar, Chapter 9 - Consequences of **US and EU biodiesel policies** on global food security. Editor(s): Deepayan Debnath, Suresh Chandra Babu. *Biofuels, Bioenergy and Food Security*, Academic Press, 2019, <https://doi.org/10.1016/B978-0-12-803954-0.00009-7>
- Declerck, Francis and Indjehagopian, Jean-Pierre and Lantz, Frederic, Dynamics of Biofuel Prices on the European Market: Impact of the EU Environmental Policy on the Resources Markets (February 21, 2020). Available at SSRN: <https://ssrn.com/abstract=3542376> or <http://dx.doi.org/10.2139/ssrn.3542376>

- Oleynikova, E. N., Yanova, M. A., Sharopatova, A. V., & Grishina, I. I. (2020, January). Comparative evaluation of the economic efficiency of the rapeseed cultivation by the traditional method and using the principles of **organic production**. In IOP Conference Series: Earth and Environmental Science (Vol. 421, No. 3, p. 032005). IOP Publishing. <https://iopscience.iop.org/article/10.1088/1755-1315/421/3/032005/meta>
- PREPRINT: Meijaard, E., Brooks, T., Carlson, K., Slade, E. M., Ulloa, J. G., Gaveau, D. L., ... & Ancrenaz, M. (2020). The environmental **impacts of palm oil in context**. <https://eartharxiv.org/e69bz/>
- Beyer, R. M., Duran, A. P., Rademacher, T. T., Martin, P., Tayleur, C., Brooks, S. E., ... & Sanderson, F. J. (2020). The environmental **impacts of palm oil and its alternatives**. bioRxiv. <https://doi.org/10.1101/2020.02.16.951301>
- Tao, J., Wu, W., Liu, W., & Xu, M. (2020). Exploring the Spatio-Temporal Dynamics of Winter Rape on the Middle Reaches of **Yangtze River Valley** Using Time-Series MODIS Data. Sustainability, 12(2), 466. <https://doi.org/10.3390/su12020466>
- Kaliakparova, G. S., Gridneva, Y. E., Alpysbayev, K. S., & Sevindik, T. (2020). Market of repae plant: condition and prospects for **Kazakhstan**. ҚАЗАҚСТАН РЕСПУБЛИКАСЫ, 80. <http://rmebrk.kz/journals/5922/16870.pdf#page=80>
- Goncharov, S. V., & Shokhina, L. V. (2020, March). How to **Share Value in Seed Market?**. In International Scientific Conference " Far East Con"(ISCFEC 2020) (pp. 1803-1807). Atlantis Press. <https://doi.org/10.2991/aebmr.k.200312.250>
- Reed, C. (2019). The Exploratory Study of the Effects of **Selling Canola by Seed Size** on Farmers: Variability of Hybrids and Years. <https://lib.dr.iastate.edu/creativecomponents/420/>

MUSTARD and Other Brassicae

- Karpavičienė, Birutė, Nijolė Maršalkienė, and Liuda Žilėnaitė. "Seed composition of different *Camelina sativa* and *Crambe abyssinica* cultivars." 26th NJF Congress: Agriculture for the Next 100 Years, 27-29 June, 2018 Kaunas r. Lithuania: Programme and Summaries of Presentations. Akademija, 2018. 2018. <https://hdl.handle.net/20.500.12259/92459>
- Salas, H., Castillejos, L., López-Suárez, M., & Ferret, A. (2019). In Vitro Digestibility, In Situ Degradability, Rumen Fermentation and N Metabolism of **Camelina** Co-Products for Beef Cattle Studied with a Dual Flow Continuous Culture System. Animals, 9(12), 1079. <https://doi.org/10.3390/ani9121079>
- Jankowski, K. J., Załuski, D., & Sokólski, M. (2020). Canola-quality **white mustard**: Agronomic management and seed yield. Industrial Crops and Products, 145, 112138. <https://doi.org/10.1016/j.indcrop.2020.112138> or [REFERENCE](#)
- Geller, D., Mulvaney, M. J., Seepaul, R., Small, I., Wright, D., Paula-Moraes, S., ... & Leon, R. Frost Damage of **Carinata**. https://secure.caes.uga.edu/extension/publications/files/pdf/B%201517_4.PDF

Upcoming International and national events

September 24-27, 2023 16th International Rapeseed Congress, Sydney, Australia

www.irc2023sydney.com



We invite you to share information with the rapeseed/canola community: let us know the scientific projects, events organized in your country, crop performances or any information of interest in rapeseed/canola R&D.

Contact GCIRC News:

Etienne Pilorgé, GCIRC Secretary-Treasurer: e.pilorge@terresinovia.fr

Contact GCIRC: contact@gcirc.org