



## Global Council for Innovation in Rapeseed and Canola

*“Building a World community for Innovation on Rapeseed and Canola”*

*N° 14, February 2023*

### Table des matières

<b>Editorial</b> .....	2
<b>Activity/ News of the association:</b> .....	3
<b>IRC-16 Sydney 2023 – September 24-27, Australia</b> .....	3
<b>Professor Jan Krzymaniński</b> .....	5
<b>Welcome to New GCIRC members</b> .....	7
<b>Value chains and regional news</b> .....	7
<b>Global rapeseed market: UFOP publishes updated Report on Global Market Supply 2022/2023</b> .....	8
<b>Australian Canola Crop Summary</b> .....	8
<b>European Union: mutagenesis and GMO</b> .....	9
<b>Highlights from Canada’s Canola Week</b> .....	10
<b>India</b> .....	11
<b>International Conference on Vegetable Oils (ICVO) 2023 at Hyderabad, India</b> .....	12
<b>Scientific news</b> .....	13
<b>Rapeseed research through a bibliometric study</b> .....	13
<b>Bioengineering to increase the yield of vegetable oil from plants</b> .....	13

<b>Publications:</b> .....	13
<b>GENETICS &amp; BREEDING</b> .....	13
<b>CROP PROTECTION</b> .....	18
<b>AGRONOMY &amp; CROP MANAGEMENT</b> .....	21
<b>PHYSIOLOGY</b> .....	24
<b>REMOTE SENSING</b> .....	26
<b>PROCESSING, QUALITY &amp; PRODUCTS</b> .....	27
<b>NUTRITION AND HEALTH</b> .....	33
<b>ANALYZES</b> .....	34
<b>ECONOMY and MARKET</b> .....	34
<b>MUSTARD and Other Brassicae</b> .....	35
<b>MISCELLANEOUS</b> .....	35
<b>Upcoming international and national events</b> .....	35

## Editorial

Greetings and welcome to GCIRC Newsletter #14, February 2023.

Welcome to our first newsletter for 2023, and begin by expressing how extremely important it is to remember the people of Ukraine on this the 1<sup>st</sup> anniversary of the war. It is hard to comprehend the devastation, destruction to cities and vital infrastructure let alone the injuries and loss of so many lives. Our thoughts are with our colleagues in Ukraine who are resiliently maintaining supply chains for oilseeds and veg oils under the most challenging conditions in that region.

Sadly, the Board has been advised of the recent passing of Prof Krzymanski. To his family, friends, and colleagues, we the Board and members of GCIRC pass on our sincere condolences.

Prof Krzymanski was a highly regarded scientist, who in 2003 received a GCIRC Rapeseed Award for his lifelong achievements including the attached paper ‘Rapeseed breeding for better oil and meal quality in Poland’.

With the ever-changing weather conditions globally, whether it be drought in one region or floods in another oilseed production will be monitored closely in 2023. That said I look forward to reviewing country crop forecasts, both area and production numbers in this newsletter.

At the February 13<sup>th</sup> GCIRC board meeting, discussion about the next technical meeting was held with a call for application submissions to host the 2025 Technical meeting be available for voting at the

Sydney board meeting. India and the US have indicated they are interested, so look forward to hearing more shortly.

Finally, remember to register for IRC-16 in Sydney where I look forward to seeing as many of you as possible in September.

Robert Wilson, GCIRC President

## Activity/ News of the association:

### IRC-16 Sydney 2023 – September 24-27, Australia



How time flies - only 30 weeks until IRC-16 Congress and already registrations are flowing in. Many regular IRC attendees are taking advantage of the early bird rate and have already registered, which is particularly encouraging. The Call for Abstracts opened this week, we look forward to these also starting to flow in as well in the lead up to the Congress.

All details for registration, abstract submission and draft program are available on the <https://www.ircsydney2023.com/> website. We are also very pleased to report that sponsorship support has been very strong with many of the high level, high exposure sponsorship opportunities already taken. BASF is the sole Diamond sponsor; Syngenta has taken a Gold sponsorship, while Nuseed has agreed to sponsor the Congress dinner. All sponsors are listed on the website, and there are still many more sponsorship opportunities available. Contact the organisers at [ircsydney2023@australia-noilseeds.com](mailto:ircsydney2023@australia-noilseeds.com) if you are interested in gaining exposure for your business or research institution through adding your name to the list of prestigious sponsors.

The Congress theme of *'Global Crop – Golden Opportunities'* will recognise and highlight the outstanding opportunities canola/rapeseed and the scientific committees are already locking in keynote and plenary speakers, and these will be notified progressively on the website as they are secured in the weeks ahead. Thank you to those of our international colleagues who have agreed to participate on a committee.

The Congress scientific program will run over 3 days and organised under 6 Core Themes:

- Genetics, Genomics and Breeding
- Agronomy, Physiology and Simulation
- Crop Protection

- Products and Quality
- End Uses
- Economy and Markets

The Themes will run as Concurrent Tracks with opportunities for up to 40 contributed talks in each Track over the 3 days.

Each day will commence with a plenary session of invited keynote talks of general interest to all delegates presented by global leaders in their fields.

The Congress Organising Committee is engaging the Keynote Speakers, and these will be notified progressively on the website as they are secured in the weeks ahead.

Preceding the Congress will be our 3-day Field tour, beginning Friday 22<sup>nd</sup>, visiting one of Australia's leading centres of canola breeding and research at Wagga Wagga in the heart of canola country.

The Tour will include visits to research trials, Australia's newest oilseed crushing plant, a live sheep shearing demonstration, plus more!

The Tour also includes return transfers to Sydney beginning Saturday traveling via the nation's capital Canberra en route, arriving back in Sydney on Sunday afternoon, in time for some sight-seeing or the pre-arranged Clubroot workshop or the GCIRC board meeting or some rest before the Welcome Reception!

We will be asking participating delegates to make their own way to Wagga Wagga to commence the Field Tour, as most international air tickets can include a more cost-effective add-on flight to Wagga Wagga. Bus and train options are also available, and details will be provided to assist with your logistics.

Call for Abstracts: Opened last week (21<sup>st</sup> February) – watch your inbox for notifications.

Pre-congress Field Tour Committee – Core members engaged, planning underway.

When booking your registration remember to also book the field tour which is separate and not included in the Congress registration and finally also arrange the necessary arrival documents such as your Visa to enter Australia.

We look forward to welcoming as many friends and colleagues as possible to Australia in September 2023. Remember, Sydney – ***“it's closer than you think”***.

For further info go to <https://www.ircsydney2023.com>

Congress Executive Progress report - meeting regularly, hitting timelines for activities.

- ✓ Congress venue booked.
- ✓ Congress Dinner venue booked.
- ✓ Key major Sponsors secured along with supporting sponsors.
- ✓ Registration opened late 2022 already have good uptake on Early-Bird offer.
- ✓ Budget (ongoing)

- ✓ Plenary and Keynote speakers being assembled.
- ✓ Call for Abstracts opened late February.
- ✓ Pre-congress field tour finalised, and activities started.
- ✓ Website live.
  - <https://www.ircsydney2023.com> & [IRC Program/Itinerary](#)

## Professor Jan Krzymański

We are deeply saddened to inform of the passing of Prof Jan Krzymański, and express our sympathy to his family, friends, and colleagues.

Prof Jan Krzymański greatly contributed to rapeseed research and development. His involvement and the excellence of his work were recognised through the Rapeseed Award in 2003, in Copenhagen.



*Prof Jan Krzymański receiving the Award during the 11<sup>th</sup> IRC in Copenhagen, in 2003.*

Prof Jan Krzymański was one of the first members of the GCIRC, having joined the association in 1978 (the GCIRC constitution was adopted in November 1977, after 5 years of discussions) and participated actively to the GCIRC Board until 2005. He was president of GCIRC and chaired the 7<sup>th</sup> IRC in Poznan, in 1987.

He played a key role in the elaboration of the first varieties of low erucic and low glucosinolates winter rapeseed, at the basis of cultivars grown in Europe until today.

We give here a summary of his history, extract from an article written by Prof Krzymański himself for the 4<sup>th</sup> IRC, in Giessen, Germany, in 1974. It illustrates his personal role and also the importance of international collaborations, and the progress of knowledge on rapeseed genetics in the 1960<sup>ies</sup> and 70<sup>ies</sup>:

<<The lower seed value of rape is determined mainly by two factors – high erucic acid content in oil and toxic properties of thioglucosides (glucosinolates) which occur in meal. These two undesirable factors can be changed only in small degree by modifications in oil industry technology. It looks now the best solution of the problem can be obtained with genetical means breeding of new varieties of rape with improved chemical composition. Especial research project was made [in Poznan, Poland] by Oil Crop Department of Institute of Plant Breeding and Acclimatization (IHAR) .

Elaboration of method for fatty acid composition analysis by quantitative paper chromatography (Krzymański, 1961, 1965) allowed to undertake genetical researches and rape breeding for low erucic acid content in seed oil. This method was replaced later by gas chromatography (Byczynska & Krzynański, 1968, Krzynański and Downey, 1969). Works on thioglucoside required also certain and proper analytical method. Different methods were examined and a new modification of Youngs-Wetter's methods were proposed (Byczynska, 1974).

This method , based on gas chromatography, makes possible the estimation of individual isothiocyanates and individual oxazolidinethiones in seed meal. It is well adapted for needs of breeding and genetic investigation. Oil content in seeds is not analyzed now by destructive and quick method based on nuclear magnetic resonance measurement (Krzynański, 1970).

Survey of all varieties and strains of winter rape in our collection showed that we had none winter form low in erucic acid or thioglucoside content (Byczynska 1974, Krzynański 1965). For this reason, it was necessary to use spring forms for obtaining essential genetic variability. The following lines were genetical sources of desired traits in our researches and breeding works:

1. Zero erucic line selected from 'Liho' variety of spring rape in Canada (Stephansson et al., 1961)
2. Low erucic line selected from "Bronowski" variety of spring rape in Poland (Krzynański, 1966, Krzynański et al 1967)
3. Lines with very low thioglucoside content selected from "Bronowski" variety of spring rape (Finlayson et al, 1973; Krzynański, 1970)

Investigation was realized on inheritance of erucic acid content in rapeseed oil (Krzynański et al 1967; Krzynański & Downey, 1969; Krzynański 1970) and on inheritance of thioglucoside content in rape seed (Krzynański et al 1970). The results obtained were conformable to the published data of other authors (Harvey & Downey, 1964; Kondra & Stefansson 1965, 1970) (based on these results it can be concluded:

1. Erucic acid content in rape seed oil is a hereditary trait controlled by embryo genotype;
2. Erucic acid content is controlled by one or two pair system – the zero or low erucic forms represents  $\frac{1}{4}$  or  $\frac{1}{16}$  of the F2 generation depending on cross combination;
3. There are alleles or pseudoalleles controlling different levels of eruci acid acting in an additive manner without distinct domination;

4. *Thioglucosides content in rapeseed is controlled mainly by the maternal plant genotype in respect of both quantity and quality;*
5. *The trait of high thioglucoside content is a dominant one in reference to total content of these compounds, but for individual thioglucosides different results were obtained. Overdominancy was observed in the case of pentenyl isothiocyanate, dominance for butenyl isothiocyanate and incomplete dominance for oxazolidinethiones.;*
6. *A differentiation in thioglucosides composition was also observed in segregating generations of hybrids.*

*Low erucic or low thioglucosides strains of winter rape were obtained by crosses between winter varieties of rape and above-mentioned lines of spring rape. But these desired traits were strongly linked with many other traits typical for spring forms. These traits were usually unfavourable and caused the new strains had much lower agricultural value than normal old varieties which have been cultivated till now. These strains make only a raw material which needs further improvement especially for better vigour, higher yielding ability and better winter hardiness. We try to achieve this goal by using breeding methods based on backcrossing and intensive selection in segregating conditions... >>*

The full article and many others of that time are available on the GCIRC website (<https://www.gcirc.org/publications/archives/irc-proceedings-until-2015> )

## Welcome to New GCIRC members

Since last October we have welcomed four new members:

ZHANG Liangxiao	Chinese Academy of Agricultural Sciences	CHINA
HILTPOLD Ivan	AGROSCOPE	SWITZERLAND
GREEN Allan	AGRENEW Pty Ltd	AUSTRALIA
FALAK Igor	Corteva	CANADA

*You may visit their personal pages on the GCIRC website directory, to better know their fields of interest. We take this opportunity to remind all members that they can modify their personal page, especially indicating their fields of interest in order to facilitate interactions.*

## Value chains and regional news

## Global rapeseed market: UFOP publishes updated Report on Global Market Supply 2022/2023

“In more than 50 pages, the UFOP Report on Global Market Supply provides detailed information on the current state of global grain, oilseed and vegetable oil production, main uses in human and animal nutrition and use as a renewables feedstock in biofuels production.” See: Ufop news:

<https://www.ufop.de/english/news/ufop-publishes-updated-report-global-market-supply-20222023/> and the report available to download at: [https://www.ufop.de/index.php/download\\_file/12108/](https://www.ufop.de/index.php/download_file/12108/)

## Australian Canola Crop Summary

### **2022 Season:**

The season surprised even the most experienced pundits, with another record crop produced for Australia of close to 7.8mmt. This result was despite very wet conditions in the states of NSW and Victoria with heavy rain and floods affecting many crops towards the end of the season. Resilient crops, with maturing seeds pods were able to withstand these conditions in most cases. Very large biomass created challenges for harvest, as many fields were too wet to access for swathing/ windrowing forcing farmers to resort to direct heading/harvesting, which is unusual in Australia.

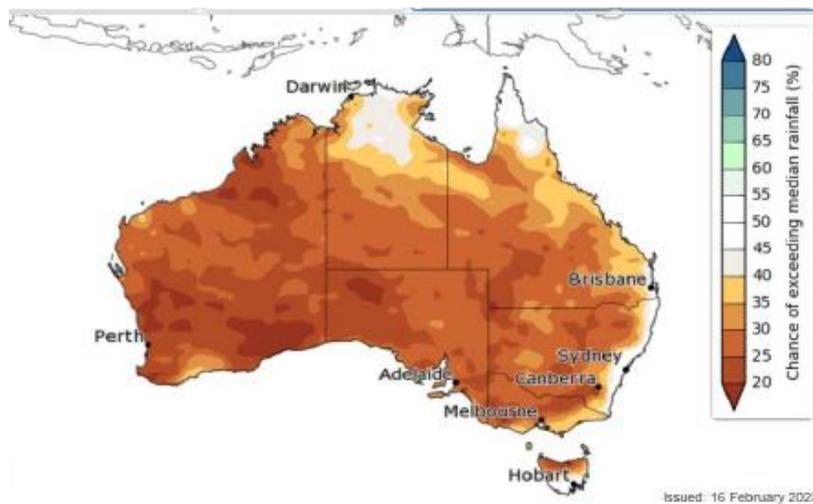
In the other canola producing states of South Australia and Western Australia, conditions with ideal, with record crops recorded on both those states. Western Australia, at 4.25mmt achieved what would be regarded only a few years ago as a good national crop.

2022/23	AREA	YIELD	PROD'N
NSW	856	1.76	1,507
VIC	630	2.17	1,367
SA	307	2.14	657
WA	1,986	2.14	4,250
AUS	3,779	2.06	7,781

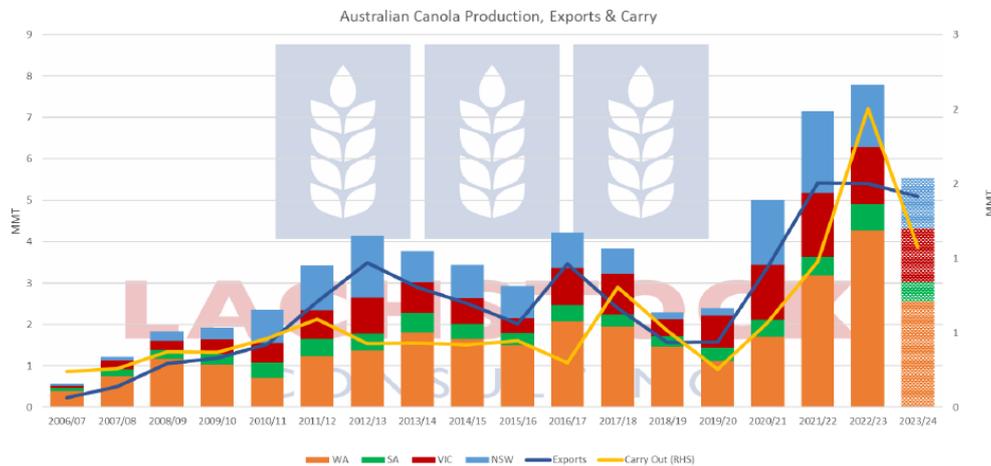
### **2023 Season:**

The upcoming season is expected to see a return to the longer-term trend in terms of area to be sown (in April) and expected yield. While there is good subsoil moisture throughout the Eastern states of NSW and Victoria, and no indication Western Australia won't receive its usual May seasonal break (rain), the overall trend is towards El Niño conditions, which would tend to result in below-average rainfall. Heavy canola rotations over the last 2 years, combined with a softening price and still relatively high input costs will also temper grower enthusiasm to go very strong on canola.

The forecast is for only 20-35% chance of exceeding median rainfall during the seeding and establishment phase, (April-June) which will mean that the crops will rely on accessing deeper soil moisture during the critical biomass production stage leading up to flowering. If El Niño conditions continue through to pod and seed development, yields and oil content will be impacted.



Early indications are for an area sown to be down around 10% on last year with yields closer to the national long-term average of around 1.6t/ha. This will deliver a national production volume of 5.5mmt, with as much as 2 million tonnes carry over from the last harvest.



## European Union: mutagenesis and GMO

In a ruling published on 7 February, the Court of Justice of the European Union ruled that organisms obtained by "in vitro" random mutagenesis are excluded from the scope of the 2001 Directive on the deliberate release of GMOs or their placing on the market.

For more information: the original text of the Court of Justice at <https://curia.europa.eu/jcms/upload/docs/application/pdf/2018-07/cp180111en.pdf>

And for comments on context: <https://www.euractiv.com/section/agriculture-food/news/eu-court-exempts-gene-modification-technique-from-stricter-rules/>

This decision comes as the European Commission plans to clarify the regulation of new genomic techniques (directed mutagenesis, intragenesis and cisgenesis). It is expected to present a draft regulation on new GMOs by the end of the first quarter of 2023.

## Highlights from Canada's Canola Week

The Canola Council of Canada co-hosted Canola Week December 6-9, 2022, in Saskatoon, Saskatchewan, host city of the 2015 International Rapeseed Congress. Here are a few market opportunities, productivity enhancements and new technologies.

By Jay Whetter

**The aquaculture opportunity for canola meal.** Data from the Food and Agriculture Organization of the United Nations show wild caught and farmed fish neck and neck in terms of supply in 2020, with aquaculture trending upward quickly and wild caught trending downward. Brittany Wood, director of communications for the Canola Council of Canada, says aquaculture – fish farming – “plays to canola meal’s strengths.” Carp, catfish, tilapia, and salmonids are the top four farmed fish. Shrimp are also farmed in large quantities. Standard canola meal is a great fit for tilapia and carp farms, common in China. Specialty canola meal, like the high-protein, low-fibre product from Botaneco, a Canadian company, works for farmed salmon and shrimp.

**Ag and petroleum companies partner on renewable diesel.** In Canada, Saskatchewan-based food company AGT is working with Federated Co-op on a renewable diesel and canola protein meal project. This is just one example of new relationships forming between agriculture companies and petroleum companies. In the U.S., ADM food company and Marathon fuel company have a joint project in North Dakota, Bunge and Chevron are working together in Louisiana and Illinois, and Shell Rock Soy Processing and P66 in Iowa.

**Fix or set aside unprofitable acres?** Land is not going to fix itself, says Jason Casselman, Canola Council of Canada agronomy specialist. “With data analysis and mapping technology, farmers now have the opportunity to not only identify areas of low productivity, but also see how deep the problem is when they look at the bottom line,” Casselman says.

–**Fix: Moving topsoil back to hilltops.** Marla Riekman, soil management specialist with Manitoba Agriculture, cites Manitoba studies showing that topsoil added back to hilltops increases yields significantly, while having minimal effect on yield in low-lying areas where soil was removed.

–**Fix: Strategic tile drainage.** Tile drainage removes water that exceeds the holding capacity of the soil. This excess water impedes root function and limits field activities. By removing this water, tile drainage can improve plant health and plant uniformity and allow farmers to get on fields faster in the spring or after a big rain.

–**Set aside: Convert unfixable acres to grass.** Mark McConnell, assistant professor and upland birds’ specialist at Mississippi State University, uses field profit maps to show chronically unprofitable areas.

Given the shape of these areas, it may not be practical to take them all out of production, but he says it can make sense to set aside some field edges with grasses and forages. In a published research paper, McConnell wrote: “I suggest targeted conservation be defined as the application of conservation practices only where they increase profitability to the producer.”

**Technology that excites.** Joy Agnew moderated a Canola Discovery Forum panel on precision agriculture technology. Agnew is associate vice president, applied research, at Olds College of Agriculture & Technology in Olds, Alberta. She asked her three panelists, what upcoming technology most excites you?

–**Bonnie Mandziak, product marketing manager with Climate FieldView:** “If we can use data and digital tools to help farmers answer spray questions – Do I spray? When should I spray? And where should I spray? – we can help them make better more informed decisions.”

–**Christian Hansen, small grains corporate agronomist with John Deere:** “I’m excited for Innerplant, which is a company inserting fluorescent proteins into plants that can make them signal certain stressors throughout their life cycle. While Innerplant is at a very early stage, the commercial application of this tech is limitless to help agronomists and growers make proactive decisions on their farms. It could be used to signal fields that are at high risk for disease infection, insect infestations, nutrient deficiencies or even water stress.”

–**Garth Donald, manager of agronomy with Decisive Farming by Telus Agriculture:** “Hyperspectral imaging. With this technology, one will be able to identify plant diseases before the human eye can see them. That way one can be more proactive than reactive.” Hyperspectral imaging captures wavelengths beyond visible light to show things the eye can’t see. Low earth orbit satellites, once launched, will capture these high-resolution images at a broad regional scale.

(Jay Whetter is the editor of Canola Digest. Read the magazine online at [canoladigest.ca](http://canoladigest.ca). Read his insightful column at [canoladigest.ca/department/the-editors-desk/](http://canoladigest.ca/department/the-editors-desk/))

## India

India is the fourth-largest contributor of oilseeds in the world, Indian rapeseed and mustard contribute for about 28.6% of total oilseeds production. Rapeseed–mustard crops in India are grown under diverse agro climatic conditions, e.g. north-eastern / north western hills to down south under irrigated/rainfed, timely/late sown, saline soils and mixed cropping. During 2021-22, the all-time highest production of 11.75 MT was 13 percent higher than the one of 2020-21 year (10.21MT). Area under mustard in the current post-rainy (rabi) season has been reported at a record 9.4 million hectare (MH) which is 49% more than last five years’ average sown area of 6.3 MH.

## International Conference on Vegetable Oils (ICVO) 2023 at Hyderabad, India

Indian Council of Agricultural Research, ICAR-Indian Institute of Oilseeds Research (IIOR), and Indian Society of Oilseeds Research (ISOR) in collaboration other ICAR oilseed institutes and the societies engaged with vegetable oil research were jointly organized the International Conference on Vegetable Oils (ICVO) 2023 during January 17- 21, 2023 at Hyderabad, India. The conference was envisaged to be a convergent point for priority persuasion and provided a platform to deliberate on research strategies, infrastructure developmental needs, trade and value chain ecosystems, and policy perspective to promote increased vegetable oil production on short-, medium- and long-term basis at global as well as national levels. Several invited talks, plenary talks, contributory oral as well as poster presentations and technology exhibitions were organised during the International conference. Also, five satellite symposia dedicated to specific issues of major vegetable oil crops were held during the conference, including Satellite symposium on Rapeseed-mustard.



Inauguration of ICVO 2023

During satellite symposium on Rapeseed-mustard, Dr. PK Rai Director, ICAR-DRMR, Bharatpur, presented the current status and future development strategies in rapeseed-mustard for nutritional security. Dr. SR Bhat presented a talk on Pre-breeding for genetic enhancement of oilseed Brassica. Dr. Etienne Pilorgé, Terres Inovia, and Secretary GCIRC, France, made a presentation on integrated management to enhance productivity of rapeseed, through online mode. Dr. HC Sharma made presentation on new paradigms in insect-pest management in oilseed Brassica. Dr. Samantha Cook, Rothamsted Research, UK made a talk on IPM strategies for insect pests in European rapeseed, through online mode. Five presentations were made on different aspects by Drs. VV Singh, Pankaj Sharma, AK Sharma, RS Jat and H.K. Sharma, ICAR-DRMR. Symposium was chaired by Dr. Arvind Kumar, co-chaired by Dr. P.K. Rai and coordinated by Dr. Pankaj Sharma. Later a panel discussion was also held under chairmanship of Dr. S.R. Bhat.

## Scientific news

### Rapeseed research through a bibliometric study

An interesting bibliometric study on rapeseed research, showing the main countries making efforts on rapeseed, in which names of several GCIRC members appear, and many others that GCIRC would welcome with pleasure.

Zheng, Q., & Liu, K. (2022). Worldwide **rapeseed** (*Brassica napus* L.) **research: A bibliometric analysis** during 2011–2021. *Oil Crop Science*, 7(4), 157-165. <https://doi.org/10.1016/j.ocsci.2022.11.004>

### Bioengineering to increase the yield of vegetable oil from plants

In November 2022, the ScienceDaily website (University of Singapore) reported results showing, in the laboratory, the possibility to increase the yield of oil production by a plant. This method is patent pending. Scientists have successfully bioengineered an important protein in plants to increase the yield of oil from their fruits and seeds -- a holy grail for the global agri-food industry. Their patent-pending method can increase oil content in seeds by 15 to 18 per cent, which is a significant improvement that could be applied to numerous oilseeds. This innovation can help the world in its quest for sustainability, helping to reduce the amount of arable land needed for oil-yielding crops while increasing the oil yield to meet the world's growing demand for vegetable oil.

Read more at <https://www.sciencedaily.com/releases/2022/11/221109124301.htm> and original article at <https://doi.org/10.1126/sciadv.abq1211>

### Publications:

**To the authors: we identify publications through research with 2 key words only: “rapeseed” and “canola”. If a publication does not contain one of these two words, but for example only *Brassica napus* or terms implicitly linked to rapeseed/canola (for example names of diseases or insects or genes, etc....), it will not be detected.**

### GENETICS & BREEDING

Orantes-Bonilla M, Makhoul M, Lee H, Chawla HS, Vollrath P, Langstroff A, Sedlazeck FJ, Zou J and Snowdon RJ (2022) Frequent spontaneous structural rearrangements promote rapid **genome diversification** in a *Brassica napus* F1 generation. *Front. Plant Sci.* 13:1057953. <https://doi.org/10.3389/fpls.2022.1057953>

Li, J., Li, Y., Wang, R., Fu, J., Zhou, X., Fang, Y., ... & Liu, Y. (2022). Multiple Functions of **MiRNAs** in *Brassica napus* L. *Life*, 12(11), 1811. <https://doi.org/10.3390/life12111811>

Katche, E. I., Schierholt, A., Becker, H. C., Batley, J., & Mason, A. S. (2022). Fertility, genome stability, and homozygosity in a diverse set of **resynthesized rapeseed lines**. *The Crop Journal*. <https://doi.org/10.1016/j.cj.2022.07.022>

- Houmanat, K., Nabloussi, A., Rhazlaoui, Y., Bahri, H., EL FECHTALI, M., & CHARAFI, J. (2022). First report of **genetic relationship and diversity** among Moroccan and introduced rapeseed (*Brassica napus* L.) varieties as revealed by molecular markers. <https://doi.org/10.21203/rs.3.rs-2129788/v1>
- Dolatabadian, A., Yuan, Y., Bayer, P. E., Petereit, J., Severn-Ellis, A., Tirnaz, S., ... & Batley, J. (2022). **Copy Number Variation among Resistance Genes Analogues** in *Brassica napus*. *Genes*, 13(11), 2037. <https://doi.org/10.3390/genes13112037>
- Mei, D., Liu, J., & Wei, W. (2023). Identification and **phylogenetic analysis** of R2R3-MYB subfamily in *Brassica napus*. <https://doi.org/10.21203/rs.3.rs-2344198/v1>
- Wang, Z., Wang, F., Yu, Z., Shi, X., Zhou, X., Wang, P., ... & Yang, G. (2022). **Pyramiding of multiple genes** generates rapeseed introgression lines with clubroot and herbicide resistance, high oleic acid content, and early maturity. *The Crop Journal*. <https://doi.org/10.1016/j.cj.2022.10.009>
- Wang, A., Kang, L., Yang, G., & Li, Z. (2022). Transcriptomic and iTRAQ-Based Quantitative Proteomic Analyses of inap **CMS** in *Brassica napus* L. *Plants*, 11(19), 2460. <https://doi.org/10.3390/plants11192460>
- Wang, Z., Zhang, Y., Song, M., Tang, X., Huang, S., Linhu, B., ... & Xie, C. (2023). Genome-Wide Identification of the Cytochrome P450 Superfamily Genes and Targeted Editing of BnCYP704B1 Confers **Male Sterility** in Rapeseed. *Plants*, 12(2), 365. <https://doi.org/10.3390/plants12020365>
- Hu, R., Zhu, M., Chen, S., Li, C., Zhang, Q., Gao, L., ... & Qu, C. (2022). BnbHLH92a negatively regulates **anthocyanin and proanthocyanidin** biosynthesis in *Brassica napus*. *The Crop Journal*. <https://doi.org/10.1016/j.cj.2022.07.015>
- Li, S., Chang, T., Li, X., Peng, Z., Guan, C., & Guan, M. (2022). Regulatory mechanisms of rapeseed **petal color** formation: Current research status and future perspectives. *Oil Crop Science*. <https://doi.org/10.1016/j.ocsci.2022.11.005>
- Xu, Y., Yang, Y., Yu, W., Liu, L., Hu, Q., Wei, W., & Liu, J. (2022). Dissecting the Genetic Mechanisms of **Hemicellulose Content** in Rapeseed Stalk. *Agronomy*, 12(11), 2886. <https://doi.org/10.3390/agronomy12112886>
- Zhao, W., Liu, J., Qian, L., Guan, M., & Guan, C. (2022). Genome-wide identification and characterization of **oil-body-membrane proteins** in polyploid crop *Brassica napus*. *Plants*, 11(17), 2241. <https://doi.org/10.3390/plants11172241>
- Jia, Y., Yao, M., He, X., Xiong, X., Guan, M., Liu, Z., ... & Qian, L. (2022). Transcriptome and Regional Association Analyses Reveal the Effects of **Oleosin Genes** on the Accumulation of **Oil Content** in *Brassica napus*. *Plants*, 11(22), 3140. <https://doi.org/10.3390/plants11223140>
- Shi, J., Ni, X., Huang, J., Fu, Y., Wang, T., Yu, H., & Zhang, Y. (2022). CRISPR/Cas9-Mediated Gene Editing of BnFAD2 and BnFAE1 Modifies **Fatty Acid Profiles** in *Brassica napus*. *Genes*, 13(10), 1681. <https://doi.org/10.3390/genes13101681>
- Lipşa, F.-D.; Snowdon, R.; Wittkop, B.; Friedt, W. Quantitative **genetic analysis of phenolic acids** in oilseed rape meal. *Journal of Applied Life Sciences and Environment* 2022, 55 (2), 133-144. <https://doi.org/10.46909/alse-552051>

- Gao, C., Zhang, F., Hu, Y., Song, L., Tang, L., Zhang, X., ... & Wu, X. (2022). Dissecting the genetic architecture of **glucosinolate compounds** for quality improvement in flowering stalk tissues of *Brassica napus*. Horticultural Plant Journal. <https://doi.org/10.1016/j.hpj.2022.09.001>
- Zhou, X., Zhang, H., Xie, Z., Liu, Y., Wang, P., Dai, L., ... & Hong, D. (2023). Natural variation and artificial selection at the BnaC2. MYB28 locus modulate *Brassica napus* **seed glucosinolate**. Plant Physiology, 191(1), 352-368. <https://doi.org/10.1093/plphys/kiac463>
- AHMAD, I., SALEEM, S., GHAZALI, H., MASOOD, S., JAMIL, M., FAHEEM, U., & HUSSAIN, F. (2022). KHANPUR CANOLA (KN-279); NEWLY APPROVED LONG SILIQUED, BOLD SEEDED, HIGH YIELDING WITH IMPROVED QUALITY, DOUBLE ZERO (00) **VARIETY OF RAPESEED** (*Brassica Napus L.*). Biological and Clinical Sciences Research Journal, 2022(1). <https://doi.org/10.54112/bcsrj.v2022i1.119>
- Jin, Q., Gao, G., Guo, C. et al. Transposon insertions within alleles of BnaFT.A2 are associated with **seasonal crop type** in rapeseed. Theor Appl Genet 135, 3469–3483 (2022). <https://doi.org/10.1007/s00122-022-04193-x>
- Fan, S., Liu, H., Liu, J., Hua, W., & Li, J. (2022). BnGF14-2c Positively Regulates **Flowering** via the **Ver-nalization** Pathway in Semi-Winter Rapeseed. Plants, 11(17), 2312 <https://doi.org/10.3390/plants11172312>
- Chen, L., Lei, W., He, W., Wang, Y., Tian, J., Gong, J., ... & Fan, Z. (2022). Mapping of Two Major QTLs Controlling **Flowering Time** in *Brassica napus* Using a High-Density Genetic Map. Plants, 11(19), 2635. <https://doi.org/10.3390/plants11192635>
- Fu, R., Wang, J., Zhou, M. et al. Five NUCLEAR FACTOR-Y subunit B genes in rapeseed (*Brassica napus*) promote **flowering and root elongation** in Arabidopsis. Planta 256, 115 (2022). <https://doi.org/10.1007/s00425-022-04030-x>
- Liu, T., Li, Y., Wang, C., Zhang, D., Liu, J., He, M., ... & Guo, Y. (2023). *Brassica napus* Transcription Factor Bna. A07. WRKY70 Negatively Regulates **Leaf Senescence** in Arabidopsis thaliana. Plants, 12(2), 347. <https://doi.org/10.3390/plants12020347>
- Chen, Y., Zhu, W., Yan, T. et al. **Stomatal morphological variation** contributes to global ecological **ad-aptation** and diversification of *Brassica napus*. Planta 256, 64 (2022). <https://doi.org/10.1007/s00425-022-03982-4>
- Ma, L., Xu, J., Tao, X., Wu, J., Wang, W., Pu, Y., ... & Sun, W. (2022). Genome-Wide Identification of C2H2 ZFPs and Functional Analysis of BRZAT12 under **Low-Temperature Stress** in Winter Rapeseed (*Brassica rapa*). International Journal of Molecular Sciences, 23(20), 12218. <https://doi.org/10.3390/ijms232012218>
- Liu, X., Wei, R., Tian, M., Liu, J., Ruan, Y., Sun, C., & Liu, C. (2022). Combined Transcriptome and Metabolome Profiling Provide Insights into **Cold Responses** in Rapeseed (*Brassica napus L.*) Genotypes with Contrasting Cold-Stress Sensitivity. International Journal of Molecular Sciences, 23(21), 13546. <https://doi.org/10.3390/ijms232113546>
- Sharma, A., Kumari, V., & Rana, A. (2022). Genetic Variability Studies on **Drought Tolerance** using Agro-Morphological and Yield Contributing Traits in Rapeseed-Mustard. <https://doi.org/10.23910/1.2022.2878>

- Zhang, T., Zhou, T., Zhang, Y., Chen, J., Song, H., Wu, P., ... & Hua, Y. (2022). Genome-Wide Identification and Functional Characterization Reveals the Pivotal Roles of BnaA8. ATG8F in **Salt Stress Tolerance and Nitrogen Limitation Adaptation** in Allotetraploid Rapeseed. *International Journal of Molecular Sciences*, 23(19), 11318. <https://doi.org/10.3390/ijms231911318>
- Guo, Y., Li, D., Liu, T., Liao, M., Li, Y., Zhang, W., ... & Chen, M. (2022). Effect of Overexpression of  $\gamma$ -Tocopherol Methyltransferase on  $\alpha$ -Tocopherol and Fatty Acid Accumulation and Tolerance to **Salt Stress** during Seed Germination in *Brassica napus* L. *International Journal of Molecular Sciences*, 23(24), 15933. <https://doi.org/10.3390/ijms232415933>
- Zhu, J., Lei, L., Wang, W. et al. QTL mapping for **seed density per silique** in *Brassica napus*. *Sci Rep* 13, 772 (2023). <https://doi.org/10.1038/s41598-023-28066-5>
- Ma, X., Wang, J., Gu, Y., Fang, P., Nie, W., Luo, R., ... & Mei, J. (2023). Genetic analysis and QTL mapping for **silique density** in rapeseed (*Brassica napus* L.). <https://doi.org/10.21203/rs.3.rs-2475794/v1>
- Geng, R., Shan, Y., Li, L., Shi, C. L., Zhang, W., Wang, J., ... & Tan, X. L. (2022). CRISPR-mediated BnaIDA editing prevents **silique shattering**, floral organ abscission, and spreading of *Sclerotinia sclerotiorum* in *Brassica napus*. *Plant Communications*, 3(6), 100452. <https://doi.org/10.1016/j.xplc.2022.100452>
- Cao, B., Wang, H., Bai, J., Wang, X., Li, X., Zhang, Y., ... & Yu, X. (2022). miR319-Regulated TCP3 Modulates Silique Development Associated with **Seed Shattering** in *Brassicaceae*. *Cells*, 11(19), 3096. <https://doi.org/10.3390/cells11193096>
- Dong, H., Yang, L., Liu, Y., Tian, G., Tang, H., Xin, S., ... & Qian, W. (2022). Detection of new candidate genes controlling **seed weight** by integrating gene coexpression analysis and QTL mapping in *Brassica napus* L. *The Crop Journal*. <https://doi.org/10.1016/j.cj.2022.09.009>
- Cowling, W. A., Castro-Urrea, F. A., Stefanova, K. T., Li, L., Banks, R. G., Saradadevi, R., ... & Siddique, K. H. (2023). Optimal Contribution Selection Improves the Rate of Genetic Gain in **Grain Yield and Yield Stability** in Spring Canola in Australia and Canada. *Plants*, 12(2), 383. <https://doi.org/10.3390/plants12020383>
- Zhang, L., Yang, B., Zhang, C., Chen, H., Xu, J., Qu, C., ... & Li, J. (2023). Genome-Wide Identification and Posttranscriptional Regulation Analyses Elucidate Roles of Key Argonautes and Their miRNA Triggers in Regulating Complex **Yield Traits** in Rapeseed. *International Journal of Molecular Sciences*, 24(3), 2543. <https://doi.org/10.3390/ijms24032543>
- Lu, H., Chen, L., Du, M., Lu, H., Liu, J., Ye, S., ... & Shen, J. (2023). miR319 and its target TCP4 involved in **plant architecture regulation** in *Brassica napus*. *Plant Science*, 326, 111531. <https://doi.org/10.1016/j.plantsci.2022.111531>
- Tan, Y., Ren, L., Wang, J., Ran, S., Wu, L., Cheng, Z., ... & Liu, L. (2022). Identification and characterization of a **curly-leaf** locus CL1 encoding an IAA2 protein in *Brassica napus*. *The Crop Journal*. <https://doi.org/10.1016/j.cj.2022.11.001>
- Channaoui, S., Mazouz, H., Labhilili, M., El Fechtali, M., & Nabloussi, A. (2022). Inheritance of **dwarfism and narrow lobed-leaf** in two rapeseed (*Brassica napus* L.) mutant lines. *Heliyon*, e12649. <https://doi.org/10.1016/j.heliyon.2022.e12649>

- Sun, Q., Xi, Y., Lu, P., Lu, Y., Wang, Y., & Wang, Y. (2022). Genome-wide analysis of the G-box regulating factors protein family reveals its roles in response to *Sclerotinia sclerotiorum* infection in rapeseed (*Brassica napus* L.). *Frontiers in Plant Science*, 13. <https://doi.org/10.3389/fpls.2022.986635>
- Sarwar, R., Li, L., Yu, J., Zhang, Y., Geng, R., Meng, Q., ... & Tan, X. L. (2023). Functional Characterization of the Cystine-Rich-Receptor-like Kinases (CRKs) and Their Expression Response to *Sclerotinia sclerotiorum* and **Abiotic Stresses** in *Brassica napus*. *International Journal of Molecular Sciences*, 24(1), 511. <https://doi.org/10.3390/ijms24010511>
- Zhang, Y., Wang, Y., Wu, D., Qu, D., Sun, X., & Zhang, X. (2022). SNP and Haplotype Variability in the BnP5CR2 Gene and Association with Resistance and Susceptible Cultivars for *Sclerotinia sclerotiorum* in *Brassica napus*. *Agronomy*, 12(12), 2956. <https://doi.org/10.3390/agronomy12122956>
- Adhikary, D., Kisiala, A., Sarkar, A., Basu, U., Rahman, H., Emery, N., & Kav, N. N. (2022). Early-stage responses to *Plasmodiophora brassicae* at the transcriptome and metabolome levels in clubroot resistant and susceptible oilseed *Brassica napus*. *Molecular Omics*, 18(10), 991-1014. <https://doi.org/10.1039/D2MO00251E>
- Wang, Z. (2022). Genetic and molecular analysis of **clubroot resistance** in canola introgressed from rutabaga cvs. Polycross and Brookfield. <https://doi.org/10.7939/r3-0456-2r30>
- Liu, Y. (2022). Use of Turnip (*Brassica rapa* var. *rapifera*) and Rutabaga (*B. napus* var. *napobrassica*) for the Improvement of **Clubroot Resistance** in Spring *B. napus* Canola. <https://doi.org/10.7939/r3-irfy-gw21>
- Hu, L., Zhai, Y., Xu, L., Wang, J., Yang, S., Sun, Y., ... & Fan, C. (2022). Precise A·T to G·C base **editing** in the allotetraploid rapeseed (*Brassica napus* L.) genome. *Journal of Cellular Physiology*. <https://doi.org/10.1002/jcp.30904>
- Qing-Yong, Y., Yang, Z., Wang, S., Wei, L., Huang, Y., Liu, D., ... & Guo, L. (2023). BnIR: a **multi-omics collaborative resource** and tool platform for *Brassica napus* biological study. *bioRxiv*, 2023-01. <https://doi.org/10.1101/2023.01.12.523736>
- Li, X., Liu, X., Fan, Y., Li, S., Yu, M., Qian, M., ... & Lu, K. (2022). Development of a target capture sequencing SNP genotyping platform for genetic analysis and genomic breeding in rapeseed. *The Crop Journal*. <https://doi.org/10.1016/j.cj.2022.08.008>
- Kozar, E. V., Kozar, E. G., & Domblides, E. A. (2022). Effect of the Method of Microspore Isolation on the Efficiency of Isolated Microspore Culture In Vitro for *Brassicaceae* Family. *Horticulturae*, 8(10), 864. <https://doi.org/10.3390/horticulturae810086>
- Jiao, Y., Liang, B., Yang, G. et al. A simple and efficient **method to quantify the cell parameters** of the seed coat, embryo and silique wall in rapeseed. *Plant Methods* 18, 117 (2022). <https://doi.org/10.1186/s13007-022-00948-1>
- Gholizadeh, A., Oghan, H. A., Alizadeh, B., Rameeh, V., Payghamzadeh, K., Bakhshi, B., ... & Shariati, F. (2022). **Phenotyping** new rapeseed lines based on multiple traits: Application of GT and GYT biplot analyses. *Food Science & Nutrition*. <https://doi.org/10.1002/fsn3.3119>

## CROP PROTECTION

- Kuyper, T.W. The smartest plant?. *Plant Soil* (2023). <https://doi.org/10.1007/s11104-022-05859-7>
- Jamalzadeh, A., Darvishnia, M., Khodakaramian, G. et al. Effect of **antagonistic bacteria** associated with canola on **disease** suppression. *Eur J Plant Pathol* (2023). <https://doi.org/10.1007/s10658-022-02633-4>
- Zhang, M., Gong, Q., Su, X., Cheng, Y., Wu, H., Huang, Z., ... & Yu, C. (2022). Microscopic and Transcriptomic Comparison of **Powdery Mildew Resistance** in the Progenies of *Brassica carinata* × *B. napus*. *International Journal of Molecular Sciences*, 23(17), 9961. <https://doi.org/10.3390/ijms23179961>
- Priyanka, B. S., Singh, H. K., & Srivastava, J. P. (2022). Noble Indices for Degree of Host Plant Resistance against **Alternaria Blight** in Rapeseed-mustard Genotypes. *International Journal of Environment and Climate Change*, 12(12), 1379-1387. <https://doi.org/10.9734/ijecc/2022/v12i121577>
- Huang, Y. J., Sidique, S. N. M., Karandeni Dewage, C. S., Gajula, L. H., Mitrousia, G. K., Qi, A., ... & Fitt, B. D. (2022). Effective control of **Leptosphaeria maculans** increases importance of *L. biglobosa* as a cause of phoma stem canker epidemics on oilseed rape. *Pest Management Science*. <https://doi.org/10.1002/ps.7248>
- Tao Chen, Yanli Zhao, Chao Li, et al. Overexpression of a chitinase gene *PbChia1* from **Plasmodiophora brassicae** increases broad spectrum disease resistance. *Authorea*. October 27, 2022. <https://doi.org/10.22541/au.166686672.20472606/v1>
- Javed, M. A., Schwelm, A., Zamani-Noor, N., Salih, R., Silvestre Vañó, M., Wu, J., ... & Pérez-López, E. (2022). The **clubroot** pathogen **Plasmodiophora brassicae**: A profile update. *Molecular Plant Pathology*. <http://dx.doi.org/10.1111/mpp.13283>
- Wang, Y. (2022). Factors stimulating germination of **Plasmodiophora brassicae** resting spores. PhD thesis University of Göttingen <http://dx.doi.org/10.53846/goediss-9600>
- Zuzak, K. A., Strelkov, S. E., Turnbull, G. D., Manolii, V. P., & Hwang, S. F. (2022). **Soil fumigation** with Vapam (metam sodium) to control **clubroot** (*Plasmodiophora brassicae*) of canola (*Brassica napus*). *Canadian Journal of Plant Science*. <https://doi.org/10.1139/cjps-2022-0086>
- Roth, M. N. (2022). Evaluation of **Plasmodiophora brassicae** for the occurrence of pH insensitive isolates <https://doi.org/10.7939/r3-1yh9-cj49>
- McBain, E. (2022). **Monitoring Airborne Inoculum of Sclerotinia sclerotiorum** at Canola Flowering and Relationships to Weather Conditions and Disease Incidence and Severity. <https://doi.org/10.7939/r3-qwme-3584>
- Duarte Riveros, P. (2022). Lab-on-a-Chip Designs for Airborne **Spore Detection**: Towards the Forecasting of **Sclerotinia** Stem Rot of Canola. <https://doi.org/10.7939/r3-ggpy-7p68>
- Gupta N.C., S. Arora, Aditi Kundu, Pankaj Sharma, M. Rao and R.Bhattarchatya. 2022. UPLC-Q-TOF-MS-based untargeted studies of the secondary metabolites secreted by **Sclerotinia sclerotiorum** under the axenic conditions. *J Plant Science and Phytopathology*.6:173-182. <https://doi.org/10.29328/journal.jpss.1001095>
- Gupta N.C., S. Yadav, S. Arora, D.C. Mishra, N. Budhlakoti, K. Gaikwas, M. Rao, L. Prasad, P.K. Rai and Pankaj Sharma. 2022. Draft genome sequencing and secretome profiling of **Sclerotinia**

- sclerotiorum*** revealed effector repertoire diversity and allied broad-host range necrotrophy. Scientific Reports . 12:21855. <https://doi.org/10.1038/s41598-022-22028-z>
- Cui, J., Strelkov, S. E., Fredua-Agyeman, R., & Hwang, S. F. (2022). Development of optimized ***Verticillium longisporum*** inoculation techniques for canola (*Brassica napus*). Canadian Journal of Plant Pathology, 1-11. <https://doi.org/10.1080/07060661.2022.2120913>
- Wang, Y., Strelkov, S. E., & Hwang, S. F. (2023). Blackleg Yield Losses and Interactions with ***Verticillium*** Stripe in Canola (*Brassica napus*) in Canada. Plants, 12(3), 434. <https://doi.org/10.3390/plants12030434>
- Umer, M., Qadeer, A., Razaq, Z., Anwar, N., & Kiptoo, J. J. Mycovirus: **Biocontrol agent against *S. sclerotiorum*** of Rapeseed. [REFERENCE](#)
- Jayasena, K. W., van BURGEL, A. N. D. R. E. W., Galloway, J., & Thomas, G. (2022). **Detection** and quantification of ***Sclerotinia sclerotiorum*** infestation of canola petals using a clearing and staining technique. Canadian Journal of Plant Pathology, 1-6. <https://doi.org/10.1080/07060661.2022.2119603>
- Sharma, Pankaj, Ritu Mawar and K.S. Jadon. 2023. Diseases of oilseed crops in India: Diagnosis and management. Indian Phytopathological Society, New Delhi. pp.153. (book)
- Laforest, M., Martin, S., Soufiane, B., Bisailon, K., Maheux, L., Fortin, S., ... & Simard, M. J. (2022). Distribution and genetic characterization of bird rape mustard (*Brassica rapa*) populations and analysis of **glyphosate resistance introgression**. Pest Management Science, 78(12), 5471-5478. <https://doi.org/10.1002/ps.7170>
- Soltani, N., Geddes, C., Laforest, M., Dille, J. A., & Sikkema, P. H. (2022). Economic impact of **glyphosate-resistant weeds** on major field crops grown in Ontario. Weed Technology, 36(5), 629-635. <https://doi.org/10.1017/wet.2022.72>
- Geddes, C.M., Pittman, M.M. First report of **glyphosate-resistant downy brome** (*Bromus tectorum* L.) in Canada. Sci Rep 12, 18893 (2022). <https://doi.org/10.1038/s41598-022-21942-6>
- Anderson, J. V., Bigger, B. B., Howatt, K., Mettler, J., & Berti, M. T. (2022). Weed Pressure, Nutrient Content, and Seed Yield in Field Grown **Sulfonylurea-Resistant *Camelina sativa*** and *Brassica napus*. Agronomy, 12(11), 2622. <https://doi.org/10.3390/agronomy12112622>
- Geddes, C. M., Tidemann, B. D., Ikley, J. T., Dille, J. A., Soltani, N., & Sikkema, P. H. Potential spring canola yield **losses due to weeds** in Canada and the United States. Weed Technology, 1-19. <https://doi.org/10.1017/wet.2022.88>
- Bellec, L., Hervé, M. R., Mercier, A. S., Lenal, P. A., Faure, S., & Cortesero, A. M. (2022). A protocol for increased throughput phenotyping of **plant resistance to the pollen beetle**. Pest Management Science. <https://doi.org/10.1002/ps.7266>
- Seimandi-Corda, G., Hall, J., Jenkins, T., & Cook, S. M. (2022). Relative efficiency of methods to estimate **cabbage stem flea beetle** (*Psylliodes chrysocephala*) larval infestation in oilseed rape (*Brassica napus*). Pest Management Science. <https://doi.org/10.1002/ps.7341>
- Desroches, C., Bouchard, P., Labrie, G., & Lucas, E. (2022). Assemblage of **Ceutorhynchinae Weevils** Associated With Brassicaceae in Quebec (Canada) Agroecosystems. Environmental Entomology. <https://doi.org/10.1093/ee/nvac097>

- Herbertsson, L., Klatt, B. K., Blasi, M., Rundlöf, M., & Smith, H. G. (2022). Seed-coating of rapeseed (*Brassica napus*) with the **neonicotinoid** clothianidin affects behaviour of red mason bees (*Osmia bicornis*) and **pollination** of strawberry flowers (*Fragaria x ananassa*). Plos one, 17(9), e0273851. <https://doi.org/10.1371/journal.pone.0273851>
- Yadav, S., Jat, M. K., Yadav, S. S., & Kumar, H. (2022). Diversity, abundance and **foraging behaviour of pollinators** in early sown rapeseed-mustard genotypes. Journal of Agriculture and Ecology, 14, 104-112. <https://journals.saaer.org.in/index.php/jae/article/view/519>
- Perig, N. (2022). Study of the influence of **protein food** on the development and productivity of **queen bees**. Scientific Messenger of LNU of Veterinary Medicine and Biotechnologies. Series: Agricultural sciences, 24(97), 76-81. <https://doi.org/10.32718/nvlvet-a9713>
- Pineaux, M., Grateau, S., Lirand, T., Aupinel, P., & Richard, F. J. (2023). **Honeybee queen exposure** to a widely used fungicide disrupts reproduction and colony dynamic. Environmental Pollution, 121131. <https://doi.org/10.1016/j.envpol.2023.121131>
- Roy, N. **Behavioral pattern** of generalist and specialist **insect pests** to brassicaceous leaf cuticular n-alkanes and free fatty acids. Arthropod-Plant Interactions 16, 537–551 (2022). <https://doi.org/10.1007/s11829-022-09917-w>
- Karthik, S., Yashaswini, G., Mukherjee, U. et al. **Phylogenetic divergence** of *Lipaphis erysimi pseudo-brassicae* (Aphididae: Homoptera): a dominant specialist aphid on Rapeseed-mustard, India. Biologia 77, 3603–3614 (2022). <https://doi.org/10.1007/s11756-022-01213-5>
- Sharma, O., & Singh, D. K. (2022). Assessment of rapeseed-mustard genotypes against mustard aphid, *Lipaphis erysimi* (Kalt.) under natural infestation conditions. <https://www.thepharmajournal.com/archives/2022/vol11issue10S/PartL/S-11-10-140-650.pdf>
- Kirkland, L. S., Chirgwin, E., Ward, S. E., Congdon, B. S., van Rooyen, A., & Umina, P. A. P450-mediated resistance in *Myzus persicae* (Sulzer)(Hemiptera: Aphididae) reduces the efficacy of neonicotinoid seed treatments in *Brassica napus*. Pest Management Science. <https://doi.org/10.1002/ps.7362>
- Slavíková, L., Ibrahim, E., Alquicer, G., Tomašechová, J., Šoltys, K., Glasa, M., & Kundu, J. K. (2022). **Weed Hosts** Represent an Important Reservoir of **Turnip Yellows Virus** and a Possible Source of Virus Introduction into Oilseed Rape Crop. Viruses, 14(11), 2511. <https://doi.org/10.3390/v14112511>
- Armand Pilón, A., Silva, H., Abbate, S., Bentancur, Ó., & Huguaburu, V. (2022). Development and reproductive potential of *Plutella xylostella* (Lepidoptera: Plutellidae) in five Brassicaceae hosts. International Journal of Pest Management, 68(4), 381-389. <https://doi.org/10.1080/09670874.2022.2134942>
- Solà, M., Dumont, F., Provost, C., & Lucas, E. *Lygus lineolaris* nutritional needs and host choices for IPM strategies with predators and trap crops. [https://www.cram-mirabel.com/wp-content/uploads/2022/11/Sola-Vegephy12021\\_final.pdf](https://www.cram-mirabel.com/wp-content/uploads/2022/11/Sola-Vegephy12021_final.pdf)
- Kaivosoja, J., Ronkainen, A., Hautsalo, J., Backman, J., Linkolehto, R., Emeterio, M. S., & Soininen, J. P. (2022, November). Automated Rapeseed **Pest Detection and Management with Drones**. In ROBOT2022: Fifth Iberian Robotics Conference: Advances in Robotics, Volume 2 (pp. 427-437).

Cham: Springer International Publishing. [https://doi.org/10.1007/978-3-031-21062-4\\_35](https://doi.org/10.1007/978-3-031-21062-4_35) or [REFERENCE](#)

Fricke, U., Redlich, S., Zhang, J., Benjamin, C. S., Englmeier, J., Ganuza, C., ... & Steffan-Dewenter, I. (2022). **Earlier flowering** of winter oilseed rape compensates for higher **pest pressure** in warmer climates. *Journal of Applied Ecology*. <https://doi.org/10.1111/1365-2664.14335>

Naz, S., & Abbas, M. DIVERSITY, RICHNESS, AND ASSOCIATED **PREDATORS OF ARTHROPOD PESTS ON CANOLA CROP** (*Brassica napus* L.). <https://www.doi.org/10.56726/IRJMETS32491>

### **AGRONOMY & CROP MANAGEMENT**

Zandberg, J. D., Fernandez, C. T., Danilevycz, M. F., Thomas, W. J., Edwards, D., & Batley, J. (2022). The Global Assessment of Oilseed Brassica Crop Species Yield, Yield Stability and the Underlying Genetics. *Plants*, 11(20), 2740. <https://doi.org/10.3390/plants11202740>

Shcherbyna, O., Tymoshenko, O., Levchenko, T., Baidiuk, T., Veresenko, O., & Hurenko, A. (2022). Characteristics of new breeding material of winter rape by important economic and valuable characteristics at different stages of breeding process. *Agriculture and Plant Sciences: Theory and Practice*, (3), 76-82. Retrieved from <http://journal-agriplant.com/index.php/journal/article/view/51>

Secchi, M. (2022). New insights of winter canola survival, seed quality, and yield for the **Great Plains** region and the United States (Doctoral dissertation). <https://hdl.handle.net/2097/42843>

Arinaitwe, U., Clay, S. A., & Nleya, T. Growth, **Yield and Yield Stability** of Canola (*Brassica Napus*) in the Northern **Great Plains** of the **US**. *Agronomy Journal*. <https://doi.org/10.1002/agj2.21269>

Xie, Z., Kong, J., Tang, M., Luo, Z., Li, D., Liu, R., ... & Zhang, C. (2023). Modelling Winter Rapeseed (*Brassica napus* L.) Growth and Yield under Different Sowing Dates and Densities Using **Aqua-Crop Model**. *Agronomy*, 13(2), 367. <https://doi.org/10.3390/agronomy13020367>

Kakati, N., Deka, R.L., Das, P. et al. **Forecasting yield** of rapeseed and mustard using multiple linear regression and ANN techniques in the Brahmaputra valley of Assam, North East India. *Theor Appl Climatol* 150, 1201–1215 (2022). <https://doi.org/10.1007/s00704-022-04220-3>

Rajković, D., Jeromela, A. M., Pezo, L., Lončar, B., Grahovac, N., & Špika, A. K. (2023). Artificial neural network and random forest regression models for **modelling fatty acid and tocopherol content** in oil of winter rapeseed. *Journal of Food Composition and Analysis*, 115, 105020. <https://doi.org/10.1016/j.jfca.2022.105020>

Guirrou, I.; El Harrak, A.; El Antari, A.; Hssaini, L.; Hanine, H.; El Fechtali, M.; Nabloussi, A. **Bioactive Compounds** Assessment in Six Moroccan **Rapeseed** (*Brassica napus* L.) Varieties Grown in Two Contrasting Environments. *Agronomy* **2023**, *13*, 460. <https://doi.org/10.3390/agronomy13020460>

Han, S., Liu, J., Zhou, G., Jin, Y., Zhang, M., & Xu, S. (2022). InceptionV3-LSTM: A Deep Learning Net for the Intelligent **Prediction of Rapeseed Harvest Time**. *Agronomy*, 12(12), 3046. <https://doi.org/10.3390/agronomy12123046>

- Nelson, M. N., Nesi, N., Barrero, J. M., Fletcher, A. L., Greaves, I. K., Hughes, T., ... & Kirkegaard, J. A. (2022). Strategies to improve **field establishment** of canola: A review. *Advances in Agronomy*. <https://doi.org/10.1016/bs.agron.2022.05.001>
- Jiang, L., Tang, Q., Wu, J., Yu, W., Zhang, M., Jiang, D., & Wei, D. (2022). Design and Test of **Seedbed Preparation** Machine before Transplanting of Rapeseed Combined Transplanter. *Agriculture*, 12(9), 1427. <https://doi.org/10.3390/agriculture12091427>
- Wang, R., Wu, W., Cheng, X., & Peng, W. (2023). **High plant density** increases sunlight interception and yield of **direct-seeded** winter canola in China. *Experimental Agriculture*, 59, e2. <https://doi.org/10.1017/S0014479722000564>
- Yan, E., Carozzi, M., Munier-Jolain, N., & Martin, P. (2022, August). Potential of **crop mixtures to reduce pesticide** use in France. A data analysis. In XVII. Congress of the European Society for Agronomy. <https://hal.inrae.fr/hal-03927116>
- Wang, Z., Song, L., Wang, C., Guo, M., El-Badri, A. M., Batool, M., ... & Zhou, G. (2023). Rapeseed-maize **double-cropping** with high biomass and high economic benefits is a soil environment-friendly forage production mode in the Yangtze River Basin. *European Journal of Agronomy*, 142, 126675. <https://doi.org/10.1016/j.eja.2022.126675>
- Dowling, A., Roberts, P., Doolette, A., Zhou, Y., & Denton, M. D. (2023). **Oilseed-legume intercropping** is productive and profitable in low input scenarios. *Agricultural Systems*, 204, 103551. <https://doi.org/10.1016/j.agsy.2022.103551>
- Imran, Amanullah, & Al Tawaha, A. R. (2022). Regenerating Potential of **Dual Purpose Rapeseed** (*Brassica Napus L.*) as Influenced by Decapitation Stress and Variable Rates of Phosphorous. *Communications in Soil Science and Plant Analysis*, 1-10. <https://doi.org/10.1080/00103624.2022.2118297>
- Jamar, D., Daniaux, C., & Regibeau, C. (2022). Rapeseed regrowth and white clover cover crop valorisation through **sheep grazing** (DiverIMPACTS Practice Abstract). <https://zenodo.org/record/6514829#.YyHWCHZByUk>
- Hegewald, H. Oilseed rape in **high-intensity crop rotations** (Doctoral dissertation, Dissertation, Halle (Saale), Martin-Luther-Universität Halle-Wittenberg, 2022). [https://opendata.uni-halle.de/bitstream/1981185920/92660/1/Hegewald\\_Diss\\_Bibliothek\\_online.pdf](https://opendata.uni-halle.de/bitstream/1981185920/92660/1/Hegewald_Diss_Bibliothek_online.pdf)
- Abdirazzakovich, B. A., & Razzak, O. (2022). **EFFECT OF PREVIOUS CROPS ON SOIL FERTILITY AND WINTER WHEAT YIELD**. *Web of Scientist: International Scientific Research Journal*, 3(12), 723-727. <https://wos.academiascience.org/index.php/wos/article/view/3015>
- Schillinger, W. F., Paulitz, T. C., & Hansen, J. C. **Canola Rotation** Effects on Soil Water and Subsequent Wheat in the Pacific Northwest USA. *Agronomy Journal*. <https://doi.org/10.1002/agj2.21248>
- Wen, G., Ma, B. L., Vanasse, A., Caldwell, C. D., & Smith, D. L. (2022). Optimizing **machine learning**-based site-specific **nitrogen application recommendations** for canola production. *Field Crops Research*, 288, 108707. <https://doi.org/10.1016/j.fcr.2022.108707>
- Mohammed, Y., Abdelghani, N., Abdelwahed, M., Abdellatif, B., & Khalid, D. (2022). Effects of **nitrogen rates** on yield, yield components, and other related attributes of different rapeseed (*Brassica napus L.*) varieties. <https://doi.org/10.1051/ocl/2022001>

- Li, Z., Liu, F., & Wu, W. (2023). Optimising **nitrogen management** strategies to minimise lodging risk while sustaining high seed yield in rapeseed. *European Journal of Agronomy*, 142, 126671. <https://doi.org/10.1016/j.eja.2022.126671>
- Ebaid, M., Abd El-Hady, M.A., El-Temseh, M.E. et al. Response of Canola productivity to integration between **mineral nitrogen** with yeast extract under poor fertility sandy soil condition. *Sci Rep* 12, 20216 (2022). <https://doi.org/10.1038/s41598-022-24645-0>
- Li, Jing and Zhou, Yangguo and Gu, Hehe and Lu, Zhifeng and Cong, Rihuan and Li, Xiaokun and Ren, Tao and Lu, Jianwei, Synergistic **Nitrogen and Potassium** Improvement in Winter Oilseed Rape (*Brassica Napus L.*) Yield for Sustainable Nitrogen Management. Available at SSRN: <https://ssrn.com/abstract=4210121> or <http://dx.doi.org/10.2139/ssrn.4210121>
- Wang, R., Liu, A., Chen, X., Wu, Y., & Peng, W. (2023). Effect of **potassium rate** on yield, potassium uptake and canopy radiation interception of direct-seeded winter canola. *Chilean journal of agricultural research*, 83(1), 107-118. <http://dx.doi.org/10.4067/S0718-58392023000100107>
- Tomczyk, M., Zaguła, G., Kaczmarek, M., Puchalski, C., & Dżugan, M. (2023). The Negligible Effect of **Toxic Metal** Accumulation in the Flowers of Melliferous Plants on the Mineral Composition of **Monofloral Honeys**. *Agriculture*, 13(2), 273. <https://doi.org/10.3390/agriculture13020273>
- Meyer-Aurich, A., & Karatay, Y. N. (2022). **Greenhouse Gas Mitigation** Costs of Reduced **Nitrogen** Fertilizer. *Agriculture*, 12(9), 1438. <https://doi.org/10.3390/agriculture12091438>
- ÖZPINAR, S. Analysis of **Energy Use** Efficiency and **Greenhouse Gas** Emission in Rainfed Canola Production (Case study: Çanakkale Province, Turkey). *Tekirdağ Ziraat Fakültesi Dergisi*, 20(1), 197-210. <https://doi.org/10.33462/jotaf.1121863>
- Bami, S. K., Ardakani, M. R., Dameghani, A. M., Rad, A. H. S., & Manavi, P. N. (2022). The relative contribution of applied *inputs* of rapeseed (*Brassica napus L.*) agro-ecosystem on **environmental** factors. *Plant Science Today*, 9(4), 874-880. <https://doi.org/10.14719/pst.1707>
- Wang, X., Ma, H., Guan, C., & Guan, M. (2022). Decomposition of **Rapeseed Green Manure** and Its Effect on Soil under Two Residue Return Levels. *Sustainability*, 14(17), 11102. <https://doi.org/10.3390/su141711102>
- Zakharova, O. V., Baranchikov, P. A., Grodetzkaya, T. A., Kuznetsov, D. V., & Gusev, A. A. (2022). Highly Dispersed **Blast-Furnace Sludge** as a New **Micronutrient Fertilizer**: Promising Results on Rapeseed. *Agronomy*, 12(12), 2929. <https://doi.org/10.3390/agronomy12122929>
- Tian, X., Li, Z., Wang, Y., Li, B., Xie, X., & Wang, L. (2022). Effects of biochar combined with **nitrogen** reduction on the **soil microbial community** of rapeseed. *Archives of Agronomy and Soil Science*, 1-15. <https://doi.org/10.1080/03650340.2022.2132479>
- Liu, Z., Li, S., Liu, N., Huang, G., & Zhou, Q. (2022). **Soil microbial community** driven by soil moisture and nitrogen in milk vetch (*Astragalus sinicus L.*)–rapeseed (*Brassica napus L.*) intercropping. *Agriculture*, 12(10), 1538. <https://doi.org/10.3390/agriculture12101538>
- Chen, S., Yang, D., Wei, Y., He, L., Li, Z., & Yang, S. (2023). Changes in **Soil Phosphorus** Availability and **Microbial Community** Structures in Rhizospheres of Oilseed Rapes Induced by **Intercropping** with White Lupins. *Microorganisms*, 11(2), 326. <https://doi.org/10.3390/microorganisms11020326>

- Iqbal, M., Naveed, M., Sanaullah, M. et al. **Plant microbe** mediated enhancement in growth and yield of canola (*Brassica napus L.*) plant through auxin production and increased nutrient acquisition. *J Soils Sediments* (2022). <https://doi.org/10.1007/s11368-022-03386-7>
- Li, Y., Vail, S. L., Arcand, M. M., & Helgason, B. (2023). Contrasting **nitrogen fertilization** and *Brassica napus* (canola) variety development impact recruitment of the root-associated **microbiome**. *Phytobiomes Journal*, (ja). <https://doi.org/10.1094/PBIOMES-07-22-0045-R>
- Spiridonov, Y.Y., Chkanikov, N.D., Pastukhov, A.V. et al. Protection of Spring Rapeseed from the Phytotoxic Effect of **Metsulfuron-Methyl Residues** Using Zeolites. *Russ. Agricult. Sci.* 48 (Suppl 1), S84–S88 (2022). <https://doi.org/10.3103/S1068367422070187>
- El Gafary, R. F., Eid, S. F., Gameh, M. A., & Abdelwahab, M. K. (2022). Irrigation Water Management of Canola Crop under Surface and Subsurface **Drip Irrigation Systems** at Toshka Area. *Egypt. Journal of Soil Sciences and Agricultural Engineering*, 13(10), 331-337. <https://dx.doi.org/10.21608/jssae.2022.162492.1105>

## PHYSIOLOGY

- Jankauskienė, J., Mockevičiūtė, R., Gavelienė, V., Jurkonienė, S., & Anisimovienė, N. (2022). The application of auxin-like compounds promotes **cold acclimation** in the oilseed rape plant. *Life*, 12(8), 1283. <https://doi.org/10.3390/life12081283>
- Luo, T., Sheng, Z., Zhang, C., Li, Q., Liu, X., Qu, Z., & Xu, Z. (2022). Seed Characteristics Affect **Low-Temperature Stress Tolerance** Performance of Rapeseed (*Brassica napus L.*) during Seed Germination and Seedling Emergence Stages. *Agronomy*, 12(8), 1969. <https://doi.org/10.3390/agronomy12081969>
- Zhu, K., Liu, J., Luo, T., Zhang, K., Khan, Z., Zhou, Y., ... & Hu, L. (2022). Wood Vinegar Impact on the Growth and **Low-Temperature Tolerance** of Rapeseed Seedlings. *Agronomy*, 12(10), 2453. <https://doi.org/10.3390/agronomy12102453>
- Lei, Y., He, H., Raza, A., Liu, Z., Xiaoyu, D., Guijuan, W., ... & Xiling, Z. (2022). Exogenous melatonin confers **cold tolerance** in rapeseed (*Brassica napus L.*) seedlings by improving antioxidants and genes expression. *Plant Signaling & Behavior*, 17(1), 2129289. <https://doi.org/10.1080/15592324.2022.2129289>
- Cheng, P., Feng, L., Zhang, S. et al. Ammonia borane positively regulates **cold tolerance** in *Brassica napus* via hydrogen sulfide signaling. *BMC Plant Biol* 22, 585 (2022). <https://doi.org/10.1186/s12870-022-03973-3>
- Ma, J., Islam, F., Ayyaz, A., Fang, R., Hannan, F., Farooq, M. A., ... & Zhou, W. (2022). Wood vinegar induces **salinity tolerance** by alleviating oxidative damages and protecting photosystem II in rapeseed cultivars. *Industrial Crops and Products*, 189, 115763. <https://doi.org/10.1016/j.indcrop.2022.115763>
- Hasanuzzaman, M., Raihan, M., Hossain, R., Alharby, H. F., Al-Zahrani, H. S., Alsamadany, H., ... & Nahar, K. (2023). Foliar Application of Ascorbic Acid and Tocopherol in Conferring **Salt Tolerance** in Rapeseed by Enhancing K<sup>+</sup>/Na<sup>+</sup> Homeostasis, Osmoregulation, Antioxidant Defense, and Glyoxalase System. *Agronomy*, 13(2), 361. <https://doi.org/10.3390/agronomy13020361>

- Gul, H. S., Ulfat, M., Zafar, Z. U., Haider, W., Ali, Z., Manzoor, H., ... & Athar, H. U. R. (2022). Photosynthesis and Salt Exclusion Are Key Physiological Processes Contributing to Salt **Tolerance** of Canola (*Brassica napus* L.): Evidence from Physiology and Transcriptome Analysis. *Genes*, 14(1), 3. <https://doi.org/10.3390/genes14010003>
- Raihan, M. R. H., Rahman, M., Mahmud, N. U., Adak, M. K., Islam, T., Fujita, M., & Hasanuzzaman, M. (2022). Application of Rhizobacteria, *Paraburkholderia fungorum* and *Delftia* sp. Confer **Cadmium Tolerance** in Rapeseed (*Brassica campestris*) through Modulating Antioxidant Defense and Glyoxalase Systems. *Plants*, 11(20), 2738. <https://doi.org/10.3390/plants11202738>
- Liao, Y., Tang, Y., Wang, S., Su, H., Chen, J., Zhang, D., ... & Liu, L. (2023). Abscisic acid modulates differential physiological and biochemical responses to **cadmium stress** in *Brassica napus*. *Environmental Pollutants and Bioavailability*, 35(1), 2168216. <https://doi.org/10.1080/26395940.2023.2168216>
- Yu, Y., Dong, J., Li, R., Zhao, X., Zhu, Z., Zhang, F., ... & Lin, X. (2023). Sodium hydrosulfide alleviates **aluminum toxicity** in *Brassica napus* through maintaining H<sub>2</sub>S, ROS homeostasis and enhancing aluminum exclusion. *Science of The Total Environment*, 858, 160073. <https://doi.org/10.1016/j.scitotenv.2022.160073>
- Elahi, N. N., Raza, S., Rizwan, M. S., Albalawi, B. F. A., Ishaq, M. Z., Ahmed, H. M., ... & Ditta, A. (2022). **Foliar Application of Gibberellin** Alleviates Adverse Impacts of **Drought Stress** and Improves Growth, Physiological and Biochemical Attributes of Canola (*Brassica napus* L.). *Sustainability*, 15(1), 78. <https://doi.org/10.3390/su15010078>
- Hemati, A., Alikhani, H.A., Babaei, M. et al. Effects of **foliar application** of **humic acid** extracts and indole acetic acid on important **growth** indices of canola (*Brassica napus* L.). *Sci Rep* 12, 20033 (2022). <https://doi.org/10.1038/s41598-022-21997-5>
- Brown, C., Gulden, R. H., Shirliffe, S. J., & Vail, S. (2022). A Review of the Genetic, Physiological and Agronomic Factors Influencing **Secondary Dormancy** Levels and **Seed Vigour** in *Brassica napus* L. *Canadian Journal of Plant Science*, (ja). <https://doi.org/10.1139/cjps-2022-0155>
- Haj Sghaier, A., Tarnawa, Á., Khaeim, H., Kovács, G. P., Gyuricza, C., & Kende, Z. (2022). The Effects of Temperature and Water on the **Seed Germination** and Seedling Development of Rapeseed (*Brassica napus* L.). *Plants*, 11(21), 2819. <https://doi.org/10.3390/plants11212819>
- Gu, Z., Wang, D., Gong, Q. et al. Metabolomic analysis of rapeseed **priming** with H<sub>2</sub>O<sub>2</sub> in response to germination under chilling stress. *Plant Growth Regul* (2022). <https://doi.org/10.1007/s10725-022-00918-6>
- Mazhar, M. W., Ishtiaq, M., Maqbool, M., & Akram, R. (2022). **Seed priming** with Calcium oxide nanoparticles improves germination, biomass, antioxidant defence and yield traits of canola plants **under drought stress**. *South African Journal of Botany*, 151, 889-899. <https://doi.org/10.1016/j.sajb.2022.11.017>
- Zhao, Y., Gao, L., Gao, Z., Tian, B., Chen, T., Xie, J., ... & Jiang, D. (2022). Exploring a rhizobium to **fix nitrogen in non-leguminous plants** by using a tumor-formation root pathogen. *Phytopathology Research*, 4(1), 48. <https://doi.org/10.1186/s42483-022-00154-w>

- Tatarintsev, N.P., Zakharchenko, N.S., Shmarev, A.N. et al. Effects of **Diphenylurea** on Energy-Storage Reactions of Photosynthesis in Rapeseed Ontogenesis. *Russ. Agricult. Sci.* 48 (Suppl 1), S69–S73 (2022). <https://doi.org/10.3103/S1068367422070205>
- Zhang, X., Fang, T., Huang, Y. et al. Transcriptional regulation of photomorphogenesis in seedlings of *Brassica napus* under different light qualities. *Planta* 256, 77 (2022). <https://doi.org/10.1007/s00425-022-03991-3>
- Tomaszewska-Sowa, M., Lisiecki, K., & Pańska, D. (2022). Response of Rapeseed (*Brassica napus* L.) to Silver and Gold Nanoparticles as a Function of Concentration and Length of Exposure. *Agronomy*, 12(11), 2885. <https://doi.org/10.3390/agronomy12112885>
- Saffari, M.R., Jafarzadeh Kenarsari, M., Farnia, A. et al. Growth Analysis and Oil Quality of Canola (*Brassica napus* L.) Treated with Zinc Nanochelate and Zinc Sulfate Under Different Irrigation Regimes. *Gesunde Pflanzen* (2023). <https://doi.org/10.1007/s10343-022-00825-w>
- Dong, R., Liu, R., Xu, Y., Liu, W., & Sun, Y. (2022). Effect of foliar and root **exposure to** polymethyl methacrylate **microplastics** on biochemistry, ultrastructure, and arsenic accumulation in *Brassica campestris* L. *Environmental Research*, 215, 114402. <https://doi.org/10.1016/j.envres.2022.114402>
- Yang, J., Yu, Y., Ma, C., & Zhang, H. (2023). Direct absorption of atmospheric lead by rapeseed siliques is the leading cause of **seed lead pollution**. *Journal of Hazardous Materials*, 443, 130284. <https://doi.org/10.1016/j.jhazmat.2022.130284>

## REMOTE SENSING

- Liu, F., Wang, F., Wang, X., Liao, G., Zhang, Z., Yang, Y., & Jiao, Y. (2022). Rapeseed **Variety Recognition** Based on Hyperspectral Feature Fusion. *Agronomy*, 12(10), 2350. <https://doi.org/10.3390/agronomy12102350>
- Hu, F., Lin, C., Peng, J., Wang, J., & Zhai, R. (2022). Rapeseed **Leaf Estimation Methods** at Field Scale by Using Terrestrial LiDAR Point Cloud. *Agronomy*, 12(10), 2409. <https://doi.org/10.3390/agronomy12102409>
- TAO, J. B., ZHANG, X. Y., WU, Q. F., & Yun, W. A. N. G. (2022). **Mapping** winter rapeseed in South China using Sentinel-2 data based on a novel separability index. *Journal of Integrative Agriculture*. <https://doi.org/10.1016/j.jia.2022.10.008>
- Liu, W., & Zhang, H. (2023). **Mapping** annual 10 m rapeseed extent using multisource data in the Yangtze River Economic Belt of China (2017–2021) on Google Earth Engine. *International Journal of Applied Earth Observation and Geoinformation*, 117, 103198. <https://doi.org/10.1016/j.jag.2023.103198>
- Fernando, H., Ha, T., Attanayake, A., Benaragama, D., Nketia, K. A., Kanmi-Obembe, O., & Shirtliffe, S. J. (2022). **High-Resolution Flowering Index** for Canola Yield Modelling. *Remote Sensing*, 14(18), 4464. <https://doi.org/10.3390/rs14184464>
- Shirtliffe, S., Johnson, E., & Duddu, H. (2022). **Image-based** remote approach of Canola **yield modelling** with cumulative temporal ground cover for precision agronomy. *Authorea Preprints*. <https://doi.org/10.1002/essoar.10508285.1>

Lawes, R., Mata, G., Richetti, J. et al. Using remote sensing, process-based crop models, and machine learning to **evaluate crop rotations** across 20 million hectares in Western Australia. *Agron. Sustain. Dev.* 42, 120 (2022). <https://doi.org/10.1007/s13593-022-00851-y>

### **PROCESSING, QUALITY & PRODUCTS**

Černilová, B., Kuře, J., Linda, M., & Chotěborský, R. (2022). Tracing of the rapeseed movement by using the contrast point tracking method for DEM model verification. <https://doi.org/10.15159/ar.22.052>

Wang, W., Yang, B., Huang, F., Zheng, C., Li, W., Liu, T., & Liu, C. (2022). **Synchronous pressing and refining** after solid-phase preadsorption technology as a new method for rapeseed oil preparation. *LWT*, 168, 113939. <https://doi.org/10.1016/j.lwt.2022.113939>

Uquiche, E., Sánchez, B., Marillán, C., & Quevedo, R. (2022). Simultaneous **extraction** of lipids and minor lipids from microalga (*Nannochloropsis gaditana*) and rapeseed (*Brassica napus*) using **supercritical carbon dioxide**. *The Journal of Supercritical Fluids*, 190, 105753. <https://doi.org/10.1016/j.supflu.2022.105753>

Wang, S., Wang, J., Dong, G., Chen, X., Wang, S., Feng, L., ... & Bai, Q. (2022). Effect of Different **Extraction** Methods on **Quality** Characteristics of Rapeseed and Flaxseed Oils. *Journal of Food Quality*, 2022. <https://doi.org/10.1155/2022/8296212>

Ning, N. I. N. G., Bing, H. U., BAI, C. Y., LI, X. H., Jie, K. U. A. I., HE, H. Z., ... & ZHAO, S. M. (2023). Influence of **two-stage harvesting** on the properties of **cold-pressed rapeseed** (*Brassica napus* L.) oils. *Journal of Integrative Agriculture*, 22(1), 265-278. <https://doi.org/10.1016/j.jia.2022.09.015>

Sendzikiene, E., Makareviciene, V., & Santaraite, M. (2022). Simultaneous **Extraction** of Rapeseed Oil and **Enzymatic Transesterification** with Butanol in the Mineral Diesel Medium. *Energies*, 15(18), 6837. <https://doi.org/10.3390/en15186837>

Yan, B., Meng, L., Huang, J., Liu, R., Zhang, N., Jiao, X., ... & Fan, D. Evaluation of **Oxidative Stability** of Rapeseed Oils by Microwave Heating: Compared with Conduction Heating. Available at SSRN 4288814. <https://ssrn.com/abstract=4288814> or <http://dx.doi.org/10.2139/ssrn.4288814>

Zhang, Y., Xiao, H., Lv, X., Zheng, C., Wu, Z., Wang, N., ... & Wei, F. (2023). Profiling and spatial distribution of **phenolic compounds** in rapeseed by two-step extraction strategy and targeted metabolomics combined with chemometrics. *Food Chemistry*, 401, 134151. <https://doi.org/10.1016/j.foodchem.2022.134151>

Zhang, Y., Xiao, H., Lv, X., Wang, D., Chen, H., & Wei, F. (2022). Comprehensive review of composition distribution and advances in profiling of **phenolic compounds** in oilseeds. *Frontiers in Nutrition*, 2646. <https://doi.org/10.3389/fnut.2022.1044871>

Fadairo, O. S., Nandasiri, R., Nguyen, T., Eskin, N., Aluko, R. E., & Scanlon, M. G. (2022). Improved Extraction Efficiency and Antioxidant Activity of Defatted Canola Meal Extract **Phenolic Compounds** Obtained from Air-Fried Seeds. *Antioxidants*, 11(12), 2411. <https://doi.org/10.3390/antiox11122411>

Tian, Y., Zhou, Y., Kriisa, M., Anderson, M., Laaksonen, O., Kütt, M. L., ... & Yang, B. (2023). Effects of **fermentation** and enzymatic treatment **on phenolic compounds** and soluble proteins in oil press

- cakes of canola (*Brassica napus*). Food Chemistry, 409, 135339. <https://doi.org/10.1016/j.foodchem.2022.135339>
- Pham, V. D., Korver, D. R., & Gänzle, M. G. (2023). Conversion of **Phenolic Acids** in Canola Fermentation: Impact on Antimicrobial Activity against *Salmonella enterica* and *Campylobacter jejuni*. Journal of Agricultural and Food Chemistry. <https://doi.org/10.1021/acs.jafc.2c08322>
- Tileuberdi, N., Turgumbayeva, A., Yeskaliyeva, B., Sarsenova, L., & Issayeva, R. (2022). Extraction, Isolation of Bioactive **Compounds and Therapeutic** Potential of Rapeseed (*Brassica napus L.*). Molecules, 27(24), 8824. <https://doi.org/10.3390/molecules27248824>
- Fadairo, O.S., Nandasiri, R., Eskin, N.A.M. et al. Air Frying as a Heat Pre-treatment Method for Improving the **Extraction and Yield of Canolol** from Canola Seed Oil. Food Bioprocess Technol 16, 639–651 (2023). <https://doi.org/10.1007/s11947-022-02961-7>
- Nandasiri, R., Fadairo, O., Nguyen, T., Zago, E., Anas, M., & Eskin, N. M. (2022). Optimized Production of **Canolol** Using Microwave Digestion as a Method of Pre-Treatment. <https://doi.org/10.3390/foods12020318>
- Nandasiri, R., Fadairo, O., Nguyen, T., Zago, E., Anas, M. U., & Eskin, N. A. (2023). Optimization of **Canolol Production** from Canola Meal Using Microwave Digestion as a Pre-Treatment Method. Foods, 12(2), 318. <https://doi.org/10.3390/foods12020318>
- Marcinkowski, D., Czwartkowski, K., Bochniak, M., Wereńska, M., & Krzaczek, P. (2022). **Reuse of Bleaching Earth**: The Green Solution for Rapeseed Oil Producers. Sustainability, 14(20), 13071. <https://doi.org/10.3390/su142013071>
- Hou, Z., Jiang, S., Cao, X., Cao, L., Pang, M., Yang, P., & Jiang, S. (2023). Performances of phospholipids and changes of antioxidant capacity from rapeseed oil during **enzymatic degumming**. LWT, 173, 114222. <https://doi.org/10.1016/j.lwt.2022.114222>
- Rashidian, M., Gharachorloo, M., Bahmaei, M., Ghavami, M., & Mirsaeedghazi, H. (2022). Effects of **solvent concentration on refining** (degumming, dewaxing and deacidification) of canola oil using membrane filtration. Iranian Journal of Chemistry and Chemical Engineering. [https://www.ijcce.ac.ir/article\\_699874.html](https://www.ijcce.ac.ir/article_699874.html)
- Lu, T., Guo, Y., Shi, J., Li, X., Wu, K., Li, X., ... & Xiong, Y. (2022). Identification and Safety Evaluation of **Ochratoxin A** Transformation Product in Rapeseed **Oil Refining** Process. Journal of Agricultural and Food Chemistry, 70(47), 14931-14939. <https://doi.org/10.1021/acs.jafc.2c04532>
- Wen, Y. Q., Xue, C. H., Zhang, H. W., Xu, L. L., Wang, X. H., Bi, S. J., ... & Jiang, X. M. (2023). Concomitant **oxidation** of fatty acids other than DHA and EPA plays a role in the characteristic **off-odor of fish oil**. Food Chemistry, 404, 134724. <https://doi.org/10.1016/j.foodchem.2022.134724>
- Rabiej-Kozioł, D., Tymczewska, A., & Szydłowska-Czerniak, A. (2022). Changes in **Quality** of Cold-Pressed Rapeseed Oil with Sinapic Acid Ester-Gelatin Films during **Storage**. Foods, 11(21), 3341. <https://doi.org/10.3390/foods11213341>
- Zhang, L., Chen, J., Zhang, J., Sagymbek, A., Li, Q., Gao, Y., ... & Yu, X. (2022). Lipid oxidation in **fragrant rapeseed oil**: Impact of seed **roasting** on the generation of key volatile compounds. Food Chemistry: X, 16, 100491. <https://doi.org/10.1016/j.fochx.2022.100491>

- Zhang, L., Chen, J., Zhao, X., Wang, Y., & Yu, X. (2022). Influence of **roasting** on the thermal degradation pathway in the **glucosinolates** of fragrant rapeseed oil: Implications to **flavour** profiles. *Food Chemistry*: X, 16, 100503. <https://doi.org/10.1016/j.fochx.2022.100503>
- Tan, M., Zhang, HB., Ye, PP. et al. Distinguishing strong, mellow and light **fragrant rapeseed oils** in China using physicochemical, nutritional and aroma profiles. *Food Measure* (2022). <https://doi.org/10.1007/s11694-022-01729-z>
- Drabińska, N., Siger, A. & Jeleń, H. Comprehensive two-dimensional gas chromatography-time of flight mass spectrometry as a tool for tracking roasting-induced changes in the **volatilome of cold-pressed rapeseed oil**. *Anal Bioanal Chem* (2022). <https://doi.org/10.1007/s00216-022-04486-6>
- Coughlan, R., Kilcawley, K., Skibinska, I., Moane, S., & Larkin, T. (2023). Analysis of **volatile organic** compounds in Irish rapeseed oils. *Current Research in Food Science*, 6, 100417. <https://doi.org/10.1016/j.crfs.2022.100417>
- Liang, Q., Xiong, W., Zhou, Q., Cui, C., Xu, X., Zhao, L., ... & Yao, Y. (2023). Glucosinolates or erucic acid, which one contributes more to volatile **flavor of fragrant rapeseed oil**?. *Food Chemistry*, 135594. <https://doi.org/10.1016/j.foodchem.2023.135594>
- Yang, Z., Huang, Z. & Cao, L. Biotransformation technology and high-value application of **rapeseed meal**: a review. *Bioresour. Bioprocess.* 9, 103 (2022). <https://doi.org/10.1186/s40643-022-00586-4>
- Vlassa, M., Filip, M., Țăranu, I., Marin, D., Untea, A. E., Ropotă, M., ... & Sărăcilă, M. (2022). The **Yeast Fermentation** Effect on Content of Bioactive, Nutritional and Anti-Nutritional Factors in Rapeseed **Meal**. *Foods*, 11(19), 2972. <https://doi.org/10.3390/foods11192972>
- Taranu, I., Pistol, G. C., Anghel, A. C., Marin, D., & Bulgaru, C. (2022). **Yeast-Fermented Rapeseed Meal** Extract Is Able to Reduce Inflammation and Oxidative Stress Caused by *Escherichia coli* Lipopolysaccharides and to Replace ZnO in Caco-2/HTX29 Co-Culture Cells. *International Journal of Molecular Sciences*, 23(19), 11640. <https://doi.org/10.3390/ijms231911640>
- Alpiger, S., & Corredig, M. (2022). Changes in the Physicochemical Properties of Rapeseed-derived **Protein** Complexes During **Enzyme-Assisted Wet Milling**. <https://doi.org/10.22541/au.166786941.10286887/v1>
- Guo, L., Guo, Y., Wu, P., Liu, S., Gu, C., Wu, M., ... & He, R. (2022). Enhancement of **Polypeptide Yield** Derived from Rapeseed Meal with Low-Intensity Alternating **Magnetic Field**. *Foods*, 11(19), 2952. <https://doi.org/10.3390/foods11192952>
- Plankensteiner, L., Yang, J., Bitter, J. H., Vincken, J. P., Hennebelle, M., & Nikiforidis, C. V. (2023). High **yield extraction of oleosins**, the proteins that plants developed to stabilize oil droplets. *Food Hydrocolloids*, 137, 108419. <https://doi.org/10.1016/j.foodhyd.2022.108419>
- Huang, W., Xu, H., Pan, J., Dai, C., Mintah, B. K., Dabbour, M., ... & Ma, H. (2022). Mixed-Strain Fermentation Conditions Screening of Polypeptides from Rapeseed Meal and the Microbial Diversity Analysis by High-Throughput Sequencing. *Foods*, 11(20), 3285. <https://doi.org/10.3390/foods11203285>
- Kaugarenia, N., Beaubier, S., Durand, E., Aymes, A., Villeneuve, P., Lesage, F., & Kapel, R. (2022). Optimization of **selective hydrolysis of cruciferins** for production of potent mineral chelating

- peptides and napins purification to valorize total rapeseed meal **proteins**. *Foods*, 11(17), 2618. <https://doi.org/10.3390/foods11172618>
- Wang, Y., Rosa-Sibakov, N., Edelman, M., Sozer, N., Katina, K., & Coda, R. (2022). Enhancing the utilization of **rapeseed protein ingredients in bread** making by tailored lactic acid fermentation. *Food Bioscience*, 50, 102028. <https://doi.org/10.1016/j.fbio.2022.102028>
- Momen, S., & Aider, M. (2023). Production of highly soluble and functional **whey/canola proteins** through complexation using alkaline electro-activation. *Food Hydrocolloids*, 137, 108395. <https://doi.org/10.1016/j.foodhyd.2022.108395>
- Shen, P., Yang, J., Nikiforidis, C. V., Mocking-Bode, H. C., & Sagis, L. M. (2023). Cruciferin versus napin–Air-water interface and **foam** stabilizing properties of rapeseed storage **proteins**. *Food Hydrocolloids*, 136, 108300. <https://doi.org/10.1016/j.foodhyd.2022.108300>
- Li, C., Shi, D., Stone, A., Wanasundara, J., Tanaka, T., & Nickerson, M. (2022). Select functional **properties of protein isolates** obtained from canola meals modified by solid-state fermentation. *Authorea Preprints*. <https://doi.org/10.22541/au.166576503.35339303/v1>
- Li, C., Shi, D., Stone, A., Wanasundara, J., Tanaka, T., & Nickerson, M. (2022). Effect of solid-state fermentation on select antinutrients and **protein digestibility** of cold-pressed and hexane-extracted canola meals. *Authorea Preprints*. <https://doi.org/10.22541/au.167157004.42104415/v1>
- Schubert, M., Erlenbusch, N., Wittland, S., Nikolay, S., Hetzer, B., & Matthäus, B. (2022). Rapeseed Oil Based **Oleogels** for the Improvement of the Fatty Acid Profile Using Cookies as an Example. *European Journal of Lipid Science and Technology*, 124(11), 2200033. <https://doi.org/10.1002/ejlt.202200033>
- Bagnani, M., Ehrenguber, S., Soon, W. L., Peydayesh, M., Miserez, A., & Mezzenga, R. (2022). Rapeseed Cake Valorization into **Bioplastics Based on Protein Amyloid Fibrils**. *Advanced Materials Technologies*, 2200932. <https://doi.org/10.1002/admt.202200932>
- Dissanayake, T., Trinh, B. M., Mekonnen, T. H., Sarkar, P., Aluko, R. E., & Bandara, N. (2023). Improving properties of **canola protein-based nanocomposite films** by hydrophobically modified nanocrystalline cellulose. *Food Packaging and Shelf Life*, 35, 101018. <https://doi.org/10.1016/j.fpsl.2022.101018>
- Zhang, Q., Ma, Y., Qi, Z., Jia, C., Yao, Y., & Zhang, D. (2022). Optimisation on uniformity and compressibility of rapeseed straw cellulose fiber mixtures for straw/mineral hybrid natural **fiber composite**. *Industrial Crops and Products*, 189, 115852. <https://doi.org/10.1016/j.indcrop.2022.115852>
- Martinez Diaz, J., Grande, P. M., & Klose, H. (2023). Small-scale OrganoCat processing to screen rapeseed **straw** for efficient **fractionation** into cellulose, sugars, and lignin. *Frontiers in Chemical Engineering*, 5, 1. <https://doi.org/10.3389/fceng.2023.1098411>
- Jerman, M., Böhm, M., Dušek, J., & Černý, R. (2022, November). Effect of surface treatment of straw on microstructure and mechanical properties of rapeseed **particleboards**. In *AIP Conference Proceedings* (Vol. 2611, No. 1, p. 040005). AIP Publishing LLC. <https://doi.org/10.1063/5.0119795>

- Jerman, M., Böhm, M., Dušek, J., & Černý, R. (2022, November). Water vapor and thermal properties of newly developed rapeseed **particleboards**. In AIP Conference Proceedings (Vol. 2611, No. 1, p. 040004). AIP Publishing LLC. <https://doi.org/10.1063/5.0119793>
- Tene Tayo, J. L., Bettelhäuser, R. J., & Euring, M. (2022). Canola Meal as Raw Material for the Development of **Bio-Adhesive for Medium Density Fiberboards (MDFs)** and Particleboards Production. *Polymers*, 14(17), 3554. <https://doi.org/10.3390/polym14173554>
- Abookleesh, F., Mosa, F. E., Barakat, K., & Ullah, A. (2022). Assessing Molecular Docking Tools to Guide the Design of **Polymeric Materials Formulations**: A Case Study of Canola and Soybean Protein. *Polymers*, 14(17), 3690. <https://doi.org/10.3390/polym14173690>
- Sousa, D., Salgado, J. M., Cambra-López, M., Dias, A., & Belo, I. (2023). **Bioprocessing of oilseed cakes by fungi** consortia: Impact of enzymes produced on antioxidants release. *Journal of Biotechnology*. <https://doi.org/10.1016/j.jbiotec.2023.01.008>
- Heidari, F., Øverland, M., Hansen, J. Ø., Mydland, L. T., Urriola, P. E., Chen, C., ... & Hu, B. (2022). **Solid-state fermentation** of *Pleurotus ostreatus* to improve the nutritional profile of mechanically-fractionated canola meal. *Biochemical Engineering Journal*, 187, 108591. <https://doi.org/10.1016/j.bej.2022.108591>
- Beaubier, S., Pineda-Vadillo, C., Mesieres, O., Framboisier, X., Galet, O., & Kapel, R. (2023). Improving the **in vitro digestibility** of rapeseed albumins resistant to gastrointestinal proteolysis while preserving the functional properties using enzymatic hydrolysis. *Food Chemistry*, 407, 135132. <https://doi.org/10.1016/j.foodchem.2022.135132>
- Jingting Yao, Ying Hang, Xueming Hua, Ningyu Li, Xiang Li, "Hepatopancreas-Intestinal Health in **Grass Carp** (*Ctenopharyngodon idella*) Fed with Hydrolyzable Tannin or Rapeseed Meal", *Aquaculture Nutrition*, vol. 2022, Article ID 6746201, 14 pages, 2022. <https://doi.org/10.1155/2022/6746201>
- Zhang, B., Liu, N., Hao, M., Xie, Y., & Song, P. (2022). Effects of substitution of soybean meal with **rapeseed meal** and glutamine supplementation on growth performance, intestinal morphology, and intestinal mucosa barrier of Qiandongnan Xiaoxiang **Chicken**. *Animal Bioscience*, 35(11), 1711-1724. <https://doi.org/10.5713/ab.21.0467>
- Wiśniewska, Z., Kołodziejki, P., Pruszyńska, E., Konieczka, P., Kinsner, M., Górka, P., ... & Kaczmarek, S. A. (2023). Effect of emulsifier and multicarbohydase enzyme supplementation on performance and nutrient digestibility in **broiler diets** containing rapeseed meal. *Poultry Science*, 102(1), 102268. <https://doi.org/10.1016/j.psj.2022.102268>
- Li, P., Ji, X., Deng, X., Hu, S., Wang, J., Ding, K., & Liu, N. (2023). Effect of rapeseed meal degraded by enzymolysis and fermentation on the growth performance, nutrient digestibility and health status of **broilers**. *Archives of Animal Nutrition*, 1-12. <https://doi.org/10.1080/1745039X.2022.2162801>
- Hamadi, Sana and Salari, Somayyeh and Aghayi, Ali and Ghorbani, Mohammadreza, Changes in Performance and Apparent **Ileal Digestibility** of **Broiler** Chickens Fed Diets Containing Electron Irradiated Full- Fat Canola Seed. Available at SSRN: <https://ssrn.com/abstract=4252601> or <http://dx.doi.org/10.2139/ssrn.4252601>

- Khalil, M. M., Abdollahi, M. R., Zaefarian, F., Chrystal, P. V., & Ravindran, V. (2023). **Broiler** Age Influences the Apparent Metabolizable Energy of Soybean Meal and Canola Meal. *Animals*, 13(2), 219. <https://doi.org/10.3390/ani13020219>
- Czech, A., Nowakowicz-Debek, B., Łukaszewicz, M. et al. Effect of **fermented rapeseed meal** in the mixture for **growing pigs** on the gastrointestinal tract, antioxidant status, and immune response. *Sci Rep* 12, 15764 (2022). <https://doi.org/10.1038/s41598-022-20227-2>
- Stødkilde, L., Mogensen, L., Bache, J. K., Ambye-Jensen, M., Vinther, J., & Jensen, S. K. (2023). Local protein sources for **growing-finishing pigs** and their effects on pig performance, sensory quality and climate impact of the produced pork. *Livestock Science*, 267, 105128. <https://doi.org/10.1016/j.livsci.2022.105128>
- Wlazło, Ł., Nowakowicz-Dębek, B., Ossowski, M., Łukaszewicz, M., & Czech, A. (2022). Effect of **Fermented Rapeseed Meal** in Diets for **Piglets** on Blood Biochemical Parameters and the Microbial Composition of the Feed and Faeces. *Animals*, 12(21), 2972. <https://doi.org/10.3390/ani12212972>
- Razzaghi, A., Leskinen, H., Ahvenjärvi, S., Aro, H., & Bayat, A. R. (2022). Energy utilization and milk fat responses to rapeseed oil when fed to **lactating dairy cows** receiving different dietary forage to concentrate ratio. *Animal Feed Science and Technology*, 293, 115454. <https://doi.org/10.1016/j.anifeedsci.2022.115454>
- Subrahmanyeswar, G., Murugan, S. S., Juliet, S., Sudharsan, C., & Chacko, B. (2022). EFFECT OF DIETARY SUPPLEMENTATION OF RUMEN PROTECTED RAPESEED OIL ON MILK OMEGA-3 FATTY ACID PROFILE OF **LACTATING CROSSBRED COWS**. <https://agris.fao.org/agris-search/search.do?recordID=IN2022027689>
- Alvarez-Hess, P. S., Jacobs, J. L., Kinley, R. D., Roque, B. M., Neachtain, A. O., Chandra, S., & Williams, S. R. O. (2023). Twice daily feeding of canola oil steeped with *Asparagopsis armata* reduced **methane emissions** of lactating **dairy cows**. *Animal Feed Science and Technology*, 115579. <https://doi.org/10.1016/j.anifeedsci.2023.115579>
- Bernard, L., Chilliard, Y., Hove, K., Volden, H., Inglingstad, R. A., & Eknæs, M. (2022). Feeding of palm oil fatty acids or rapeseed oil throughout lactation: Effects on mammary gene expression and milk production in Norwegian **dairy goats**. *Journal of Dairy Science*, 105(11), 8792-8805. <https://doi.org/10.3168/jds.2021-21372>
- Shahini, E., Luhovyi, S., Kalynychenko, H., Starodubets, O., & Trybrat, R. (2022). Rational use of **oilseed waste** to increase **dairy** productivity. *International Journal of Environmental Studies*, 1-9. <https://doi.org/10.1080/00207233.2022.2147727>
- Paya, H., **Taghizadeh**, A., Hosseinkhani, A., Mohammadzadeh, H., Janmohammadi, H., & Moghaddam, G. (2022). Effects of different **heat processing** methods of rapeseed on **ruminal** and post-ruminal nutrient disappearance. *Journal of the Hellenic Veterinary Medical Society*, 73(3), 4425–4432. <https://doi.org/10.12681/jhvms.27293>
- Chi, Y. P., Haese, E., & Rodehutschord, M. (2023). Ruminal and post-ruminal **phytate degradation** of diets containing rapeseed meal or soybean meal. *Archives of Animal Nutrition*, 1-15. <https://doi.org/10.1080/1745039X.2022.2164158>

- Mierlita, D., Santa, A., Mierlita, S., Daraban, S. V., Suteu, M., Pop, I. M., ... & Macri, A. M. (2022). The Effects of **Feeding Milled Rapeseed Seeds** with Different Forage: Concentrate Ratios in Jersey Dairy Cows on Milk Production, Milk Fatty Acid Composition, and Milk Antioxidant Capacity. *Life*, 13(1), 46. <https://doi.org/10.3390/life13010046>
- Röder, H., Kumar, K., Fuchsl, S., & Sieber, V. (2022). Ex-ante **life cycle assessment** and scale up: A **protein production** case study. *Journal of Cleaner Production*, 376, 134329. <https://doi.org/10.1016/j.jclepro.2022.134329>
- Lascu, I., Tănase, A. M., Jablonski, P., Chiciudean, I., Preda, M. I., Avramescu, S., ... & Stoica, I. (2022). Revealing the Phenotypic and Genomic Background for PHA Production from Rapeseed-Biodiesel Crude Glycerol Using *Photobacterium ganghwense* C2. 2. *International Journal of Molecular Sciences*, 23(22), 13754. <https://doi.org/10.3390/ijms232213754>
- Azargohar, R., Nanda, S., Cheng, H., & Dalai, A. K. (2022). Potential Application of **Canola Hull Fuel Pellets** for the Production of Synthesis Gas and Hydrogen. *Energies*, 15(22), 8613. <https://doi.org/10.3390/en15228613>
- Longwic, R., Sander, P., Zdziennicka, A., Szymczyk, K., & Jańczuk, B. (2023). Changes of Some Physico-chemical Properties of Canola Oil by Adding n-Hexane and Ethanol Regarding Its **Application as Diesel Fuel**. *Applied Sciences*, 13(2), 1108. <https://doi.org/10.3390/app13021108>

## **NUTRITION AND HEALTH**

- Yao, M., Xu, F., Yao, Y., Wang, H., Ju, X., & Wang, L. (2022). Assessment of Novel **Oligopeptides** from Rapeseed Napin (*Brassica napus*) in Protecting HepG2 Cells from Insulin Resistance and Oxidative Stress. *Journal of Agricultural and Food Chemistry*, 70(39), 12418-12429. <https://doi.org/10.1021/acs.jafc.2c03718>
- Ma, K., Wang, Z., Ju, X., Huang, J., & He, R. (2022). **Rapeseed peptide** inhibits HepG2 cell proliferation by regulating the mitochondrial and P53 signaling pathways. *Journal of the Science of Food and Agriculture*. <https://doi.org/10.1002/jsfa.12243>
- Ferrero, R. L., Weinstein-Opppenheimer, C. R., Cabrera-Muñoz, Z., & Zúñiga-Hansen, M. E. (2023). The **Antiproliferative Activity** of a Mixture of **Peptide and Oligosaccharide** Extracts Obtained from Defatted Rapeseed Meal on Breast Cancer Cells and Human Fibroblasts. *Foods*, 12(2), 253. <https://doi.org/10.3390/foods12020253>
- Monié, A., Habersetzer, T., Sureau, L., David, A., Clemens, K., Perez, E., ... & Delample, M. (2023). Modulation of the **crystallization of rapeseed oil** using lipases and the impact on **ice cream** properties. *Food Research International*, 112473. <https://doi.org/10.1016/j.foodres.2023.112473>
- EFSA Panel on Nutrition, Novel Foods and, Food Allergens (NDA), Turck, D., Bohn, T., Castenmiller, J., De Henauw, S., Hirsch-Ernst, K. I., ... & Knutsen, H. K. (2023). Safety of **whole seeds** of oilseed rape (*Brassica napus L emend. Metzg.*) as a **novel food** pursuant to Regulation (EU) 2015/2283. *EFSA Journal*, 21(1), e07706. <https://doi.org/10.2903/j.efsa.2023.7706>
- Guriec, N., Le Foll, C., & Delarue, J. (2023). **Long chain n-3** polyunsaturated fatty acids given before and throughout gestation and lactation in rats prevent high fat-diet-induced **insulin-resistance** in

male offspring in a tissue specific manner. *British Journal of Nutrition*, 1-34.  
<https://doi.org/10.1017/S000711452300017X>

## ANALYZES

- Broothaerts, W., Beaz Hidalgo, R., Buttinger, G., Corbisier, P., Cordeiro, F., Cubria Radio, M., ... & Robouch, P. (2022). **Determination of GM Maize in Bakery Mix (T1) and GM Oilseed Rape in Rapeseed Meal (T2)**. [https://publications.jrc.ec.europa.eu/repository/bitstream/JRC130779/JRC130779\\_01.pdf](https://publications.jrc.ec.europa.eu/repository/bitstream/JRC130779/JRC130779_01.pdf)
- Li, D., Wang, D., Xiao, H., Lv, X., Zheng, C., Liu, C., ... & Wei, F. (2022). Simultaneous Analysis of **Free/Combined Phytosterols in Rapeseed** and Their Dynamic Changes during Microwave Pre-treatment and Oil Processing. *Foods*, 11(20), 3219. <https://doi.org/10.3390/foods11203219>
- Castellaneta, A., Losito, I., Cisternino, G., Leoni, B., Santamaria, P., Calvano, C. D., ... & Cataldi, T. R. (2022). All Ion Fragmentation Analysis Enhances the Untargeted Profiling of **Glucosinolates** in Brassica Microgreens by Liquid Chromatography and High-Resolution Mass Spectrometry. *Journal of the American Society for Mass Spectrometry*, 33(11), 2108-2119. <https://doi.org/10.1021/jasms.2c00208>
- Yu, X., Le Quéré, J. M., Sotin, H., Citeau, M., Dauguet, S., & Guyot, S. (2023). Characterisation and quantification of condensed **tannins** in rapeseed hulls and meals by depolymerization methods. *Journal of Food Composition and Analysis*, 115, 105004. <https://doi.org/10.1016/j.jfca.2022.105004>
- Tsegay, G., Ammare, Y., Tollassa, K., & Shiferaw, L. Development of non-destructive models to predict **oil content and fatty acid composition** of Gomenzer (Ethiopian mustard) using **near-infrared** reflectance spectroscopy. [REFERENCE](#)

## ECONOMY and MARKET

- Chmielewski, Ł. (2022). Tendencies for Usage of Rapeseed Oil and Maize for Biocomponent Production in Poland Between 2015 and 2020 (No. 916-2022-1304, pp. 85-107). <http://dx.doi.org/10.22004/ag.econ.329860>
- Chatellier, V. (2022). World, **European and French trade in oilseeds** [Les échanges mondiaux, européens et français d'oléagineux] (No. hal-03937481). <https://hal.inrae.fr/hal-03937481/document>
- Li, F., Guo, K., & Liao, X. (2023). Risk Assessment of **China Rapeseed Supply Chain and Policy** Suggestions. *International Journal of Environmental Research and Public Health*, 20(1), 465. <https://doi.org/10.3390/ijerph20010465>
- Zhang, W., Qiu, F., Luckert, M. M., Anderson, J., & McPhee, A. (2022). **Potential supplies of fuel-grade canola oil** for low-carbon fuel production in Alberta, Canada: GIS analysis using an improved service-area approach. <https://doi.org/10.21203/rs.3.rs-2011324/v1>
- Shang, Y., Cai, H., & Wei, Y. (2022). The impacts of infectious disease **pandemic on China's edible vegetable oil futures** markets: A long-term perspective. *Economic Research-Ekonomska Istraživanja*, 1-20. <https://doi.org/10.1080/1331677X.2022.2119425>

Xu, X., Zhang, Y. Canola and soybean oil **price forecasts** via neural networks. Adv. in Comp. Int. 2, 32 (2022). <https://doi.org/10.1007/s43674-022-00045-9>

### **MUSTARD and Other Brassicae**

Muhammad, G., Manaf, A., Khalid, A., Sher, A., Lovatt, C. J., Syed, A., ... & Qayyum, A. (2023). Allometric dynamics of *Sinapis alba* under different ecological conditions. Journal of King Saud University-Science, 35(1), 102403. <https://doi.org/10.1016/j.jksus.2022.102403>

Pandit, T. K., Roy, S., & Das, B. (2022). Optimization of Intra-Row Spacing for Yield Enhancement in System of Mustard Intensification (SMI) Techniques. International Journal of Bio-Resource & Stress Management, 13(11). [REFERENCE](#)

Sharma, H.K., V.V. Singh, A. Kumar, H.S. Meena, B.L. Meena, Pankaj Sharma and P.K. Rai. 2022. Genetic study of **terminal heat stress** in indigenous collections of Indian mustard (*Brassica juncea L.*) germplasm. J. Environ. Biol., 43: 161-169. <http://doi.org/10.22438/jeb/43/1/MRN-1887>

Sharma, Pankaj, H.K. Sharma, A.K. Sharma and P.K. Rai. 2023. Agroecology specific production technology of rapeseed-mustard. ICAR-Directorate of Rapeseed Mustard Reseach, Bharatpur 321 303. pp 76. (book)

### **MISCELLANEOUS**

Jiacheng, Y., Xingyu, W., Qingxi, L., Daxin, G., Wenli, X., & Jia, Y. (2022). Mechanical compression characteristics of rapeseed based on continuous damage theory. Biosystems Engineering, 224, 301-312. <https://doi.org/10.1016/j.biosystemseng.2022.10.008>

*Rapeseed in Ancien Egypt?*

Barberis, E., Manfredi, M., Zilberstein, G., Zilberstein, S., & Righetti, P. G. (2022). A shabti of the Egyptian priest Amenmose unveiled. Journal of Cultural Heritage, 58, 122-129. <https://doi.org/10.1016/j.culher.2022.09.021>

## **Upcoming international and national events**

**April 30-May 3, 2023, Denver, Colorado/USA: AOCs Annual Meeting and Expo**

<https://annualmeeting.aocs.org/program-session-topics>



**2023 AOCs Annual Meeting & Expo**

**April 30–May 3, 2023, Colorado Convention Center, Denver, Colorado, USA**

22-23 May 2023, Wageningen, The Netherlands: 3<sup>rd</sup> International Conference on Lipid Droplets & Oleosomes

<https://lipiddropletsoleosomes.org/>

2-5 July 2023, Nantes, France: 15<sup>th</sup> International Congress ISSFAL International Society for the study of Fatty Acids and Lipids

<https://www.issfalcongress.com/>

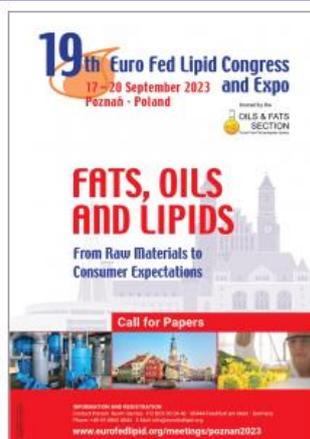


3-7 July 2023, Paris, France: biennial International Society for Seed Science (ISSS) Conference. <https://iss2023.sciencesconf.org/>



17-20 September, 2023, Poznan, Poland: 19<sup>th</sup> Euro Fed Lipid Congress and Expo

[https://veranstaltungen.gdch.de/tms/frontend/index.cfm?l=11215&sp\\_id=2](https://veranstaltungen.gdch.de/tms/frontend/index.cfm?l=11215&sp_id=2)



24-27 September, 2023: 16<sup>th</sup> International Rapeseed Congress, Sydney, Australia  
[www.irc2023sydney.com](http://www.irc2023sydney.com)



***We invite you to share information with the rapeseed/canola community: let us know the scientific projects, events organized in your country, crop performances or any information of interest in rapeseed/canola R&D.***

**Contact GCIRC News:**

Etienne Pilorgé, GCIRC Secretary-Treasurer: [e.pilorge@terresinovia.fr](mailto:e.pilorge@terresinovia.fr)

Contact GCIRC: [contact@gcirc.org](mailto:contact@gcirc.org)